

SUPPLEMENTARY INFORMATION

Supplementary Methods:

Immunoprecipitation of EGFR and HER3 following HER2 pull down in SKBr3, BT474 and BT474 Clone 5

The three cell lines were grown to 80% confluency and treated with trastuzumab or diluent control plus or minus streptavidin following the crosslinking method outlined in the main paper. After a 15 min streptavidin treatment the cells were incubated with 100 μ L lysis buffer (1% Triton X100, 20 mM Tris-HCl, 150 mM NaCl, pH 7.5, and miniComplete protease inhibitors). Cell lysate protein content was assessed using BCA assay after being centrifuged at 13,000 $\times g$ for 10 min at 4 °C and the supernatant collected. Immunoprecipitation was performed using published protocol (58) adapted and outlined below.

Generation of IP beads: anti-HER2 mAb 7C2 (BSA/glycerol free) was purchased from Insight Biotechnology (Wembley, UK) and diluted to 400 μ g/mL in 0.4 M Sodium Citrate, 0.1M Sodium HEPES, pH7.5. All 100 μ g 7C2 was added to 5 mg Ultralink Biosupport (Fisher Scientific) and allowed to react for a minimum of 3 hr at room temperature under constant rotation. 7C2 beads were pelleted at 1,200 $\times g$ for 5 min and the supernatant removed. To both 7C2 beads and 5 mg control beads 10X the bead swell volume (10xSV) of 2M glycine in PBS was added and the solution allowed to react for 3 hr at room temperature under rotation. 7C2 and control beads were washed by pelleting (1,200 $\times g$, 5 min) the beads and resuspended in 10xSV PBS for 15 min at room temperature under rotation. This was followed by further washes in 10xSV 1M NaCl, and two 10xSV PBS washes. The beads were finally resuspended in 200 μ L PBS and kept at 4°C for a maximum of 24 hr.

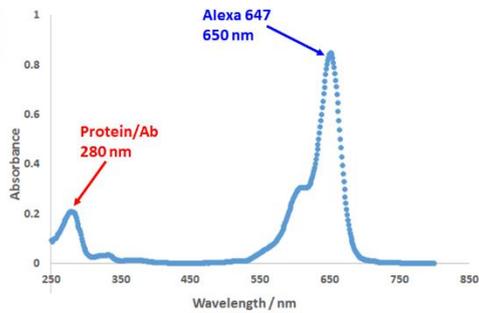
Immunoprecipitation using Ultralink Biosupport beads: 100 μ g of cell lysate was incubated with 15 μ L of 7C2 or control beads in a final volume of 200 μ L lysis buffer overnight at 4 °C under constant rotation. The beads were pelleted at 1,200 $\times g$ for 5 min and the pellet was washed twice with 500 μ L lysis buffer. The final pellet was resuspended in 15 μ L PBS prior to adding SDS loading buffer containing DTT (5 μ L). The sample was boiled for 4 min at 96 °C before being separated on a 7.5% SDS-PAGE gel, transferred to PDVF membranes and probed for EGFR, HER3 and finally HER2 as per the method section.

Supplementary Figures:

A i

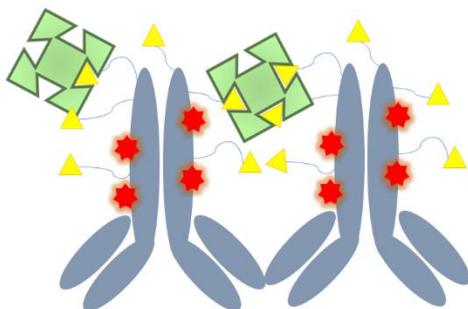
| | |
|--|---------|
| Dilution of stock before analysis | 1:2 |
| Mean Δ absorbance of diluted sample | 0.0845 |
| Biotin concentration | |
| From Beer's law | |
| absorbance = extinction coefficient(34,000) x path length (0.5) x concentration. | 4.971 |
| Biotin conc. = $\frac{0.0845}{0.5 \times 34000} \times 1000000$ | |
| Biotin concentration of stock (μ M) (above x3 for d.f. and x10 for dilution in well plate) | 149.118 |
| Protein concentration (μ M) | 25 |
| Mean number of biotins per protein | 5.965 |

