

Research Paper

Expression of programmed death receptor-1 ligand (PD-L1) in human cancer is of prognostic value and associated with macrophage infiltration

Yu Liu^{1,2†}, Qian Li^{3†}, Gangchi Cao^{4†}, Huijuan Wei⁵, Chuansong Xue¹, Jianhao Liu¹✉

1. Sanya Hospital of Traditional Chinese Medicine, Hannan, China.
2. Department of Medical Oncology, Hangzhou Cancer Hospital, Hangzhou, China.
3. Department of Hematology & Oncology, Shanghai Children's Medical Center, Shanghai Jiao Tong University School of Medicine, National Health Committee Key Laboratory of Pediatric Hematology & Oncology, Shanghai, China.
4. Department of Clinical Medicine, Sun Yat-sen University School of Medicine, Guangzhou, China.
5. Guangzhou University of Chinese Medicine, Guangzhou, China.

† These authors have contributed equally to this work.

✉ Corresponding author: Jianhao Liu (Email: Ljhamdoctor08@yeah.net).

© The author(s). This is an open access article distributed under the terms of the Creative Commons Attribution License (<https://creativecommons.org/licenses/by/4.0/>). See <https://ivyspring.com/terms> for full terms and conditions.

Received: 2024.06.18; Accepted: 2024.09.30; Published: 2024.11.04

Abstract

The tumor immune microenvironment is a crucial factor influencing tumor progression, and its molecular mechanisms have become a key topic in immunotherapy research. Programmed death receptor-1 ligand (PD-L1, CD274) is a well-known immunosuppressive molecule that can mediate the immune escape of tumor cells. The aim of this study was to evaluate the significance of PD-L1 in human cancer by integrated bioinformatics analysis. Tumor Immune Estimation Resource (TIMER), GEPIA, Kaplan-Meier plotter, TISIDB and Tumor Immune Single Cell Hub (TISCH) were used to perform the corresponding analysis. The results showed that PD-L1 was dysregulated in various cancers and was associated with the overall survival of cancer patients, which was associated with macrophage infiltration levels. Moreover, PD-L1 expression showed a significant correlation with macrophages and was universally expressed on tumor-associated macrophages (TAMs). Notably, the expression of PD-L1 on TAMs was found to be correlated with immunotherapy response in certain cancers based on analysis of single-cell RNA sequencing data. In conclusion, PD-L1 plays a significant role in cancer, which may partly be influenced by TAMs.

Keywords: PD-L1, macrophage, cancer, immune infiltration, single-cell RNA sequencing

Introduction

Cancer morbidity is rapidly rising worldwide, and cancer is among the three leading causes of death in the 21st century [1]. Although the pathogenesis of cancer has been gradually revealed and multidisciplinary therapies have been developed, the clinical prognosis of cancer patients remains poor [2]. The immune microenvironment around cancer cells plays crucial roles in cancer pathogenesis and is increasingly being investigated [3]. Notably, one of the hallmarks of cancer is evasion of immune destruction [4]. Expression profile data are being continually uploaded to public databases, providing us with the possibility of using bioinformatic

technologies to explore the immune microenvironment in cancer patients [5, 6].

Programmed death receptor-1 ligand (PD-L1, CD274), which serves as a ligand for programmed death receptor-1 (PD-1), is predominantly expressed on the membrane surface of monocytes, macrophages, dendritic cells and various tumor cells [7]. Under normal circumstances, the interaction of PD-L1 and PD-1 protects the body from damage caused by excessive inflammation and plays an important role in the treatment of autoimmune diseases [8]. When tumors occur, PD-1/PD-L1 signal transduction inhibits the function of lymphocytes and

macrophages, and causing apoptosis of lymphocytes, eventually leading to the immune escape of tumor cells. Blocking signal transduction with related anti-PD-1 and/or anti-PD-L1 monoclonal antibodies can restore the immune response, which is a new strategy for tumor immunotherapy, but the clinical effects are quite different in different tumors [9, 10]. Additionally, studies have indicated that PD-L1 serves as a crucial prognostic indicator for patients with tumors [11, 12].

Infiltrating macrophages in the tumor microenvironment are related to poor prognosis and are associated with chemotherapy resistance in most cancers [13]. Macrophages contribute to tumor progression in various ways: by stimulating angiogenesis, nurturing cancer stem cells, increasing migration, and suppressing antitumor immunity [14]. Based on signals received from tumor microenvironment, such as granulocyte-macrophage colony-stimulating factor (GM-CSF), colony-stimulating factor 1 (CSF1), IFN- γ , M0 macrophages polarize into two subtypes of tumor-associated macrophages (TAMs): anti-tumoral M1 and pro-tumoral M2-like TAMs. The phenotype of TAMs has been shown to be strongly correlated with cancer stage and prognosis [10, 15]. Multiple studies have found that a treatment scheme targeting macrophages showed excellent therapeutic effect and certain clinical benefits [16-18]. Therefore, it is particularly important to identify the key molecules that regulate the function of macrophages in the tumor microenvironment.

In this study, we analyzed the expression and prognostic value of PD-L1 in different types of cancer. Subsequently, we explored the association of PD-L1 expression with macrophage infiltration levels and the marker genes of macrophages in some cancers. Additionally, using single-cell RNA sequencing (scRNA-seq) data, we evaluated the expression of PD-L1 on TAMs and its therapeutic implications.

