

Research Paper

Stromal Infiltration of Tumor-Associated Macrophages Conferring Poor Prognosis of Patients with Basal-Like Breast Carcinoma

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Abstract

Aims: Tumor associated macrophages (TAMs) play a critical role in the initiation and progression of breast cancer. However, their prognostic significance in the molecular subtype of basal-like breast cancer (BLBC) is poorly understood. The aim of this study was to investigate the extent and patterns of TAMs in BLBC and their associations with clinicopathological features and patient survival.

Methods and Results: We evaluated TAMs in 200 cases of BLBC by immunohistochemistry using the M2 macrophage marker CD163 and the pan-macrophage marker CD68 in tumor nest and stroma, and assessed their prognostic significance.

The study demonstrated that infiltration of CD163⁺ and CD68⁺ macrophages in tumor stroma was of clinical relevance in BLBC, but not those in tumor nest. Increased stromal infiltration of CD68⁺ or CD163⁺ macrophages correlated with larger tumor size, higher histological grade, higher 5-year recurrence and 5-year breast cancer mortality. Although both of CD68⁺ and CD163⁺ macrophages in tumor stroma were associated with poor recurrence-free survival (RFS) and overall survival (OS), multivariate analysis demonstrated that only CD163⁺ macrophage was an independent predictor of RFS and OS.

Conclusions: Our results highlight the prognostic importance of TAMs' location in BLBC. CD163, a highly specific biomarker for M2 macrophages, is an independent prognostic marker for BLBC patients, and may serve as an indicator or potential target of macrophage-centred therapeutic strategies.

Key words: Tumor associated macrophages, CD163, CD68, Basal-like breast cancer, Prognosis

Introduction

Inflammatory cells are a key component of the tumor microenvironment and influence the growth and progression of tumor [1, 2]. Tumor-associated macrophages (TAMs) constitute a major component of leukocyte population infiltrating tumors of the breast [3, 4]. Numerous studies have demonstrated

that TAMs enhance tumor invasion, migration and angiogenesis in many solid tumors including breast cancer [5-7], however, with the deepening of the research, controversial conclusions have always been made.

Accumulating evidences suggests that macrophages are not a homogenous population. Thus, it should be more prudent to get a conclusion in the relationship between TAMs heterogeneity and breast cancer heterogeneity. Breast cancer is one of the few types of tumor that molecular classification has been successfully used in individualized treatment design, which can significantly improve disease specific patient survival [8]. Triple-negative breast cancer is negative for hormone receptors (HR) of estrogen (ER) and progesterone (PR), and human epidermal growth factor receptor-2 (HER2), and thus cannot benefit from the targeted treatment, such as Tamoxifen, Anastrozole, and Trastuzumab [9, 10]. A number of independent studies have demonstrated the correlation of TAM with ER-negativity, PR-negativity, and high mitotic rate [11-20]. Medrek C et al. found intensity of macrophage infiltration in tumor stroma inversely correlated with luminal A phenotype and positively correlated with triple-negative/basal-like phenotype of breast cancer [20]. We noticed that the association of TAMs with poor outcome was not always correct for HR/HER status or molecular subtypes. Thus, the prognostic significance of TAMs in breast cancer should be reassessed in larger cohorts separately depending on molecular classifications. The majority of triple-negative cancers are of basal-like phenotype [21] which has the poorest prognosis. Basal-like breast cancer (BLBC) defined by Cheang et al. [22] was adopted in our study.

Macrophages exhibit striking plasticity and change their physiology according to environmental cues. This lead to heterogeneous populations of specific subsets with distinct phenotype, polarization and functions [23, 24]. TAMs can be identified as two distinct states of polarized activation: the anti-tumor/pro-inflammatory M1 macrophages and the pro-tumoral/anti-inflammatory M2 macrophages [25, 26]. M1 macrophages are characterized by expressing high levels of HLADR, inducible nitric oxide synthase (iNOS) and tumor necrosis factor- α (TNF- α), and by high microbicidal and tumoricidal activity, which are associated with a better prognosis [27]. In contrast, M2 macrophages typically express arginase 1 (ARG1), high levels of scavenging, mannose and galactose receptors, high levels of cytokines and chemokines that support their pro-tumoral functions [28]. M2 macrophages are involved in suppression of adaptive immunity, stimulation of angiogenesis, promotion of matrix remodeling and tumor invasion and migration.