ii



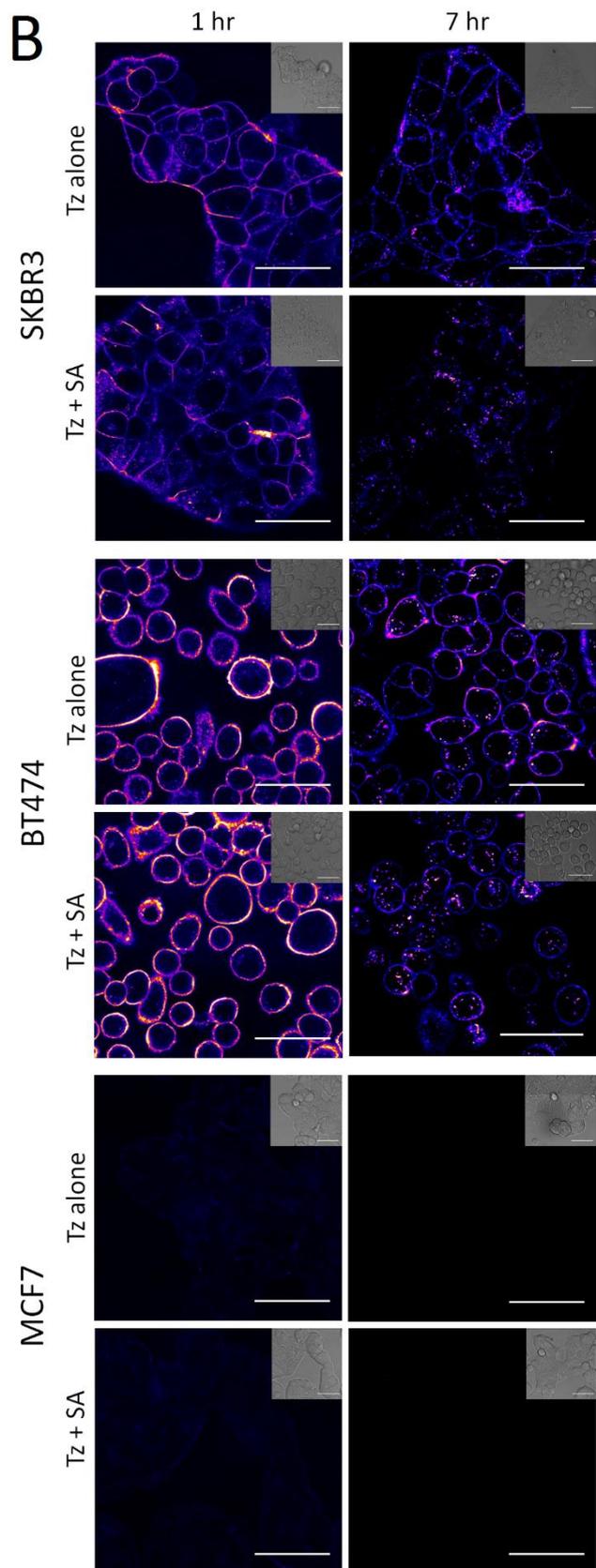
| Dilution factor | TRz-Bi-647 | |
|---|------------------|--------------------|
| | Protein (280 nm) | Alexa 647 (650 nm) |
| | 1 in 100 | |
| Extinction coefficient (M^{-1}) | 200,000 | 239,000 |
| Peak height of diluted sample | 0.207 | 0.847 |
| Corrected peak height, (corrected for dilution i.e. x100) | 20.680 | 84.726 |
| Peak heights, corrected for fluorophore absorbance at 280 nm = Corrected peak height - (Alexa 647 peak height at 650 nm x 0.03) = | 18.139 | 84.726 |
| Concentration of stock (μ M) | 90.694 | 354.503 |
| Mean number of dyes per protein | 3.909 | |

iii



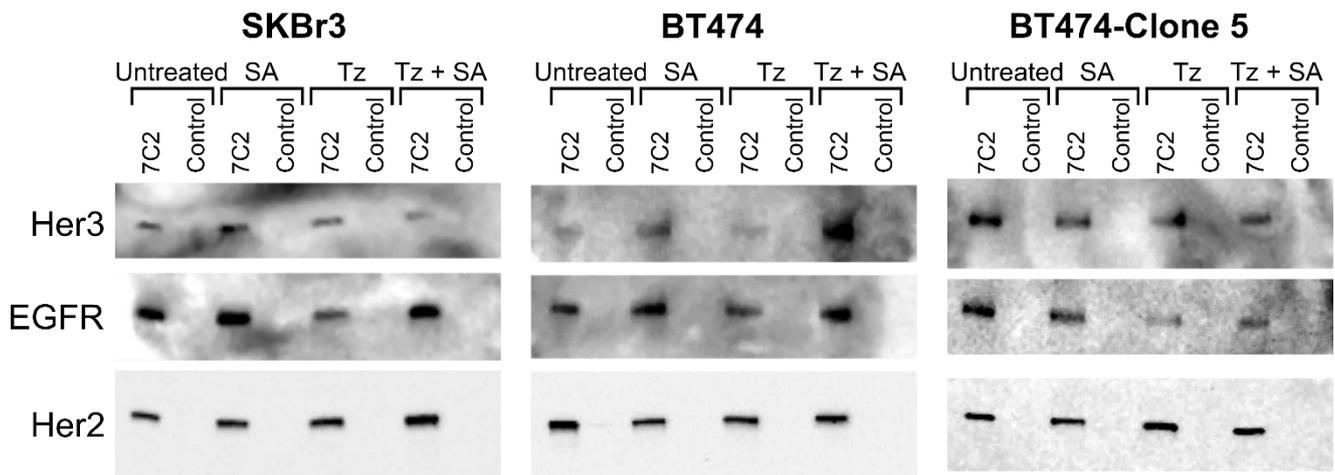
Tz-bi-647: HER2 targeted fluorescently labelled, biotinylated (crosslinkable with streptavidin)

B



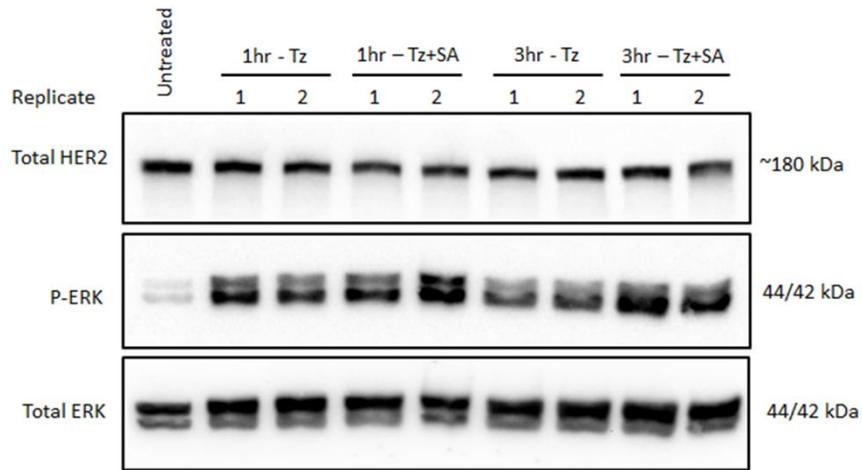
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SUPPLEMENTARY FIGURE S1: Characterisation of Tz-bi-647. A) chemical Tz-bi-647 characterisation: (i) results and analysis of biotinylation assay demonstrating an average of 6.0 biotin moieties per antibody. (ii) UV spectral analysis showing an average of 3.9 fluorophores per antibody. (iii) schematic illustrating how biotinylated Tz can be crosslinked by SA. B) Tz-bi-647 cellular uptake and crosslinking-induced endocytosis characterisation. SKBR3, BT474 and MDA-MB-231 cells were incubated with Tz for 30 min then incubated for 1 hr with either diluent control (left column) or SA (right column). Following treatments the cells were chased in complete imaging medium for 6 hr and imaged. *Scale bar = 50 μm.* **Data demonstrate that crosslinking enhances Tz internalisation in HER2⁺ SKBR3 and BT474 cells and not in HER2⁻ MDA-MB-231 cells where relatively little Tz bound and was endocytosed.**

