

Research Paper



The alterations in peripheral lymphocyte subsets predict the efficacy and prognosis of immune checkpoint inhibitors in hepatocellular carcinoma

Qu Xie^{1, 2}, Can Hu^{3,4}, Cong Luo^{1⊠}

- 1. Department of Hepato-Pancreato-Biliary & Gastric Medical Oncology, Zhejiang Cancer Hospital, Hangzhou Institute of Medicine (HIM), Chinese Academy of Sciences, Hangzhou, Zhejiang, 310022, China.
- 2. Postgraduate training base Alliance of Wenzhou Medical University (Zhejiang Cancer Hospital), Hangzhou, Zhejiang, 310022, China.
- The Second School of Clinical Medicine of Zhejiang Chinese Medical University, Hangzhou, 310053, China.
 Zhejiang Cancer Hospital, Hangzhou Institute of Medicine (HIM), Chinese Academy of Sciences, Hangzhou, Zhejiang, 310022, China.

In Encland cancer mosphas, mangenou nounce of medicine (min), chancer nauciny of occneco, mangenou, encland, or occes, change

🖂 Corresponding author: Dr. Cong Luo, Department of Hepato-Pancreato-Biliary & Gastric Medical Oncology, Zhejiang Cancer Hospital, Hangzhou Institute of Medicine (HIM), Chinese Academy of Sciences, Hangzhou, Zhejiang 310022, China. E-mail: luocong@zjcc.org.cn.

© The author(s). This is an open access article distributed under the terms of the Creative Commons Attribution License (https://creativecommons.org/licenses/by/4.0/). See http://ivyspring.com/terms for full terms and conditions.

Received: 2023.07.14; Accepted: 2023.08.31; Published: 2023.09.11

Abstract

Background: Immune checkpoint inhibitor (ICI) treatments are promising therapies for hepatocellular carcinoma (HCC) patients. However, not all HCC patients benefit from immunotherapy. Therefore, it is urgent to explore markers for the clinical efficacy and prognosis of immunotherapy for liver cancer. This study aimed to investigate changes in peripheral blood lymphocyte subsets after immunotherapy and to assess their predictive and prognostic value.

Methods: Sixty-one patients with advanced HCC were enrolled. Peripheral blood samples were collected from HCC patients before and after ICI treatment, and lymphocytes were detected by flow cytometry. The rank sum test, chi-square test, Kaplan–Meier curve, and Cox regression model were used to determine the relationship between the changes in the percentages of peripheral blood lymphocyte subsets and clinicopathological characteristics, clinical efficacy, progression-free survival (PFS) and overall survival (OS).

Results: After ICI treatment, the percentage of CD3+CD8+ T cells increased, and the percentage of B cells decreased. The changes in memory T cells percentages varied according to different immune efficacy groups. Age, history of hepatitis B infection, first-line therapy, and distant metastasis influenced the proportion of peripheral blood lymphocyte subsets in patients with advanced HCC. Furthermore, univariate analysis demonstrated that high percentage changes in the natural killer (NK) cells percentage change predicted longer PFS and OS.

Conclusions: ICI treatment alters the percentage of peripheral blood lymphocyte subsets in immunotherapy-treated HCC patients. Changes in the proportion of lymphocyte subsets are influenced by variances in immunological response and clinicopathological features. A high degree of NK cells percentage change in HCC patients treated with ICI represents an independent prognostic predictor.

Keywords: immune checkpoint inhibitor (ICI); hepatocellular carcinoma (HCC); biomarker; peripheral blood lymphocyte subsets

Background

Hepatocellular carcinoma (HCC) is the fifth most common cancer and the third leading cause of cancer death worldwide. Chinese patients with liver cancer account for more than 40% of the world's incidence(1, 2). Because the early symptoms are not obvious, the degree of malignancy is high, and the rate of development and deterioration is fast, 75% of HCC patients have developed advanced manifestations such as distant metastasis at the time of diagnosis and cannot benefit from radical surgical resection(3). Many advances have been made in the clinical diagnosis and treatment of HCC in recent years, but the 5-year overall survival (OS) rate of HCC patients is only 12%(4). How to apply existing treatment methods to improve the cure rate and prolong the survival time of liver cancer in a planned and reasonable manner has become an urgent problem to be solved in the field of advanced liver cancer treatment.