Materials and Methods

TIMER database

The Tumor IMMune Estimation Resource (TIMER) database (<http://timer.cistrome.org/>) was employed to analyze the expression of PD-L1 in cancers and normal tissues and to estimate the correlations between PD-L1 expression and macrophage infiltration levels [19]. Moreover, we explored the outcomes of cancer patients by comprehensively considering macrophage infiltration and PD-L1 expression levels. The expression of PD-L1 was also analyzed with the Gene Expression Profiling Interactive Analysis (GEPIA) database ([\[gepia2.cancer-pku.cn/#index\]\(http://gepia2.cancer-pku.cn/#index\)\) \[20\]. Spearman's correlation analysis to describe the correlation between PD-L1 expression and macrophages. P values less than 0.05 were considered statistically significant.](http://</p></div><div data-bbox=)

Kaplan-Meier plotter

The online Kaplan-Meier plotter database (<http://kmplot.com/analysis/>) and GEPIA database were adopted to explore the prognostic value of PD-L1 in various cancers [21]. Kaplan-Meier plotter includes 21 cancer types, and sources for the databases include GEO, EGA, and TCGA. Kaplan-Meier survival plots were constructed to reveal the association of PD-L1 expression with the overall survival of cancer patients. The log-rank test was used to determine the statistical significance of the correlation. P values less than 0.05 were regarded as statistically significant.

TISIDB

TISIDB (<http://cis.hku.hk/TISIDB>) was employed to estimate the relationship between the abundance of TAMs and the expression of PD-L1 [22]. TISIDB is a user-friendly web portal that has precalculated the associations between genes and immune features for 30 TCGA cancer types. The expression data are first $\log_2(\text{TPM}+1)$ transformed for differential analysis.

scRNA-seq analysis

Tumor Immune Single Cell Hub (TISCH, <http://tisch.comp-genomics.org>) was adopted to explore the expression of PD-L1 on TAMs and the corresponding clinical value. This database includes single-cell transcriptomic profiles from nearly 2 million cells from 76 high-quality tumor datasets across 27 cancer types [23].

Results

Abnormal expression of PD-L1 in cancer

We employed the TIMER database to analyze the expression of PD-L1 in different types of human cancer. PD-L1 levels in cancer tissues and in normal tissues were compared based on a TCGA dataset. As shown in Figure 1A, PD-L1 expression in various cancers was obviously different from that in normal tissues. The expression levels of PD-L1 in the tumor tissues of cervical squamous cell carcinoma (CESC), cholangiocarcinoma (CHOL), esophageal carcinoma (ESCA), head and neck squamous cell carcinoma (HNSC), and stomach adenocarcinoma (STAD) were higher than those in the corresponding control tissues.

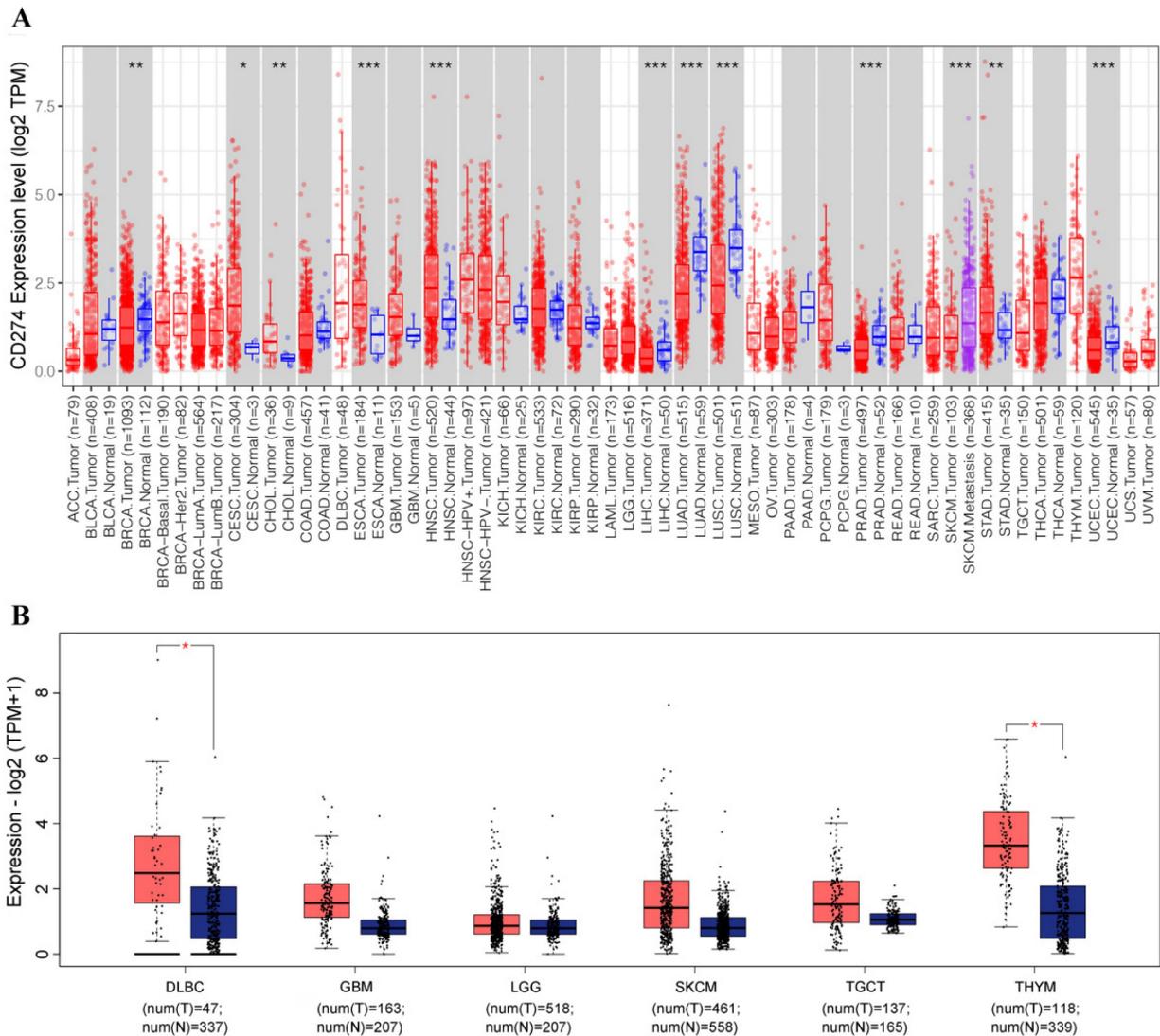


Figure 1. PD-L1 expression was abnormal in various cancers based on TIMER (A) and GEPIA (B) analyses. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, asterisks (*) stand for significance levels.