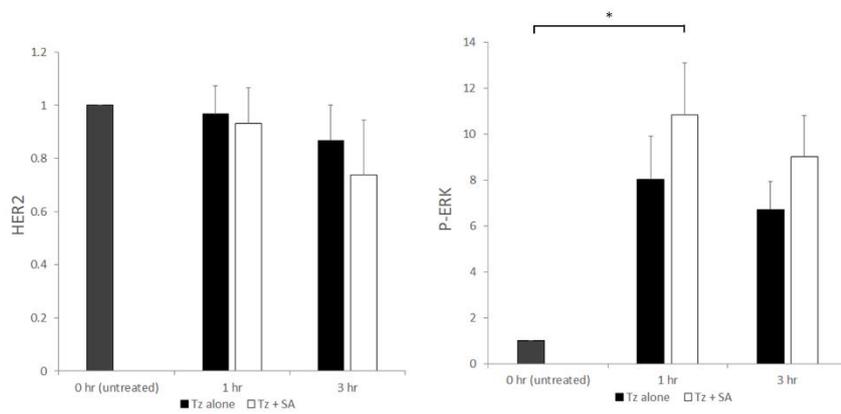


SUPPLEMENTARY FIGURE S2: HER3 and EGFR co-precipitates with HER2 following immunoprecipitation in SKBR3, BT474 and BT474 clone 5. Cells were incubated with trastuzumab or diluent control for 30 min followed by SA (or diluent control) for 15 min under tissue culture conditions. Cells were lysed and HER2 was precipitated through an overnight incubation of 100 μg of lysate with beads linked to an anti-HER2 Ab. HER3, EGFR and HER2 levels were assessed by western blot. **The data show that both EGFR and HER3 a co-precipitated following immunoprecipitation of HER2.**

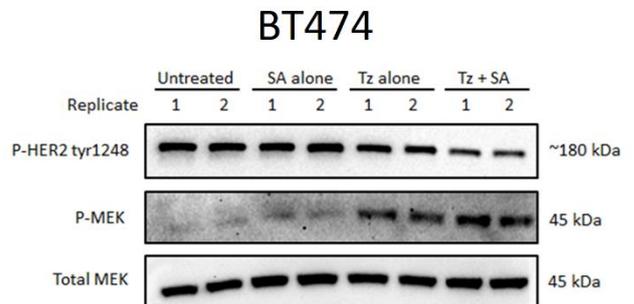
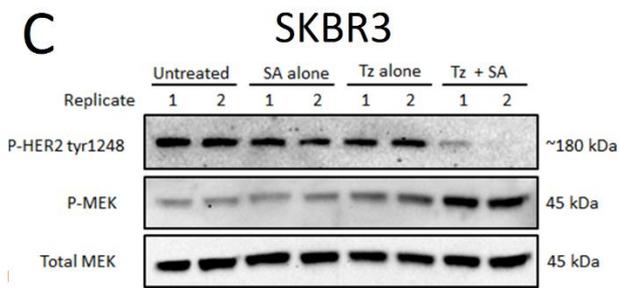
A



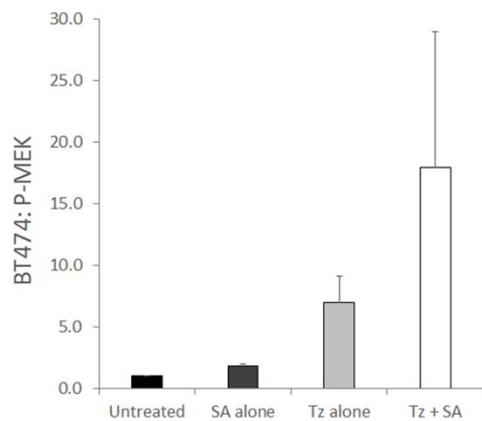
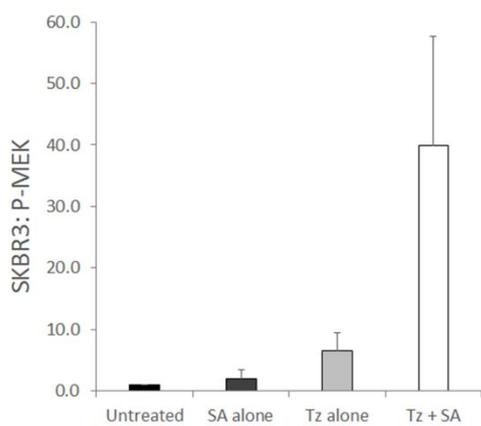
B