In some cases, CD68⁺ TAMs were considered as a prognostic predictor in breast cancer. However, in some multivariate analyses, CD68 was not confirmed

to be independent predictor of survival [14, 15, 17]. The most probable cause is both M1 and M2 macrophage can be stained by anti-CD68 Abs. CD163, the haemoglobin scavenger receptor, is typically expressed by TAMs and up-regulated in progressing neoplasms. In many studies, CD163 is regarded as a highly specific marker for M2-like polarized macrophages [29-32]. Some studies proved that CD163⁺ TAMs was more closely related to unfavorable clinicopathological features in breast cancer [20]. However, to our knowledge no paper has reported the presence and prognostic significance of CD68⁺ and CD163⁺ macrophages in the basal-like subset of breast cancer by molecular classification. Therefore, the aim of this study was to investigated the extent of infiltration and distribution patterns of macrophages in BLBC, to compare TAMs labeled by CD163 with those identified by the more frequently used pan-macrophage marker CD68, and to determine their association with clinicopathological features and prognostic value in BLBC.

Materials and Methods

Patients

We included 200 cases of BLBC from 1,971 contiguous cases of invasive breast cancer diagnosed at the Department of Pathology, Tianjin Medical University Cancer Hospital (Tianjin, China), between April 2006 and December 2007. Neoadjuvant therapy (NAT) will seriously affect the immune cells including TAMs in the tumor microenvironment, so all the patients selected in this study are BLBC patients who did not receive NAT before surgery. And we chose BLBCs that contain only IDC-NOS, which have relatively uniform molecular and pathological features (CONSORT diagram, Fig. 1). The patients were followed up for 12-86 months (median, 66 months), and the range of the patients age at the time of diagnosis was 26-81 years (median, 51 years). During the follow-up, 42 (21%) patients died of cancer, and 66 patients (33%) developed a breast cancer recurrence, included 9 (4.5%) with local or regional tumor recurrence and 57 (28.5%) with distant metastasis. All the patients underwent modified radical mastectomy or breast-conserving surgery with axillary lymph node dissection. Postoperatively, all the patients received adjuvant chemotherapy and 156 (78%) received radiation therapy. All patients signed an informed consent form for the use of their biological tissues in the study. The study was reviewed and approved by the Institutional Ethics Committee of Tianjin Medical University Cancer Institute and Hospital, Tianjin, China (No. bc2015005).

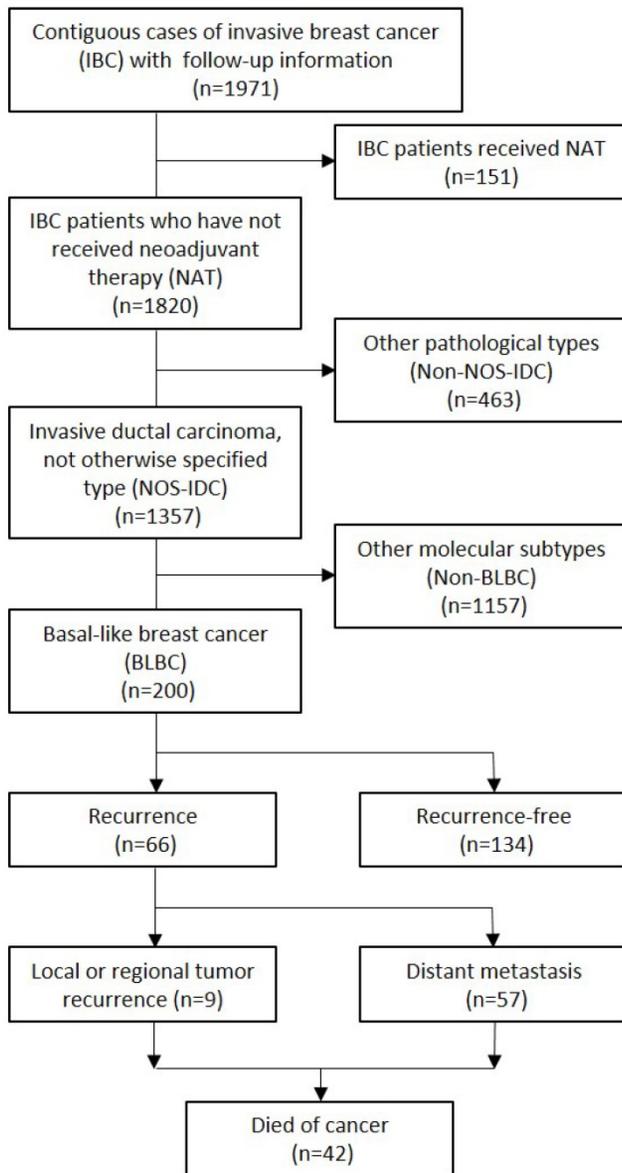


Fig. 1. CONSORT diagram.