The expression of PD-L1 was downregulated in breast invasive carcinoma (BRCA), liver hepatocellular carcinoma (LIHC), lung adenocarcinoma (LUAD), lung squamous cell carcinoma (LUSC), prostate adenocarcinoma (PRAD), and uterine corpus endometrial carcinoma (UCEC).

With the GEPIA database, we further evaluated the expression level of PD-L1 in lymphoid neoplasm diffuse large B-cell lymphoma (DLBC), glioblastoma multiforme (GBM), brain lower grade glioma (LGG), skin cutaneous melanoma (SKCM), testicular germ cell tumor (TGCT), and thymoma (THYM). The results showed that the expression of PD-L1 was significantly upregulated in DLBC and THYM (Figure 1B).

The prognostic value of PD-L1 is related to macrophages in various cancers

Subsequently, we constructed Kaplan-Meier

survival curves to determine the prognostic value of PD-L1 in cancer patients. PD-L1 was found to be significantly associated with overall survival in breast cancer, kidney renal clear cell carcinoma, LIHC, ovarian cancer (OV), pancreatic ductal adenocarcinoma (PAAD), sarcoma, TGCT, THYM, UCEC, LGG and SKCM (Figure S1).

We further performed specific analysis based on enriched macrophages. PD-L1 showed significant prognostic value in bladder carcinoma (BLCA, HR=0.37, $p=1.3e-05$), breast carcinoma (HR=0.5, $p=0.0011$), esophageal squamous cell carcinoma (ESCC, HR=0.17, $p=0.01$), OV (HR=0.48, $p=3.2e-05$), PAAD (HR=1.7, $p=0.046$), pheochromocytoma and paraganglioma (PCPG, $p=0.013$), STAD (HR=0.51, $p=0.014$), KIRC (HR=0.61, $p=0.0027$) and UCEC (HR=0.44, $p=0.026$). Notably, the specific analysis based on decreased macrophages revealed that PD-L1 was still associated with the overall survival of KIRC

and UCEC (Figure 2A) but was not associated with overall survival in the other seven types of cancer (Figure 2B). These results indicate that the prognostic value of PD-L1 is related to the content of macrophages in cancers.

PD-L1 expression is correlated with macrophage infiltration levels in cancers

With TISIDB, we assessed the relationship between PD-L1 expression and tumor-infiltrating lymphocyte abundance in cancers. The results showed that PD-L1 was universally associated with

macrophage abundance in 30 cancer types (Figure 3A). We focused on the seven types of tumors mentioned above, and the correlations between PD-L1 expression and macrophages was analyzed with Spearman. The results showed that PD-L1 expression was significantly correlated with macrophage infiltration levels in BLCA ($P < 2.2e-16$), BRCA ($P < 2.2e-16$), ESCA ($P = 1.83e-08$), OA ($P = 1.34e-12$), PAAD ($P = 1.63e-07$) and STAD ($P = 5.56e-15$), except in PCPG ($P = 0.433$) (Figure S2A).