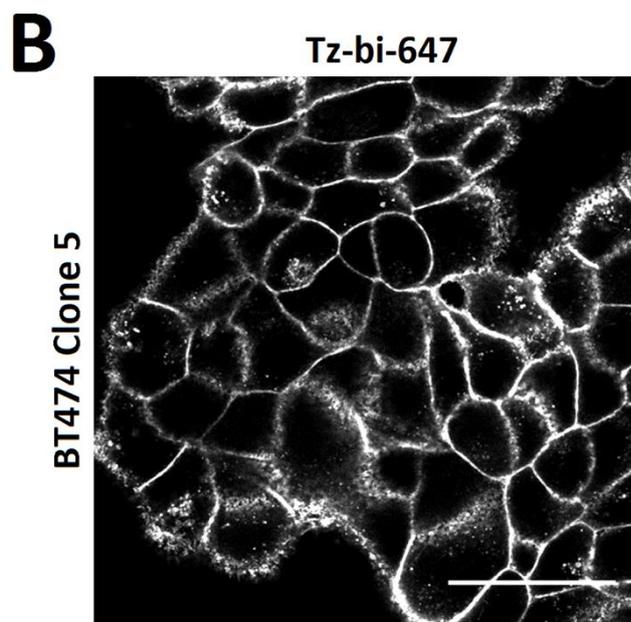
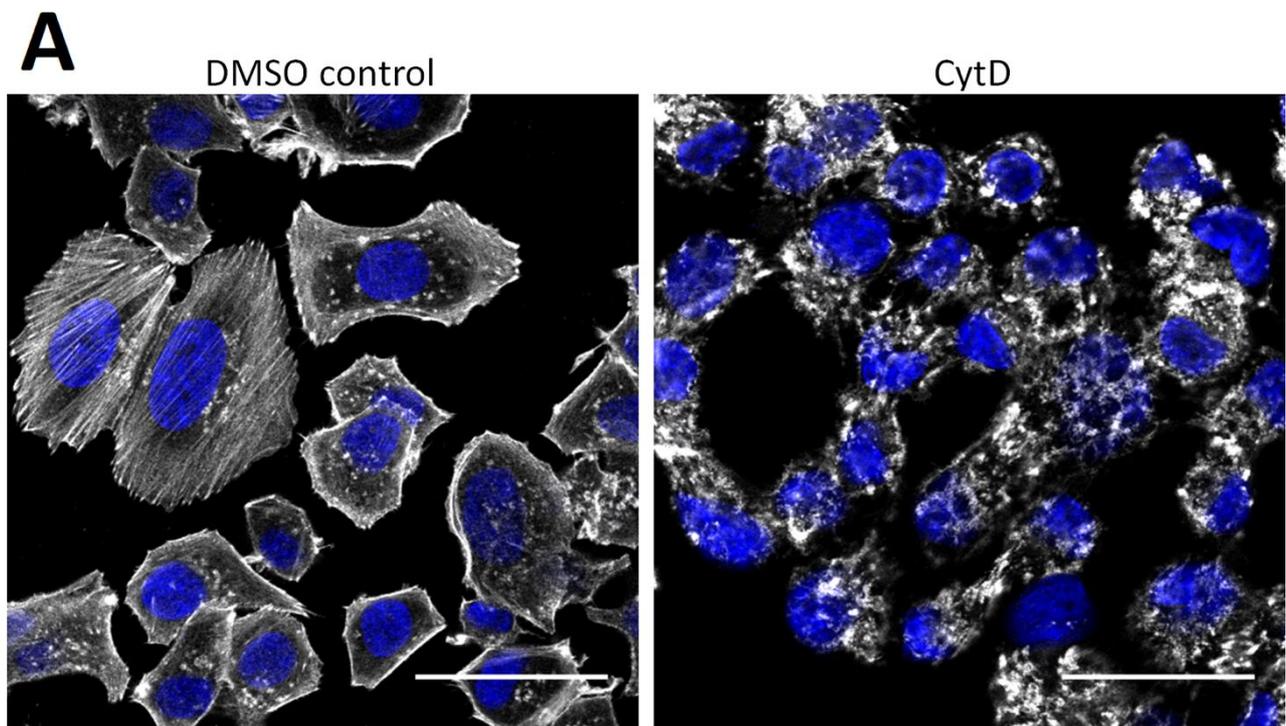
C



D



SUPPLEMENTARY FIGURE S3: Early-time point profile of ERK-activation and HER2 levels in crosslinking treated breast cancer cells and evidence of MEK activation following crosslinking in breast cancer cells. A) SKBR3 cells were either untreated (control) or incubated with Tz diluent control 30 min followed by SA alone for 1 hr, Tz alone for 30 min (+ 1 hr SA diluent control), or Tz for 30 min followed by SA for 1 hr (Tz+SA). For the 3 hr time point cells were subject to a 2 hr chase in CIM before lysate collection. Cell lysates were collected from three independent experiments, representative blots are shown. Western blotting was performed for total HER2, P-ERK (Thr202/Tyr204) and total ERK. B) Band intensities were quantified using ImageJ software. *Mean from 3 independent experiments is shown, error bars represent SE, * $p \leq 0.05$.* **Data show ERK activation and slight reduction in HER2 following crosslinking challenge over 3 hr time course, ERK was also activated by HER2 but there was evidence of this beginning to subside.** C) SKBR3 cells and BT474 cells were either untreated (control) or incubated with Tz diluent control 30 min followed by SA alone for 1 hr, Tz alone for 30 min (+ 1 hr SA diluent control), or Tz for 30 min followed by SA for 1 hr (Tz+SA) then a 6 hr chase in CIM. Cell lysates were collected from three independent experiments, representative blots are shown. Western blotting was performed for P-HER2 Tyr1248, P-MEK (Ser217/221) and total MEK. D) Band intensities were quantified using ImageJ software and mean from 3 independent experiments is shown (*Error bars represent SEM*). **The data demonstrate that crosslinking induced MEK activation of varying magnitude.**