Immunohistochemistry

Formalin-fixed, paraffin-embedded serial tissue sections of full block-face tissue from surgical specimen of each case were selected for CD68⁺ and CD163⁺ immunohistochemical stains using standard procedures. Briefly, 4 μ m tissue sections were subsequently dewaxed and rehydrated using xylene and graded alcohol washes. Antigen retrieval was performed at 121°C for 2 min, using citrate buffer, pH 6.0. After serial blocking with hydrogen peroxide and normal goat serum, the sections were incubated with primary polyclonal antibody against CD68 (Abcam, ab213096, monoclonal [KP1], 0.5 μ g/ml) and CD163 (Abcam, ab74604, monoclonal [10D6]) for 30 min at room temperature. Then, the sections were sequentially incubated with biotinylated goat

anti-mouse immunoglobulin and peroxidase-conjugated streptavidin.

TAMs were scored as the infiltration density of CD68⁺ or CD163⁺ cells with a monocyte/macrophage morphology with oval or rounded nuclei that showed strong membranous or cytoplasmic staining. In this study, TAMs in tumor nest (TN) is defined as the intraepithelial tumor-infiltrating macrophages. The definition of 'tumor stroma (TS)' in this study is the fibrous tissue surrounding tumor nest (Fig. 2); the lymphoid infiltrates surrounding the tumor bed are excluded. TAMs were evaluated by adapting the reported hotspot quantitative method.[17, 18, 33-36] The entire stained section was first scanned at low magnification ($\times 100$) to identify the hotspot areas (five areas of TN and five areas of TS) with most abundant macrophages. The average count of macrophages in all five areas and the density of infiltration was estimated (per 0.24mm²) at higher magnification ($\times 400$). For statistical analyses, the density of CD163⁺ macrophages in the TS were dichotomized into low (0-36 macrophages/0.24mm²) and high infiltration (>36 macrophages/0.24mm²) according to the median (Table 1). CD68⁺ macrophages in the TS were also scored by this critical value to unify the scoring criteria. Similarly, the density of CD163 in the TN was denoted as low (0-11 macrophages/0.24mm²) and high (>11 macrophages/0.24mm²) according to the median (Table 1), and the density of CD68 in the TN was also assessed by this critical value. Two researchers (pathologists) blinded to the clinical information carried out the evaluation process.

Additional immunohistochemistry was performed on serial tissue sections for molecular sub-classification of the tumors. Primary antibodies against ER (clone SP1, 1:150 dilution, Zymed, San Francisco, CA), PR (clone SP2, 1:150 dilution, Zymed), and HER2 (DAKO HercepTest™, Denmark), Ki-67 (clone SP6, 1:200 dilution, ThermoScientific, Fremont CA), EGFR (clone 31G7, 1:100 dilution, Zymed), and CK5/6 (clone D5/16B4, 1:100 dilution, Zymed) were applied according to manufacturer's instructions. Interpretation and scoring of ER, PR, HER2, Ki-67, EGFR, and CK5/6 staining described by Cheang et al. [22] was adopted in our study. Basal-like breast cancer was defined as ER negative, PR negative, HER2 negative, and EFGR and/or CK5/6 positive. Confirmatory fluorescence in situ hybridization (FISH) evaluation using Vysis kit (Vysis, Inc., Downers Grove, IL) was pursued on those tumors with a 2⁺ HER2 expression by immunohistochemistry. The interpretation was based up the manufacture guideline.

Statistical Analysis

Spearman's Rho test was used to examine whether the expression levels of CD68 and CD163 in macrophages were associated with clinicopathological characteristics. Wilcoxon Signed Ranks Test was performed for comparing macrophages between different tissue locations. Overall survival (OS) was measured from the date of curative surgery to the date of breast cancer-specific death. Recurrence-free survival (RFS) was measured from the date of curative surgery to the date of breast cancer recurrence, regardless of whether recurrence was local or regional, and/or distant metastasis. Kaplan-Meier analysis and long-rank tests were performed to identify the difference in OS and RFS according to CD68⁺ and CD163⁺ macrophage infiltration. Cox regression proportional hazards models were performed to estimate the hazard ratios (HR) for RFS and OS of BLBC according to CD68⁺ and CD163⁺ macrophage infiltration in univariate and multivariate models. A two-sided $P < 0.05$ was considered significant. Statistical analysis was performed with SPSS 19.0 software (SPSS, IBM).