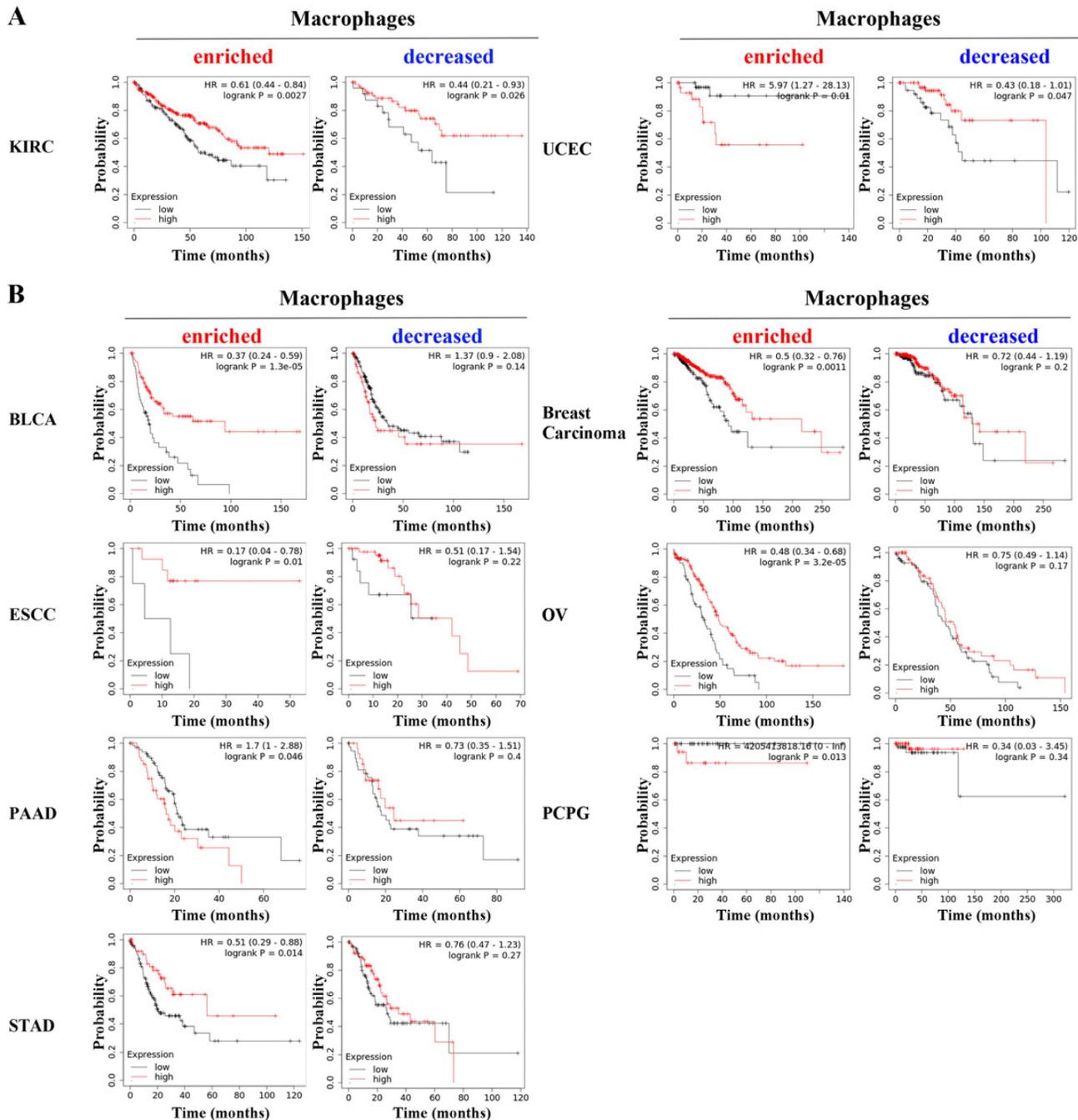


Figure 2. PD-L1 showed statistically significant prognostic value in KIRC and UCEC (A) based on the specific analysis of groups with enriched or decreased macrophage levels, while it did not show prognostic value in other seven types of cancer (B).

In addition, we employed the TIMER database to confirm the correlation between PD-L1 and macrophage infiltration level. As shown in Figure 3B, the PD-L1 expression level had a strong positive correlation with the level of infiltrating macrophages in BLCA ($r = 0.432$, $P = 3.37e-18$), BRCA ($r = 0.394$, $P = 2.60e-38$), ESCA ($r = 0.285$, $P = 1.07e-04$), OV ($r = 0.49$, $P = 2.07e-16$), PAAD ($r = 0.444$, $P = 1.15e-09$), PCPG ($r = 0.193$, $P = 1.23e-02$) and STAD ($r = 0.507$, $P = 3.63e-26$). Notably, the PD-L1 expression level also influenced the prognostic value of macrophages. In the high PD-L1 expression group, but not in the low PD-L1 expression group of BLCA (HR = 1.56, $P = 0.0327$) and STAD (HR = 1.77, $P = 0.0117$), macrophages were revealed to be negatively associated with overall survival. Additionally, the expression of PD-L1 and macrophage infiltration levels showed comprehensive prognostic value in PAAD (HR = 0.305, $P = 0.00103$) (Figure 3C).

To validate the relationship between PD-L1 and macrophages, we further determined the correlation between PD-L1 and the marker genes of macrophages with the TIMER database. Specifically, we showed that the macrophage markers CD68 (BLCA: $P = 5.42e-42$, BRCA: $P = 3.57e-104$, ESCA: $P = 2.49e-03$, OV: $P = 2.69e-30$, PAAD: $P = 3.28e-14$, PCPG: $P = 1.27e-06$, STAD: $P = 1.27e-19$), S100A9 (BLCA: $P = 5.91e-08$, BRCA: $P = 3.02e-20$, ESCA: $P = 5.48e-04$, OV: $P = 4.16e-02$, PAAD: $P = 4.91e-06$, PCPG: $P = 2.26e-02$, STAD: $P = 6.01e-08$) and CSF1R (BLCA: $P = 4.68e-35$, BRCA: $P = 5.68e-79$, ESCA: $P = 2.27e-07$, OV: $P = 2.15e-19$, PAAD: $P = 4.1e-20$, PCPG: $P = 1.85e-04$, STAD: $P = 2.09e-28$) were significantly associated with PD-L1 expression in the above seven types of cancer (Figure S2B). These findings suggest that there is a close relationship between PD-L1 and macrophages in tumors.

PD-L1 is universally expressed on TAMs based on scRNA-seq data

To further explore the expression level of PD-L1 on macrophages, we analyzed the scRNA-seq data of different cancers with TISCH. The results showed that PD-L1 was widely expressed on macrophages in various cancers (Figure S3). We then mainly focused on the above seven types of cancers, as shown in Figure 4A. PD-L1 was found to be expressed on macrophage clusters based on the scRNA-seq data of BRCA, OV and PAAD. Specifically, in the BRCA-GSE110686 dataset, the expression of PD-L1 on the macrophage cluster was higher than that on other cell clusters. The distribution of PD-L1 gene expression in different cell types across the OV-GSE118828 dataset is also shown in Figure 4B.