SUPPLEMENTARY FIGURE S4A: CytD disrupts the actin cytoskeleton of cells under the conditions of the crosslinking experiment. SKBR3 cells were incubated with 5 μ M CytD or diluent control for 7.5 hr. Following the treatments cells were fixed in 3% PFA and stained with rhodamine-phalloidin (2 μ g/mL for 15 min), nuclei were counterstained with Hoechst. *Scale bar = 50 μ m*. **Data demonstrate disruption of filamentous actin in the CytD treated cells.** **4B: Tz-b-647 binds to Tz-resistant BT474 cells.** BT474 clone 5 cells were incubated with Tz for 30 min then imaged by confocal microscopy. *Scale bar = 50 μ m*. **Data demonstrate that the Tz-resistant cell line was still able to bind Tz-bi-647 at the plasma membrane.**

A

| HER3: SKBR3 cells | |
|-------------------------------------|---------|
| F-Statistic | 43.4814 |
| Degrees of freedom - Between groups | 3 |
| Degrees of freedom - Within groups | 8 |
| P-Value | 0.0000 |
| Tukey Post-Hoc Test | |
| Untreated vs SA alone | 0.0630 |
| Untreated vs Tz alone | 0.0047 |
| Untreated vs Tz+SA | 0.0000 |
| SA alone vs Tz alone | 0.2803 |
| SA alone vs Tz+SA | 0.0001 |
| Tz alone vs Tz+SA | 0.0014 |

| EGFR: SKBR3 cells | |
|-------------------------------------|--------|
| F-Statistic | 1.195 |
| Degrees of freedom - Between groups | 3 |
| Degrees of freedom - Within groups | 8 |
| P-Value | 0.3716 |

| HER3: BT474 cells | |
|-------------------------------------|--------|
| F-Statistic | 6.8356 |
| Degrees of freedom - Between groups | 3 |
| Degrees of freedom - Within groups | 8 |
| P-Value | 0.0134 |
| Tukey Post-Hoc Test | |
| Untreated vs SA alone | 0.9988 |
| Untreated vs Tz alone | 0.6201 |
| Untreated vs Tz+SA | 0.0204 |
| SA alone vs Tz alone | 0.5442 |
| SA alone vs Tz+SA | 0.0169 |
| Tz alone vs Tz+SA | 0.1154 |

| EGFR: BT474 cells | |
|-------------------------------------|--------|
| F-Statistic | 0.6132 |
| Degrees of freedom - Between groups | 3 |
| Degrees of freedom - Within groups | 8 |
| P-Value | 0.6253 |

B

| HER2: SKBR3 cells | |
|-------------------------------------|---------|
| F-Statistic | 45.9643 |
| Degrees of freedom - Between groups | 3 |
| Degrees of freedom - Within groups | 8 |
| P-Value | 0.0000 |
| Tukey Post-Hoc Test | |
| Untreated vs SA alone | 0.0606 |
| Untreated vs Tz alone | 0.0003 |
| Untreated vs Tz+SA | 0.0000 |
| SA alone vs Tz alone | 0.0092 |
| SA alone vs Tz+SA | 0.0002 |
| Tz alone vs Tz+SA | 0.0409 |

| P-HER2 Tyr1248: SKBR3 cells | |
|-------------------------------------|---------|
| F-Statistic | 11.8297 |
| Degrees of freedom - Between groups | 3 |
| Degrees of freedom - Within groups | 8 |
| P-Value | 0.0955 |
| Tukey Post-Hoc Test | |
| Untreated vs SA alone | 0.5393 |
| Untreated vs Tz alone | 0.9863 |
| Untreated vs Tz+SA | 0.0115 |
| SA alone vs Tz alone | 0.3734 |
| SA alone vs Tz+SA | 0.0021 |
| Tz alone vs Tz+SA | 0.0179 |

| P-HER2 Tyr 877: SKBR3 cells | |
|-------------------------------------|--------|
| F-Statistic | 0.6342 |
| Degrees of freedom - Between groups | 3 |
| Degrees of freedom - Within groups | 8 |
| P-Value | 0.6136 |

| P-ERK: SKBR3 cells | |
|-------------------------------------|---------|
| F-Statistic | 61.3086 |
| Degrees of freedom - Between groups | 3 |
| Degrees of freedom - Within groups | 8 |
| P-Value | 0 |
| Tukey Post-Hoc Test | |
| Untreated vs SA alone | 1 |
| Untreated vs Tz alone | 0.5085 |
| Untreated vs Tz+SA | 0 |
| SA alone vs Tz alone | 0.5261 |
| SA alone vs Tz+SA | 0 |
| Tz alone vs Tz+SA | 0 |

| HER2: BT474 cells | |
|-------------------------------------|--------|
| F-Statistic | 8.0401 |
| Degrees of freedom - Between groups | 3 |
| Degrees of freedom - Within groups | 8 |
| P-Value | 0.0085 |
| Tukey Post-Hoc Test | |
| Untreated vs SA alone | 0.8318 |
| Untreated vs Tz alone | 0.6653 |
| Untreated vs Tz+SA | 0.0230 |
| SA alone vs Tz alone | 0.2617 |
| SA alone vs Tz+SA | 0.0075 |
| Tz alone vs Tz+SA | 0.1168 |