Results

Characterization of CD68⁺ and CD163⁺ macrophages in BLBC

As shown in Fig. 2, CD68⁺ macrophages and CD163⁺ macrophages were present in both TS and TN of BLBC. We evaluated the macrophages in TS and TN separately and found significantly more macrophages infiltrated in TS than in TN (CD68: median 40 vs. 13; $Z = -11.725$, $p < 0.001$; CD163: median 36 vs. 11; $Z = -10.665$, $p < 0.001$; Table 1 and Fig. 2). The ratio of TN/total (CD68: median 0.24; CD163: median 0.24) and TS/Total (CD68: median 0.72; CD163: median 0.73) were also shown in Table 1. There was a strong correlation between CD163 and CD68 in both TS ($r_s = 0.404$, $p < 0.001$) and TN ($r_s = 0.621$, $p < 0.001$). No correlation was identified between the infiltration density of CD163⁺ macrophages in the TS and in the TN ($r_s = 0.060$, $p = 0.399$) (Table 1), while a significant correlation between the infiltration density of CD68⁺ macrophages in the TS and in the TN ($r_s = 0.275$, $p < 0.001$) (Table 1) was identified.

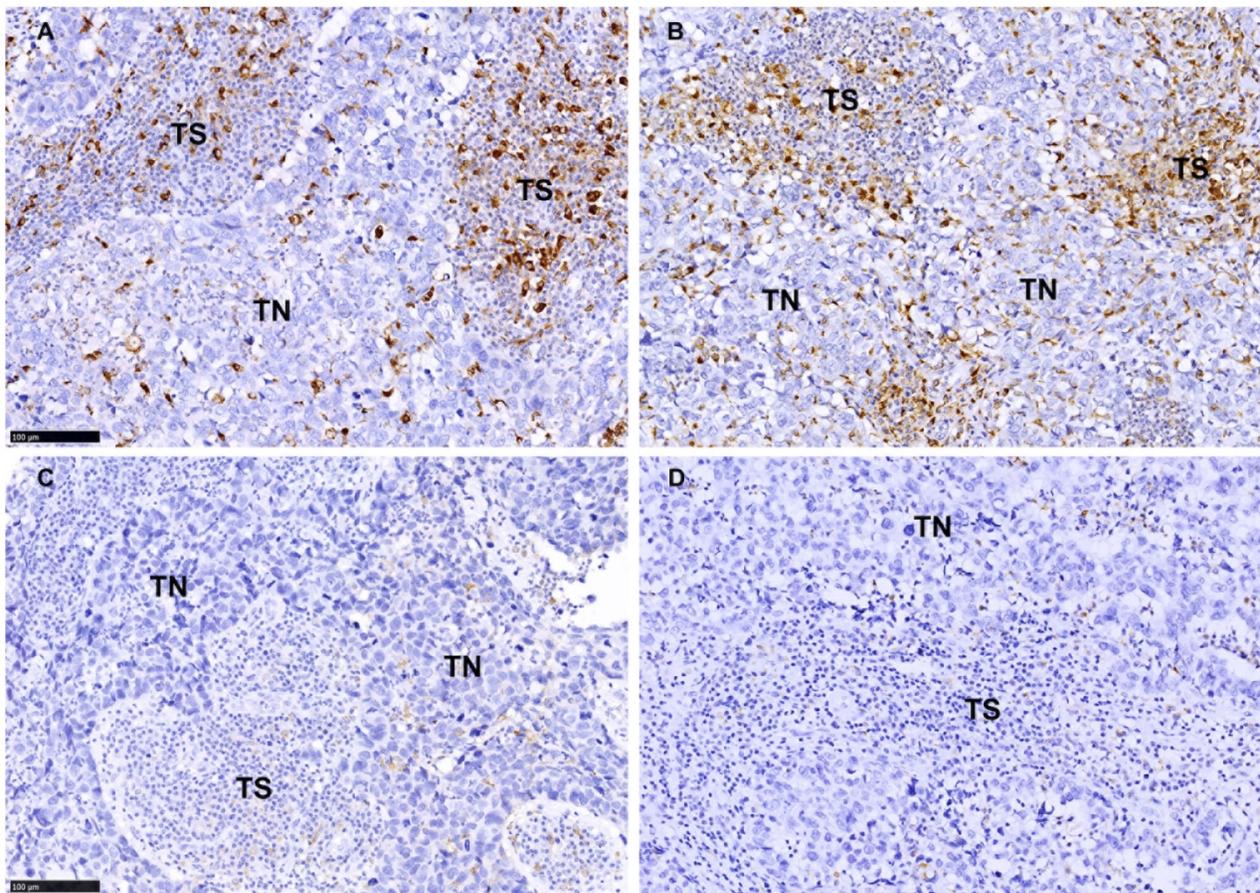


Fig. 2. The density and distribution pattern of macrophage infiltration characterized by CD68 and CD163 immunoreactivity in the tumor nest (TN) and tumor stroma (TS) of BLBC. **A-B** Representative images of high density of CD68⁺ staining (**A**) and CD163⁺ staining (**B**) in TN and TS. **C-D** Representative images of low density of CD68⁺ staining (**C**) and CD163⁺ staining (**D**) in TN and TS. Scale bar, 100 μm