PD-L1 expression on TAMs correlates with treatment response in cancers

The TISCH dataset included limited treatment information. Thus, we also analyzed the correlation between PD-L1 expression on TAMs and the therapeutic effect in cancer patients. The results showed that the expression of PD-L1 on TAMs was significantly associated with the treatment response in SKCM and basal cell carcinoma (BCC) (Figure 5A and Figure S4A). Clustering of cells based on the SKCM-GSE120575 dataset created a detailed map comprising 18 transcriptionally homogeneous subpopulations (Figure 5B). After adjusting for cell type (major lineage) based on marker gene expression (Figure 5C and D), the expression of PD-L1 on macrophages was significantly higher than that on other cells (Figure 5E). Based on the BCC-GSE123813 dataset, PD-L1 was found to be mainly expressed on macrophages (Figure S4B and C). These results indicated that PD-L1 was highly expressed on macrophages and that its expression was associated with immunotherapy response.

Discussion

With the in-depth study of the tumor immune microenvironment, it has been recognized that the immune escape of tumor cells is the key reason for tumor progression, and its molecular mechanism has also become one of the key topics in tumor immunotherapy research [24]. Recent studies have shown that PD-L1 (CD274) and PD-1 are closely related to immune evasion and tumorigenesis [25]. PD-L1 is highly expressed in a variety of tumor cells and affects the clinical outcomes of immunotherapy [26]. In this study, we found that PD-L1 was upregulated to different degrees in CESC, CHOL, ESCA, HNSC, STAD, DLBC and THYM and downregulated in BRCA, LIHC, LUAD, LUSC, PRAD and UCEC. Based on these results, PD-L1 may be a good potential diagnostic biomarker for different cancers.

Studies have shown that the expression level of PD-L1 is closely related to the prognosis of tumor patients [27], which was also observed in this study. Notably, our results demonstrated that the prognostic value of PD-L1 was different between groups with enriched and decreased macrophage levels in BLCA, BRCA, ESCA, OV, PAAD, PCPG and STAD, indicating that PD-L1 may influence tumorigenesis partly through macrophages. It is well recognized that macrophages are important immune components in the tumor microenvironment [28]. In our study, we further validated the significant correlations of PD-L1 with macrophage infiltration and with the marker

genes of macrophages in the above seven types of cancers. Consistent with these findings, studies have revealed that macrophages can increase the expression of PD-L1 on tumor cells in ESCA and STAD [29, 30]. Moreover, breast cancer cells upregulate PD-L1 expression on macrophages and promote macrophage-associated immunosuppression [31]. Interestingly, circulating sPD-L1 in the blood was found to be positively associated with PD-L1+ macrophages in the blood of OV patients [32].

Based on the scRNA-seq data in TISCH, we further found that PD-L1 was widely expressed on macrophages in cancers. It has been confirmed that TAMs with high expression of PD-L1 play an immunosuppressive role in the tumor microenvironment [33]. In hepatocellular carcinoma, the infiltration of PD-L1+ macrophages was negatively correlated with overall survival and disease-free survival [34]. In non-small-cell lung cancer (NSCLC), the 5-year overall survival of patients with low expression of PD-L1 in macrophages was significantly higher than

that of patients with high expression of PD-L1 in macrophages [35]. Notably, a previous study indicated that macrophage polarization affects the antitumor efficacy of checkpoint inhibitor therapy [36]. In this study, we found that the expression of PD-L1 on macrophages was significantly associated with the checkpoint inhibitor treatment response in SKCM and BCC. Due to the limited treatment information in the single-cell sequencing database, we did not perform a similar analysis in the other tumor types, but such analyses will be performed in future research.

In conclusion, our study found that PD-L1 is dysregulated in various cancers and is associated with prognosis; in addition, the relationship of PD-L1 with prognosis is partly influenced by macrophage infiltration levels. The correlation between PD-L1 and macrophages is widely observed in tumors. Moreover, PD-L1 is universally expressed on macrophages and significantly associated with the treatment response in some cancers.