| P-HER2 Tyr1248: BT474 cells | |
|-------------------------------------|--------|
| F-Statistic | 0.966 |
| Degrees of freedom - Between groups | 3 |
| Degrees of freedom - Within groups | 8 |
| P-Value | 0.4547 |

| P-HER2 Tyr 877: BT474 cells | |
|-------------------------------------|--------|
| F-Statistic | 0.3554 |
| Degrees of freedom - Between groups | 3 |
| Degrees of freedom - Within groups | 8 |
| P-Value | 0.7868 |

| P-ERK: BT474 cells | |
|-------------------------------------|---------|
| F-Statistic | 15.1871 |
| Degrees of freedom - Between groups | 3 |
| Degrees of freedom - Within groups | 8 |
| P-Value | 0.0011 |
| Tukey Post-Hoc Test | |
| Untreated vs SA alone | 1 |
| Untreated vs Tz alone | 0.6789 |
| Untreated vs Tz+SA | 0.0019 |
| SA alone vs Tz alone | 0.6566 |
| SA alone vs Tz+SA | 0.0018 |
| Tz alone vs Tz+SA | 0.0073 |

C

| HER2 | |
|--------------------------------------|--------|
| F-Statistic | 3.5874 |
| Degrees of freedom - Between groups | 3 |
| Degrees of freedom - Within groups | 8 |
| P-Value | 0.0324 |
| Tukey Post-Hoc Test | |
| DMSO - untreated vs DMSO - Tz | 0.3967 |
| DMSO - untreated vs DMSO - Tz+SA | 0.0471 |
| DMSO - untreated vs CytD - untreated | 0.9907 |
| DMSO - untreated vs CytD - Tz | 0.3985 |
| DMSO - untreated vs CytD - Tz+SA | 0.0799 |
| DMSO - Tz vs DMSO - Tz+SA | 0.7286 |
| DMSO - Tz vs CytD - untreated | 0.7164 |
| DMSO - Tz vs CytD - Tz | 0.4068 |
| DMSO - Tz vs CytD - Tz+SA | 0.8807 |
| DMSO - Tz+SA vs CytD - untreated | 0.1219 |
| DMSO - Tz+SA vs CytD - Tz | 0.7266 |
| DMSO - Tz+SA vs CytD - Tz+SA | 0.9994 |
| CytD - untreated vs CytD - Tz | 0.7184 |
| CytD - untreated vs CytD - Tz+SA | 0.1992 |
| CytD - Tz vs CytD - Tz+SA | 0.8793 |

D

| HER2 | |
|--------------------------------------|--------|
| F-value | 7.9526 |
| Degrees of freedom - Between groups | 3 |
| Degrees of freedom - Within groups | 8 |
| P-Value | 0.0016 |
| Tukey Post-Hoc Test | |
| DMSO - Untreated vs DMSO - Tz alone | 0.0877 |
| DMSO - Untreated vs DMSO - Tz+SA | 0.0006 |
| DMSO - Untreated vs CytD - Untreated | 0.1141 |
| DMSO - Untreated vs CytD - Tz alone | 0.1231 |
| DMSO - Untreated vs CytD Tz+SA | 0.5569 |
| DMSO - Tz alone vs DMSO - Tz+SA | 0.0813 |
| DMSO - Tz alone vs CytD - Untreated | 1.0000 |
| DMSO - Tz alone vs CytD - Tz alone | 0.9999 |
| DMSO - Tz+SA vs CytD - Tz alone | 0.7692 |
| DMSO - Tz+SA vs CytD - Untreated | 0.0621 |
| DMSO - Tz+SA vs CytD - Tz alone | 0.0574 |
| DMSO - Tz+SA vs CytD Tz+SA | 0.0089 |
| CytD - Untreated vs CytD - Tz alone | 1.0000 |
| CytD - Untreated vs CytD Tz+SA | 0.8477 |
| CytD - Tz alone vs CytD Tz+SA | 0.8680 |

| P-ERK | |
|--------------------------------------|--------|
| F-value | 9.8263 |
| Degrees of freedom - Between groups | 3 |
| Degrees of freedom - Within groups | 8 |
| P-Value | 0.0006 |
| Tukey Post-Hoc Test | |
| DMSO - Untreated vs DMSO - Tz alone | 0.4094 |
| DMSO - Untreated vs DMSO - Tz+SA | 0.0014 |
| DMSO - Untreated vs CytD - Untreated | 0.9681 |
| DMSO - Untreated vs CytD - Tz alone | 0.9813 |
| DMSO - Untreated vs CytD Tz+SA | 0.4447 |
| DMSO - Tz alone vs DMSO - Tz+SA | 0.0366 |
| DMSO - Tz alone vs CytD - Untreated | 1.0000 |
| DMSO - Tz alone vs CytD - Tz alone | 0.7826 |
| DMSO - Tz+SA vs CytD Tz+SA | 1.0000 |
| DMSO - Tz+SA vs CytD - Untreated | 0.0005 |
| DMSO - Tz+SA vs CytD - Tz alone | 0.0042 |
| DMSO - Tz+SA vs CytD Tz+SA | 0.0326 |
| CytD - Untreated vs CytD - Tz alone | 0.7021 |
| CytD - Untreated vs CytD Tz+SA | 0.1531 |
| CytD - Tz alone vs CytD Tz+SA | 0.8168 |

E

| HER2 | |
|-------------------------------------|--------|
| F-Statistic | 1.2366 |
| Degrees of freedom - Between groups | 3 |
| Degrees of freedom - Within groups | 8 |
| P-Value | 0.3585 |

| P-ERK | |
|-------------------------------------|----------|
| F-Statistic | 114.1662 |
| Degrees of freedom - Between groups | 3 |
| Degrees of freedom - Within groups | 8 |
| P-Value | 0.0000 |
| Tukey Post-Hoc Test | |
| Untreated vs SA alone | 0.3341 |
| Untreated vs Tz alone | 0.0000 |
| Untreated vs Tz+SA | 0.0000 |
| SA alone vs Tz alone | 0.0000 |
| SA alone vs Tz+SA | 0.0000 |
| Tz alone vs Tz+SA | 0.5912 |