Table 1. Distribution pattern of TAMs in BLBC

Variables	Mean ^a	SE ^a	Median ^a	Range ^a	Z ^b	P ^b	r _s ^c	P ^c
CD68⁺ macrophage					-11.725	<0.001	0.275	<0.001
Tumor nest	15	1.01	13	2-79				
Tumor stroma	46	2.56	40	5-197				
CD163⁺ macrophage					-10.665	<0.001	0.060	0.399
Tumor nest	14	0.92	11	1-75				
Tumor stroma	42	2.09	36	3-184				
TN/total								
CD68 ⁺ macrophage	0.23	0.01	0.24	0-0.63				
CD163 ⁺ macrophage	0.25	0.01	0.24	0-0.61				
TS/Total								
CD68 ⁺ macrophage	0.72	0.01	0.72	0.38-1				
CD163 ⁺ macrophage	0.74	0.01	0.73	0.35-0.94				

^a Cell count per field (cells/0.24 mm²)

^b Wilcoxon signed ranks test

^c Spearman's Rho test

Correlations of CD68⁺ and CD163⁺ macrophages to clinicopathological features

High infiltrations of CD68⁺ macrophage and CD163⁺ macrophage in the TS were both positively associated with larger tumor size ($r_s=0.203$, $p=0.004$; $r_s=0.284$, $p<0.001$, respectively), higher histological grade ($r_s=0.153$, $p=0.030$; $r_s=0.581$, $p<0.001$, respectively), higher 5-year recurrence rate ($r_s=0.217$, $p=0.002$; $r_s=0.145$, $p=0.041$, respectively) and higher 5-year breast cancer mortality ($r_s=0.215$, $p=0.002$; $r_s=0.178$, $p=0.011$, respectively). Higher density of CD163⁺ macrophage ($r_s=-0.380$, $p<0.001$), but not CD68⁺ macrophage ($r_s=-0.124$, $p=0.080$), in the TS was observed in tumors of younger patients. There was no significant association between stromal CD68⁺ and CD163⁺ macrophage infiltration and lymph node metastasis ($r_s=0.130$, $p=0.067$; $r_s=0.029$, $p=0.688$, respectively) (Table 2 and 3). In contrast, neither CD68⁺ macrophages nor CD163⁺ macrophages in the TN was found significantly correlated with any of the above clinicopathological features ($p>0.05$; Table 2 and 3).

Prognostic significance of CD68⁺ macrophages and CD163⁺ macrophages

Univariate Cox proportional hazards regression analysis identified that the high density of CD68⁺ and CD163⁺ macrophage infiltration in TS was associated with poor OS (HR=2.985, $p=0.005$; HR=3.548, $p<0.001$) and RFS (HR=2.229, $p=0.005$; HR=3.975, $p<0.001$) (Table 4; Fig. 3). In contrast, there was no correlation between CD68⁺ or CD163⁺ macrophages in TN with OS (HR=0.951, $p=0.870$; HR=1.322, $p=0.371$) and RFS (HR=0.925, $p=0.753$; HR=1.496, $p=0.108$) (Table 4; Fig. 3).

Multivariate Cox proportional hazards regression analysis, after adjusting for patient age, tumor size, histological grade, lymph node stage and radiotherapy, proved CD163⁺ macrophages in TS, but

not CD68⁺ macrophages, was an independent prognostic factor for worse OS (HR=3.427, $p=0.009$) and RFS (HR=4.304, $p<0.001$) (Table 5).

Table 2. Correlations between CD68⁺ macrophages and clinicopathological features in BLBC by their locations

Variables	CD68 in TS				CD68 in TN			
	N	High, n (%)	r _s	P	High, n (%)	r _s	P	
Age, years			-0.124	0.080		-0.061	0.388	
< 50	98	62(63.3)			55(56.1)			
≥50	102	52(51.0)			51(50.0)			
Tumor size, cm			0.203	0.004		0.061	0.389	
≤2	60	25(41.7)			29(48.3)			
> 2	140	89(63.6)			77(55.0)			
Histological Grade (SBR)			0.153	0.030		0.037	0.608	
Grade 1	0	0			0			
Grade 2	94	46(48.9)			48(51.1)			
Grade 3	106	68(64.2)			58(54.7)			
Lymph node metastasis			0.130	0.067		0.002	0.978	
Negative	123	63(51.2)			67(54.5)			
1 to 3	43	30(69.8)			19(44.2)			
4 to 9	22	14(63.6)			8(36.4)			
10 or more	12	7(58.3)			12(100)			
Recurrence (5 years)			0.217	0.002		0.041	0.568	
No	156	80(51.3)			81(51.9)			
Yes	44	34(77.3)			25(56.8)			
Breast cancer mortality (5 years)			0.215	0.002		0.022	0.758	
No	162	84(51.9)			85(52.5)			
Yes	38	30(78.9)			21(55.3)			

TS, tumor stroma; TN, tumor nest; BLBC, basal-like breast cancer.