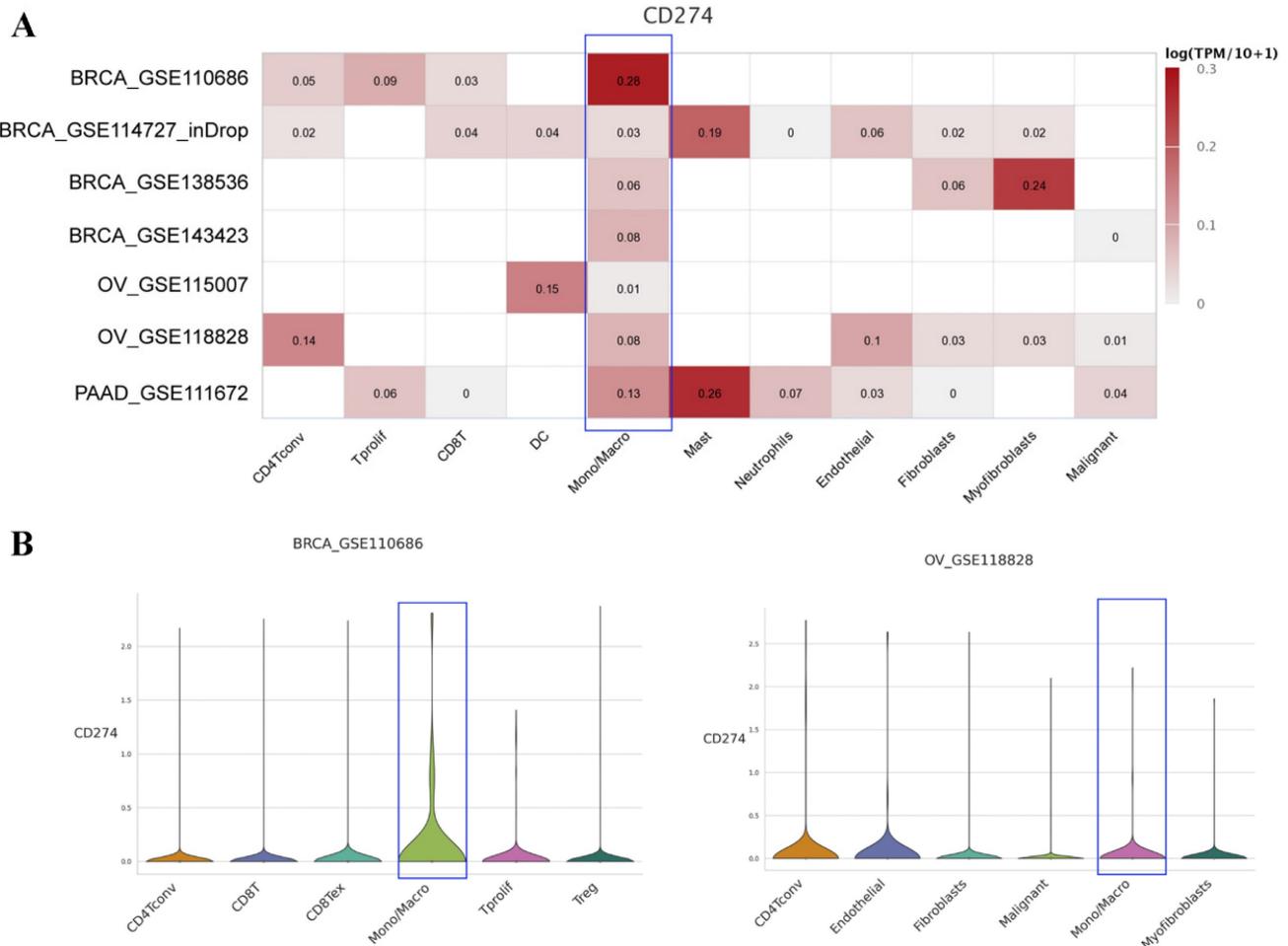


Figure 4. Average expression of PD-L1 in different cell types across some human cancer datasets (heatmap) based on the TISCH web portal (A). Distribution of PD-L1 expression in different cell types across the BRCA-GSE110686 dataset (B) and OV-GSE118828 dataset (C) in the form of violin plots.

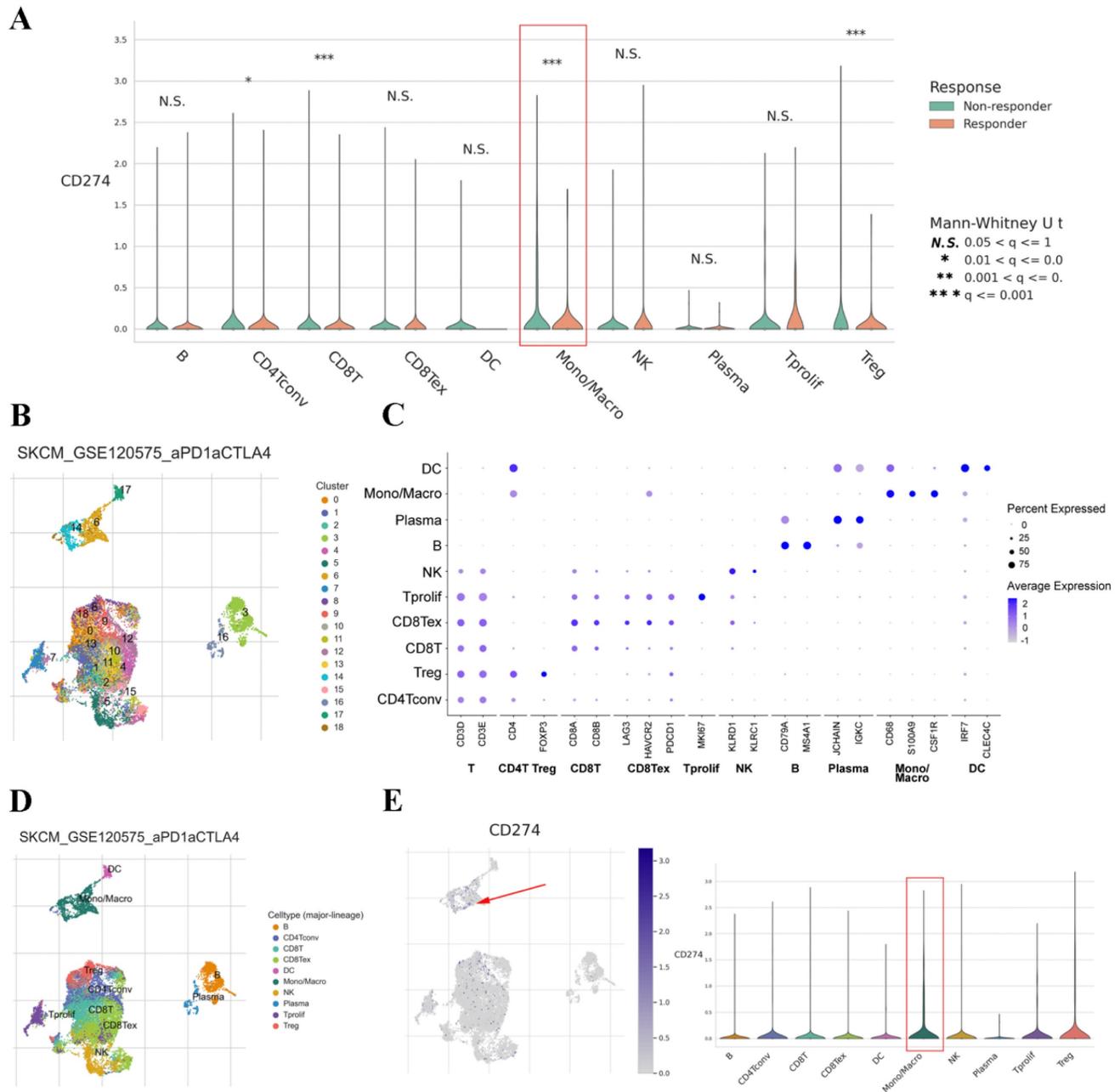


Figure 5. PD-L1 gene expression in different cell types grouped by treatment response based on the SKCM-GSE120575 dataset (A). tSNE plot depicting single cells (B). Marker gene expression according to cell type (major lineage) (C). Subjects are color coded according to the cell type (major lineage) on the tSNE (D). Distribution of PD-L1 expression in different cell types (E).