Fi

| HER2 | |
|-------------------------------------|--------|
| F-Statistic | 0.5923 |
| Degrees of freedom - Between groups | 3 |
| Degrees of freedom - Within groups | 8 |
| P-Value | 0.6761 |

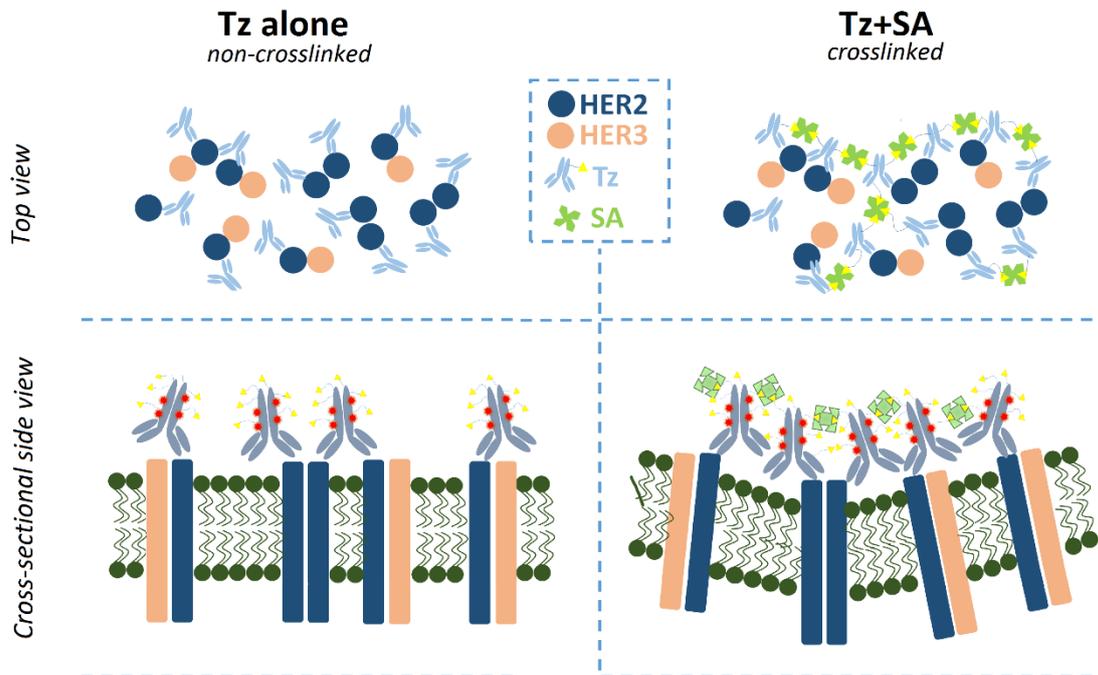
| P-ERK | |
|-------------------------------------|--------|
| F-Statistic | 4.6177 |
| Degrees of freedom - Between groups | 3 |
| Degrees of freedom - Within groups | 8 |
| P-Value | 0.0227 |
| Tukey Post-Hoc Test | |
| Untreated vs 1 hr Tz alone | 0.0999 |
| Untreated vs 1 hr Tz+SA | 0.0171 |
| Untreated vs 3 hr Tz alone | 0.2696 |
| Untreated vs 3 hr Tz+SA | 0.0535 |
| 1 hr Tz alone vs 1 hr Tz+SA | 0.7813 |
| 1 hr Tz alone vs 3 hr Tz alone | 0.9554 |
| 1 hr Tz alone vs 3 hr Tz+SA | 0.9934 |
| 1 hr Tz+SA vs 3 hr Tz alone | 0.4101 |
| 1 hr Tz+SA vs 3 hr Tz+SA | 0.9425 |
| 3 hr Tz alone vs 3 hr Tz+SA | 0.8076 |