Table 3. Correlations between CD163⁺ macrophages and clinicopathological features in BLBC by their locations

Variables	CD163 in TS				CD163 in TN			
	N	High, n (%)	r _s	P	High, n (%)	r _s	P	
Age, years			-0.380	<0.001		-0.100	0.159	
< 50	98	68(69.4)			54(55.1)			
≥50	102	32(31.4)			46(45.1)			
Tumor size, cm			0.284	<0.001		0.022	0.759	
≤2	60	17(28.3)			29(48.3)			
> 2	140	83(59.3)			71(50.7)			
Histological Grade (SBR)			0.581	<0.001		-0.020	0.778	
Grade 1	0	0			0			
Grade 2	94	18(19.1)			48(51.1)			
Grade 3	106	82(77.4)			52(49.1)			
Lymph node metastasis			0.029	0.688		0.011	0.872	
Negative	123	58(47.2)			62(50.4)			
1 to 3	43	29(67.4)			19(44.2)			
4 to 9	22	8(36.4)			11(50.0)			
10 or more	12	5(41.7)			8(66.7)			
Recurrence (5 years)			0.145	0.041		0.072	0.308	
No	156	72(46.2)			75(48.1)			
Yes	44	28(63.6)			25(56.8)			
Breast cancer mortality (5 years)			0.178	0.011		0.051	0.473	
No	162	74(45.7)			79(48.8)			
Yes	38	26(68.4)			21(55.3)			

TS, tumor stroma; TN, tumor nest; BLBC, basal-like breast cancer.

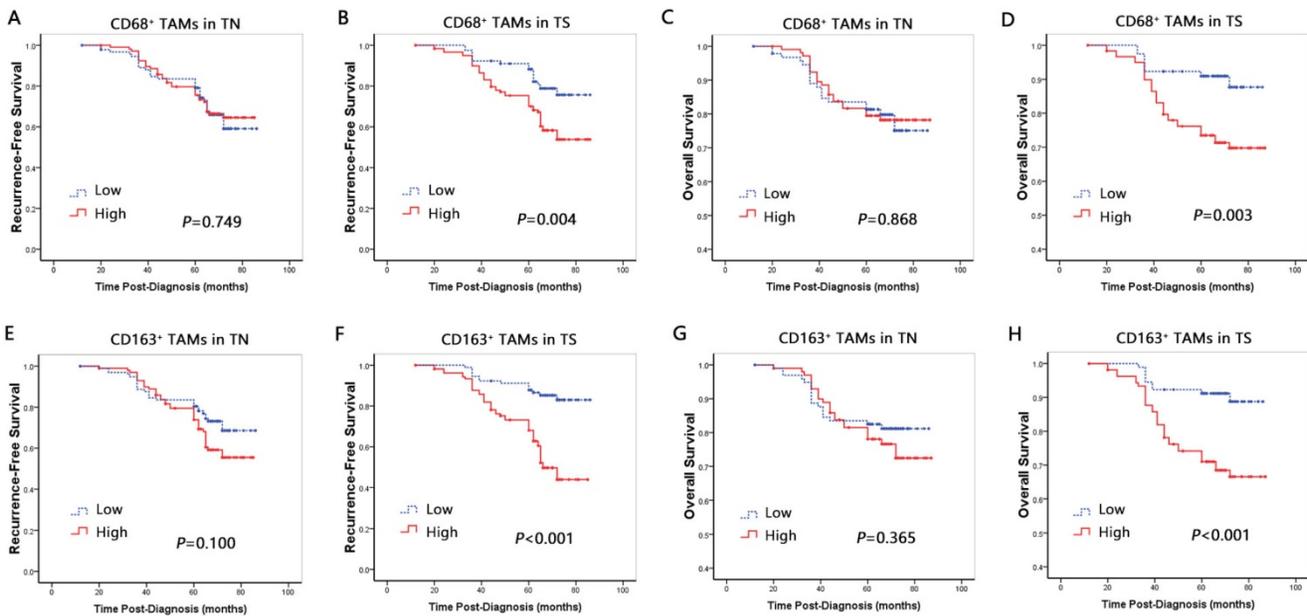


Fig. 3. RFS and OS curves based on macrophage properties. **A–D** Recurrence-free survival and Overall survival curves by CD68-positive macrophage infiltration into the tumor nest (**A, C**) and stroma (**B, D**). **E–H** Overall survival curves by CD163-positive M2 macrophage infiltration into the tumor nest (**E, G**) and stroma (**F, H**).