Abbreviations

PD-L1: programmed death receptor-1 ligand; PD-1: programmed death receptor-1; GM-CSF: granulocyte-macrophage colony-stimulating factor; CSF1: colony-stimulating factor 1; TIMER: Tumor Immune Estimation Resource; TISCH: Tumor Immune Single Cell Hub; TAMs: tumor-associated macrophages; scRNA-seq: single-cell RNA sequencing; GEPIA: Gene Expression Profiling Interactive Analysis; CESC: cervical squamous cell carcinoma; CHOL: cholangiocarcinoma; ESCA:

esophageal carcinoma; HNSC: head and neck squamous cell carcinoma; STAD: stomach adenocarcinoma; BRCA: breast invasive carcinoma; LIHC: liver hepatocellular carcinoma; LUAD: lung adenocarcinoma; LUSC: lung squamous cell carcinoma; PRAD: prostate adenocarcinoma; UCEC: uterine corpus endometrial carcinoma; DLBC: diffuse large B-cell lymphoma; GBM: glioblastoma multiforme; LGG: brain lower grade glioma; SKCM: skin cutaneous melanoma; TGCT: testicular germ cell tumor; THYM: thymoma; OV: ovarian cancer; PAAD: pancreatic ductal adenocarcinoma; BLCA: bladder

carcinoma; ESCC: esophageal squamous cell carcinoma; PCPG: pheochromocytoma and paraganglioma; BCC: basal cell carcinoma; NSCLC: non-small-cell lung cancer.

Supplementary Material

Supplementary figures.

<https://www.jcancer.org/v15p6798s1.pdf>

Acknowledgements

We thank the TIMER, GEPIA, Kaplan-Meier plotter, TISIDB and TISCH for the availability of the data.

Funding

This work was supported by grants from the National Natural Science Foundation of China (Grant number 81770217), Zhejiang Provincial Natural Science Foundation (Grant number LY19H080004) and Hainan Province Clinical Medical Center.

Ethical approval and ethical standards

The data used in this research were obtained from public databases, therefore, ethical approval was not required.

Author contributions

LJH and LY designed the study. WHJ, CGC, and XCS obtained the data and performed data analysis. LY and LQ wrote the document. LY revised the manuscript.

Competing Interests

The authors have declared that no competing interest exists.

References

- Bray F, Laversanne M, Sung H, Ferlay J, Siegel RL, Soerjomataram I, et al. Global cancer statistics 2022: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin*. 2024; 74: 229-63.
- Tompkins KD, Thorburn A. Regulation of Apoptosis by Autophagy to Enhance Cancer Therapy. *Yale J Biol Med*. 2019; 92: 707-18.
- Polak R, Zhang ET, Kuo CJ. Cancer organoids 2.0: modelling the complexity of the tumour immune microenvironment. *Nat Rev Cancer*. 2024; 24(8):523-539.
- Hanahan D. Hallmarks of Cancer: New Dimensions. *Cancer Discov*. 2022; 12: 31-46.
- Wei Z, Guo X, Li D, Wang J, Lin C, Tan C, et al. Prognostic value of CMTM6 protein in hepatocellular carcinoma involving the regulation of the immune microenvironment. *Int J Biol Macromol*. 2024; 275: 133618.
- Yan H, Qu J, Cao W, Liu Y, Zheng G, Zhang E, et al. Identification of prognostic genes in the acute myeloid leukemia immune microenvironment based on TCGA data analysis. *Cancer immunology, immunotherapy* : CII. 2019; 68: 1971-8.
- Lin X, Kang K, Chen P, Zeng Z, Li G, Xiong W, et al. Regulatory mechanisms of PD-1/PD-L1 in cancers. *Mol Cancer*. 2024; 23: 108.
- Sun C, Mezzadra R, Schumacher TN. Regulation and Function of the PD-L1 Checkpoint. *Immunity*. 2018; 48: 434-52.
- Qiu MZ, Oh DY, Kato K, Arkenau T, Tabernero J, Correa MC, et al. Tislelizumab plus chemotherapy versus placebo plus chemotherapy as first line treatment for advanced gastric or gastro-oesophageal junction

adenocarcinoma: RATIONALE-305 randomised, double blind, phase 3 trial. *BMJ*. 2024; 385: e078876.