ii

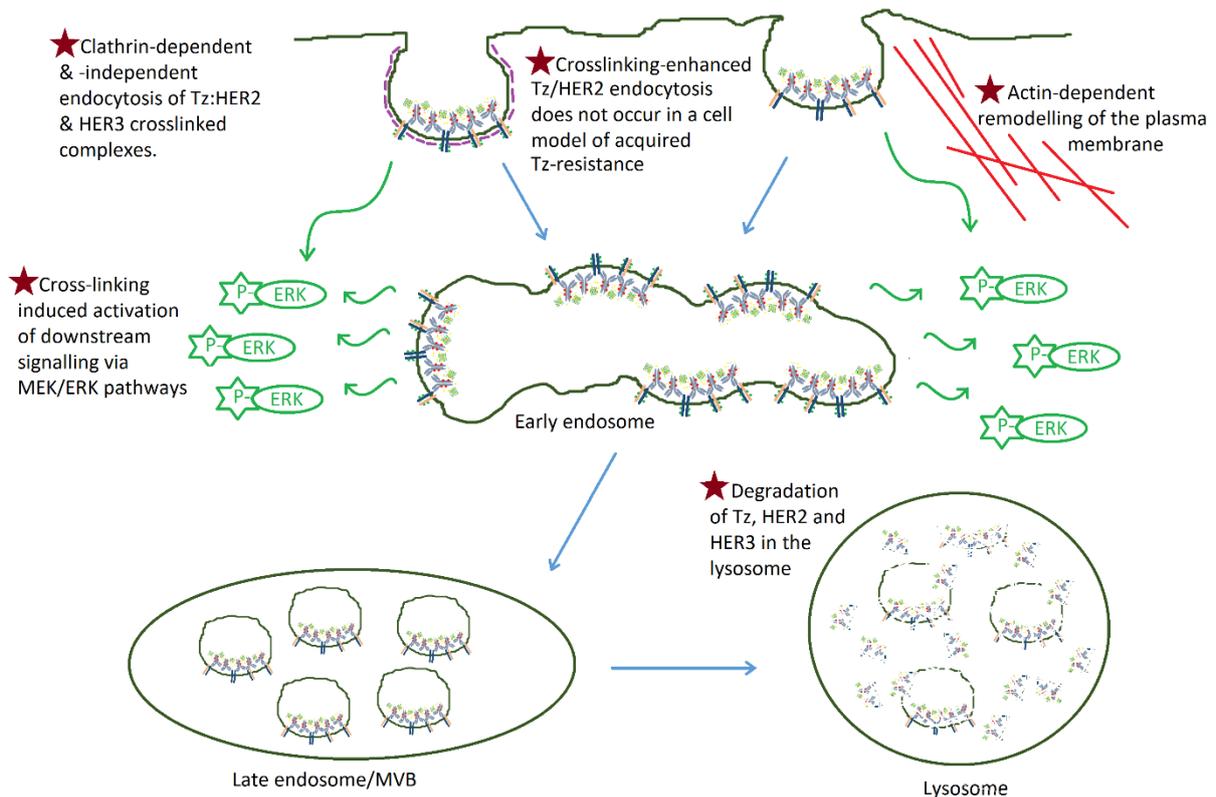
| P-MEK: SKBR3 cells | |
|-------------------------------------|--------|
| F-Statistic | 4.1975 |
| Degrees of freedom - Between groups | 3 |
| Degrees of freedom - Within groups | 8 |
| P-Value | 0.0465 |
| Tukey Post-Hoc Test | |
| Untreated vs SA alone | 0.9998 |
| Untreated vs Tz alone | 0.972 |
| Untreated vs Tz+SA | 0.0627 |
| SA alone vs Tz alone | 0.984 |
| SA alone vs Tz+SA | 0.0699 |
| Tz alone vs Tz+SA | 0.1147 |

| P-MEK: BT474 cells | |
|-------------------------------------|--------|
| F-Statistic | 1.9415 |
| Degrees of freedom - Between groups | 3 |
| Degrees of freedom - Within groups | 8 |
| P-Value | 0.2015 |

SUPPLEMENTARY FIGURE S6 Full ANOVA reporting for Western blotting quantification. A) From Figure 1. B) From Figure 2. C) From Figure 3. D) From Figure 6. E) From Figure 7. F) i) From Supplementary Figure S2B, ii) Supplementary Figure S2D).



Graphical abstract: Crosslinking-induced HER2/3 endocytosis in Tz-sensitive breast cancer cells



SUPPLEMENTARY FIGURE S7: Graphical hypothesis of Tz:HER2 mediated crosslinking at the plasma membrane. Top view and cross-sectional side view of Tz bound to HER2 at the plasma membrane with and without addition of SA crosslinks. Top row: Tz alone is largely only able to form higher-order crosslinks between HER2 homodimers, HER2:HER3 heterodimers terminate the chain of linkages. Middle row: multivalent SA-crosslinking creates extensive HER2 homodimer and HER2:HER3 heterodimer crosslinkages at the plasma membrane inducing curvature and endocytosis. Bottom section: Tz-crosslinking induces endocytosis, lysosomal trafficking and downregulation of HER2 and HERR3 in Tz-sensitive cells.

Supplementary Table 1: Primary antibodies and dilutions. All antibodies are validated and used according to manufacturer's guidelines or published protocols.

| Antibody | Supplier & product number | RRID | Dilution |
|----------------------------------|--------------------------------------|-------------|-----------------|
| Mouse—Tubulin HRP | Abcam, ab-21058 | AB_727045 | 1:50,000 |
| Rabbit—HER3 | Cell Signalling, #12708 | AB_2721919 | 1:1000 |
| Rabbit—EGFR | Cell Signalling, #2232 | AB_10692644 | 1:1000 |
| Rabbit—HER2 | Cell Signalling, #2242 | AB_823466 | 1:1000 |
| Rabbit—P-HER2 tyr-877 | Cell Signalling, #2241 | AB_2099407 | 1:500 |
| Rabbit—P-HER2 tyr-1248 | Cell Signalling, #2247 | AB_331725 | 1:500 |
| Rabbit—P-Akt ser-243 | Cell Signalling, #9271 | AB_329825 | 1:500 |
| Rabbit—Akt | Cell Signalling, #9272 | AB_329827 | 1:1000 |
| Rabbit—P-ERK 1/2 thr-202/tyr-204 | Cell Signalling, #9101 | AB_331646 | 1:500 |
| Rabbit—ERK 1/2 | Cell Signalling, #9102 | AB_330744 | 1:1000 |
| Rabbit—P-MEK ser-217/221 | Cell Signalling, #9121 | AB_331648 | 1:500 |
| Rabbit—MEK | Cell Signalling, #9122 | AB_823567 | 1:1000 |
| Mouse—AP50 | BD Biosciences, 611351 | AB_398873 | 1:250 |
| Mouse—Flotillin 1 | BD Biosciences, 610820 | AB_398139 | 1:500 |
| Rabbit—Caveolin 1 | Cell Signalling, #3238 | AB_2072166 | 1:1000 |
| Rabbit—GAPDH | Cell Signalling, #2118 | AB_2072166 | 1:1000 |