Table 4. Univariate Cox regression analyses for RFS and OS of BLBC

Variables	OS			RFS		
	HR	95%CI	P	HR	95%CI	P
Age (<50 vs. ≥50)	1.937	0.277-0.973	0.038	1.472	0.401-1.149	0.150
Tumor size, cm (≤2 vs. >2)	4.999	1.782-14.023	0.002	4.314	2.055-9.054	<0.001
Histological Grade (II vs. III)	1.105	0.599-2.040	0.749	1.235	0.753-2.026	0.402
Lymph node stage (N0 vs. N1-3)	1.932	1.490-2.504	<0.001	1.929	1.573-2.366	<0.001
Radiotherapy (no vs. yes)	3.606	2.043-6.364	<0.001	5.755	3.217-10.294	0.000
CD68+ in TS (low vs. high)	2.985	1.381-6.449	0.005	2.229	1.269-3.914	0.005
CD163+ in TS (low vs. high)	3.548	1.697-7.417	<0.001	3.975	2.200-7.180	<0.001
CD68+ in TN (low vs. high)	0.951	0.519-1.742	0.870	0.925	0.571-1.500	0.753
CD163+ in TN (low vs. high)	1.322	0.717-2.436	0.371	1.496	0.916-2.445	0.108

BLBC, basal-like breast cancer; TS, tumor stroma; TN, tumor nest; OS: overall survival; RFS: recurrence-free survival; HR: hazard ratio; CI: confidence interval.

Table 5. Multivariate Cox regression analyses for RFS and OS of BLBC

Variables	OS			RFS		
	HR	95%CI	P	HR	95%CI	P
Age (<50 vs. ≥50)	2.463	0.184-0.897	0.026	2.032	0.244-0.991	0.047
Tumor size, cm (≤2 vs. >2)	3.020	1.030-8.854	0.044	2.581	1.192-5.588	0.016
Histological Grade (II vs. III)	1.435	0.647-3.180	0.374	1.367	0.722-2.588	0.338
Lymph node stage (N0 vs. N1-3)	1.748	1.311-2.331	<0.001	1.836	1.468-2.297	<0.001
Radiotherapy (no vs. yes)	1.328	0.938-1.882	0.110	1.190	0.906-1.564	0.212
CD68+ in TS (low vs. high)	1.850	0.796-4.298	0.153	1.250	0.666-2.345	0.487
CD163+ in TS (low vs. high)	3.427	1.368-8.587	0.009	4.304	1.994-9.286	<0.001

BLBC, basal-like breast cancer; TS, tumor stroma; OS: overall survival; RFS: recurrence-free survival; HR: hazard ratio; CI: confidence interval.

Discussion

Macrophages are heterogeneous subsets with different functions. CD68 has been used to recognize tumoricidal M1 and pro-tumoral M2 macrophages. Even though multiple clinical studies had evidenced

that CD68+ macrophage positively associated with high vascularity and nodal metastasis and negatively correlated with RFS and OS in breast cancer, as a pan-macrophage marker, CD68 was found not an independent predictor of OS in some multivariate analyses [14, 15, 17]. A prior study using tissue microarrays (TMAs) with a large sample size suggests that overall numbers of CD68+ macrophages are not related to prognosis in breast cancer [15]. These results demand further investigation to identify the subset of TAMs that may service as the prognosis-related biomarkers. Meanwhile, CD163, a highly specific biomarker for M2 macrophages [29, 32], was found to be closely correlated with unfavorable clinicopathological characters in some study.[20] It has been recently found that BLBC hosts more TAMs infiltration, particularly the CD163+ cells, than other types of breast cancers [20]. However, the study with a small number of BLBC cases did not explore the prognostic significance of CD68+ and CD163+ macrophages in BLBC. We therefore undertook the study to evaluate the infiltration of stromal and intraepithelial CD68+ macrophages along with the identification of CD163+ macrophages to assess their prognostic value in a larger BLBC cohort. In addition, TAMs in different intratumoral regions are subsets with distinct molecular and functional features, and thus may exhibit different prognostic values. Unfortunately, limited studies have considered the locations in the evaluation of clinical significance. Although three studies (two by marker of CD68 [15, 33] and one by markers of CD68 and CD163 [20]) in human breast cancer have included the locations of TAMs, two of which used the TMA

samples to put it into practice [15, 20]. We used full block-face tissue sections in the study to overcome the limitation of core materials that only contain small volume of tumor tissue may suffer from bias in the assessment of TAMs due to tumor heterogeneity. Full block-face tissue sections allow the selection of tumor areas with abundant TAMs for accurate scoring in tumor nest and in tumor stroma separately.