- Toledo B, Zhu Chen L, Paniagua-Sancho M, Marchal JA, Peran M, Giovannetti E. Deciphering the performance of macrophages in tumour microenvironment: a call for precision immunotherapy. *J Hematol Oncol*. 2024; 17: 44.
- Hu X, Deng X, Xie J, Tang H, Zou Y. Heterogeneous PD-L1 expression in metastases impacts immunotherapy response. *EBioMedicine*. 2023; 97: 104816.
- Zhang Y, Wu J, Zhao C, Zhang S, Zhu J. Recent Advancement of PD-L1 Detection Technologies and Clinical Applications in the Era of Precision Cancer Therapy. *J Cancer*. 2023; 14: 850-73.
- Mantovani A, Marchesi F, Malesci A, Laghi L, Allavena P. Tumour-associated macrophages as treatment targets in oncology. *Nature reviews Clinical oncology*. 2017; 14(7): 399-416
- Komohara Y, Fujiwara Y, Ohnishi K, Takeya M. Tumor-associated macrophages: Potential therapeutic targets for anti-cancer therapy. *Advanced drug delivery reviews*. 2016; 99: 180-5.
- Ding Y, Wang Y, Hu Q. Recent advances in overcoming barriers to cell-based delivery systems for cancer immunotherapy. *Exploration (Beijing)*. 2022; 2: 20210106.
- Cassetta L, Pollard JW. Targeting macrophages: therapeutic approaches in cancer. *Nat Rev Drug Discov*. 2018; 17: 887-904.
- Zheng Y, Han Y, Sun Q, Li Z. Harnessing anti-tumor and tumor-tropism functions of macrophages via nanotechnology for tumor immunotherapy. *Exploration (Beijing)*. 2022; 2: 20210166.
- Li X, Wang H, Chen Y, Li Z, Liu S, Guan W, et al. Novel emerging nano-assisted anti-cancer strategies based on the STING pathway. *Acta Materia Medica*. 2023; 2: 323-341.
- Li T, Fan J, Wang B, Traugh N, Chen Q, Liu JS, et al. TIMER: A Web Server for Comprehensive Analysis of Tumor-Infiltrating Immune Cells. *Cancer research*. 2017; 77: e108-e10.
- Tang Z, Li C, Kang B, Gao G, Li C, Zhang Z. GEPIA: a web server for cancer and normal gene expression profiling and interactive analyses. *Nucleic Acids Res*. 2017; 45: W98-W102.
- Pan JH, Zhou H, Cooper L, Huang JL, Zhu SB, Zhao XX, et al. LAYN Is a Prognostic Biomarker and Correlated With Immune Infiltrates in Gastric and Colon Cancers. *Frontiers in immunology*. 2019; 10: 6.
- Ru B, Wong CN, Tong Y, Zhong JY, Zhong SSW, Wu WC, et al. TISIDB: an integrated repository portal for tumor-immune system interactions. *Bioinformatics*. 2019; 35: 4200-2.
- Sun D, Wang J, Han Y, Dong X, Ge J, Zheng R, et al. TISCH: a comprehensive web resource enabling interactive single-cell transcriptome visualization of tumor microenvironment. *Nucleic Acids Res*. 2021; 49: D1420-D1430.
- Vinay DS, Ryan EP, Pawelec G, Talib WH, Stagg J, Elkord E, et al. Immune evasion in cancer: Mechanistic basis and therapeutic strategies. *Semin Cancer Biol*. 2015; 35 Suppl: S185-S198.
- Gong J, Chehrizi-Raffle A, Reddi S, Salgia R. Development of PD-1 and PD-L1 inhibitors as a form of cancer immunotherapy: a comprehensive review of registration trials and future considerations. *J Immunother Cancer*. 2018; 6: 8.
- Patel SP, Kurzrock R. PD-L1 Expression as a Predictive Biomarker in Cancer Immunotherapy. *Mol Cancer Ther*. 2015; 14: 847-56.
- Akinleye A, Rasool Z. Immune checkpoint inhibitors of PD-L1 as cancer therapeutics. *J Hematol Oncol*. 2019; 12: 92.
- Bronte V, Murray PJ. Understanding local macrophage phenotypes in disease: modulating macrophage function to treat cancer. *Nature medicine*. 2015; 21: 117-9.
- Ju X, Zhang H, Zhou Z, Chen M, Wang Q. Tumor-associated macrophages induce PD-L1 expression in gastric cancer cells through IL-6 and TNF- α signaling. *Exp Cell Res*. 2020; 396: 112315.
- Yagi T, Baba Y, Okadome K, Kiyozumi Y, Hiyoshi Y, Ishimoto T, et al. Tumor-associated macrophages are associated with poor prognosis and programmed death ligand 1 expression in oesophageal cancer. *Eur J Cancer*. 2019; 111: 38-49.
- Jing W, Guo X, Wang G, Bi Y, Han L, Zhu Q, et al. Breast cancer cells promote CD169(+) macrophage-associated immunosuppression through JAK2-mediated PD-L1 upregulation on macrophages. *Int Immunopharmacol*. 2020; 78: 106012.
- Okla K, Rajtak A, Czerwonka A, Bobiński M, Wawruszak A, Tarkowski R, et al. Accumulation of blood-circulating PD-L1-expressing M-MDSCs and monocytes/macrophages in pretreatment ovarian cancer patients is associated with soluble PD-L1. *J Transl Med*. 2020; 18: 220.
- Su S, Zhao J, Xing Y, Zhang X, Liu J, Ouyang Q, et al. Immune Checkpoint Inhibition Overcomes ADCP-Induced Immunosuppression by Macrophages. *Cell*. 2018; 175: 442-57.e23.
- Kuang DM, Zhao Q, Peng C, Xu J, Zhang JP, Wu C, et al. Activated monocytes in peritumoral stroma of hepatocellular carcinoma foster

- immune privilege and disease progression through PD-L1. *J Exp Med.* 2009; 206: 1327-37.
35. Sepesi B, Cuentas EP, Canales JR, Behrens C, Correa AM, Vaporciyan A, et al. Programmed Death Cell Ligand 1 (PD-L1) Is Associated With Survival in Stage I Non-Small Cell Lung Cancer. *Semin Thorac Cardiovasc Surg.* 2017; 29: 408-15.
 36. Choo YW, Kang M, Kim HY. M1 Macrophage-Derived Nanovesicles Potentiate the Anticancer Efficacy of Immune Checkpoint Inhibitors. *ACS Nano.* 2018; 12: 8977-93.