In this cohort of BLBC, CD68⁺ or CD163⁺ macrophages in TN was not found to be correlated with any of the clinicopathological features, OS or RFS. Medrek et al. recently also found no correlations between clinicopathological features and CD68⁺ or CD163⁺ macrophage infiltrates in tumor nest, and the same relationship with RFS and OS was also noted [20]. Mahmoud et al. reported that intratumoural CD68⁺ macrophage did not correlate with breast cancer-specific survival [15]. Although the methods and molecular subtypes of breast cancer patients were different in these studies, the results suggest that the TAMs infiltrates into tumor nest is not an important prognostic factor for patient outcome.

TAM promotes tumor progression through several mechanisms [37]. In contrast to TN, the study found that increased CD68⁺ or CD163⁺ macrophages in TS were correlated with larger tumor size, higher histological grade, and worse RFS and OS. It suggested that TAMs may affect the prognosis of BLBC mainly through modulating the immune response by contacting with tumor-infiltrating lymphocytes in TS, rather than through the direct interaction between macrophages and cancer cells in TN, such as phagocytosis of cancer cells. However, the specific underlying mechanisms need further study. It is worthy of notice that although univariate analysis identified the total TAM infiltrates in TS was associated with poor RFS and OS in BLBC, multivariate analysis only proved CD163⁺ macrophages in TS was an independent predictor of RFS and OS. We also got similar conclusions in multivariate Cox regression analyses using continuous variables, one-quarter and median as the cutoffs, i.e., CD163⁺ macrophages in TS was an independent prognostic factor for worse OS and RFS (Supplementary Table 1). It indicates that in contrast to the pan-macrophage marker CD68, macrophages expressing the anti-inflammatory CD163 marker (M2 macrophages) are probably the effect TAMs that facilitate the poor prognosis of BLBC and serve as an independent prognostic predictor.

A number of potential macrophage-centred therapeutic strategies are being explored [37]. Either manipulating TAMs localization at tumor sites, or targeting the signal pathways affecting TAMs polarization and functions is proved to improve the

outcomes of breast cancer [38]. The pro-tumor M2 macrophage, recognized by CD163, is reversible [39, 40]. Duluc et al. used INF- γ to reverse the immunosuppressive and pro-tumoral foundations and prevent the generation of M2 macrophage [41]. Similar to the result, Guiducci et al. activated TLR9 by its ligand CpG, along with antibody to IL-10, and switched TAMs from an M2 macrophage to an M1 macrophage [42]. And classically activated M1 macrophage can kill cancer cells and induce tumor-destructive reaction [43]. Therefore, blocking key molecular determinants of macrophage polarization may result in reorientation of macrophage polarization and activate the anti-tumor activity [39, 44, 45]. In a preclinical glioma model, a small molecule agent BZL945 (CSF-1R inhibitor) improved the survival by 're-education' of macrophages from a protumor M2-like phenotype to an antitumor M1-like type instead of TAM number depletion [46]. Although TAM-centered strategies have not been yet introduced in clinical practice in terms of effectiveness and tolerability in breast cancer patients, reducing number or reeducating of TAM seems to be a promising strategy and should be further investigated due to its clinical significance in BLBC.

There are limitations in the study. Although CD163 was regarded as a highly special M2 macrophage marker, it can also be expressed by monocyte-derived dendritic cells (MDCs) [47]. Medrek C et al. observed some CD163⁺ areas lacked CD68 expression. They attributed this to a CD163-expressing subset of immature myeloid derived suppressor cells with prognostic impact although no direct data was available [20]. Thus, we cannot exclude MDCs cells or myeloid derived cells expressing CD163, which could have impact on the poor prognosis triggered by CD163⁺ macrophage in BLBC. It is warranting to further clarify the phenotype of TAMs and the significant impact of M1/M2 macrophage ratio on the clinical course of BLBC patients. Few specific markers are so far available, and therefore there is still an urgent need to identify markers specific for M1/M2 macrophage. Some other TAM-related biomarkers that have been explored include CD206, VEGF, HIFs, TFL, PCNA and CCL18 [4]. Further investigation to identify their roles in sub-classifying TAMs into different prognosis-related biological groups will be meaningful and will be one of the focuses of our future studies.

Conclusion

This study identified that infiltration of CD68⁺ or CD163⁺ macrophages in TS, not the TN, is associated with the clinicopathological features of BLBC and

worse prognosis of patients. CD163⁺ macrophages in TS, instead of CD68⁺ macrophage, is proven to an independent risk factor for worse RFS and OS. TAMs signature is a promising therapeutic target in BLBC. However, further work is still warranting to validate the findings.

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Supplementary Material

Supplementary table.

<http://www.jcancer.org/v09p2308s1.pdf>

Competing Interests

The authors have declared that no competing interest exists.

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