In recent years, immune checkpoint inhibitor (ICI) treatment has promoted considerable survival benefits to patients with advanced HCC, making long-term survival possible(5). Immunotherapy with ICI blocks the specific binding of PD-1 on the surface of immune cells, such as T cells and B cells, to PD-L1 on the surface of tumour cells and blocks the PD-1/PD-L1 pathway through programmed death-1 (PD-1) antibodies or ligand (PD-L1) antibodies, thereby relieving immunosuppression and exerting tumour-killing effects(6). The compelling exploration of immunotherapy has broken through the bottleneck of liver cancer treatment for over a decade. The IMbrave 150 study showed that compared with sorafenib, the combination of atezolizumab and bevacizumab significantly prolonged OS (19.2 months vs. 13.4 months, hazard ratios (HR) =0.66, P = 0.0009) and progression-free survival (PFS) (6.9 months vs. 4.3 months, HR=0.65, P = 0.0001) in patients with advanced unresectable liver cancer(7). Although immunotherapy has made outstanding progress in the treatment of HCC, it has a low response rate (12-20% effective rate with single-agent immunization(8-10) and 27-46% effective rate with combination therapy(11-14)). Moreover, these treatments are expensive, which significantly limits the application of ICI. Therefore, finding appropriate immune efficacy markers, screening the population that would benefit from immunotherapy, and effectively improving the effective rate of immune combination therapy are urgent clinical problems to be solved.

Because lymphocytes in peripheral blood have the significant advantages of low invasiveness and real-time efficacy monitoring, an increasing number of studies have investigated their predictive and prognostic value in cancer patients in recent years(15, 16). For example, in 16 HCC patients treated with nivolumab, patients with progressive disease were more likely to have monocytes with increased PD-L1 positivity at 4 weeks (P = 0.020) or 6 weeks (P = 0.008) of treatment(17). In addition, patients with increased active circulating CD8+ T cells and downregulation of neutrophil-related markers during pembrolizumab treatment had better clinical benefits(18). However, the value of peripheral blood immune cells and their subsets in predicting the efficacy and prognosis of immunotherapy in patients with advanced HCC needs further exploration.

Therefore, in this study, we assessed changes in peripheral blood lymphocyte subsets before and after immunotherapy and their relationship with clinical efficacy and prognosis in patients with advanced HCC treated with ICI.

Materials and methods

Patients

This study included Barcelona Clinic Liver Cancer (BCLC) B or C patients who received ICI treatment at Zhejiang Cancer Hospital from January 2019 to December 2021. The inclusion criteria were as follows: (1) age > 18 years; (2) pathologically or clinically diagnosed HCC patients with BCLC stage B or C disease; (3) Eastern Cooperative Oncology Group performance status (ECOG) score of 0 to 2 and Child– Pugh grade A or B; (4) complete lymphocyte subsets and clinicopathological information; and (5) at least one measurable target lesion on baseline computed tomography (CT) or magnetic resonance imaging (MRI). Patients with a history of autoimmune diseases or with tumours other than HCC were excluded.

The Zhejiang Cancer Hospital's Ethics Committee granted approval for this retrospective study (IRB-2023-87), waiving the requirement for informed patient consent in adherence with the guidelines of the Declaration of Helsinki.

Treatment

The study cohort comprised individuals undergoing ICI treatment as a sole therapy or in conjunction with targeted therapies. Patient groups were administered ICI regimens, including atezolizumab (10 patients), camrelizumab (17 patients), tislelizumab (12 patients), and cedilimumab (22 patients). Intravenous administration was employed for atezolizumab at a dose of 1200 mg every three weeks, while the remaining ICI drugs were administered intravenously at 200 mg every three weeks. Among the 50 patients undergoing combined targeted therapies, 6 received a combination of the anti-vascular endothelial growth factor drug bevacizumab. In comparison, 42 patients received a combination of anti-tyrosine kinase inhibitors (including 8 cases of apatinib, 21 cases of lenvatinib, 4 cases of regorafenib, and 11 cases of sorafenib). Established clinical practice criteria guided the selection of targeted therapeutic regimens.

Data collection and response assessment

The collected clinicopathological data were analysed, including sex, age, hepatitis B infection history, Child–Pugh class, pretreatment alpha-fetoprotein (AFP) level, BCLC stage, ECOG score, line of therapy, combination with targeted therapy, combination with local therapy, and distant metastasis.

Peripheral venous blood was collected one week before ICI treatment and at the optimal response time. Various lymphocyte subsets were assessed, including CD3+ T cells, CD3+CD4+ T cells, CD3+CD8+ T cells, CD4/CD8 ratio, natural killer (NK) cells (CD3-CD56⁺), B cells (CD3⁻CD19⁺), natural killer T (NKT) cells (CD3+CD56+), Ts cells (CD4+CD45RA+), memory Т cells (CD4+CD45RO+), activated Т cells (CD45RA+CD45RO+) and activated CD8+ cells (CD8+CD38+).

Enhanced CT or enhanced MRI for clinical efficacy assessments was performed 6-8 weeks after immunotherapy and every 2-3 months thereafter. The Response Evaluation Criteria in Solid Tumours version 1.1 (RECIST 1.1) was used to assess the clinical response. The best overall response (BOR) during the whole ICI treatment phase was classified as complete response (CR), partial response (PR), stable disease (SD), or progressive disease (PD). Patients with response assessments of CR, PR, and SD were assigned to the responder group, and those with PD were assigned to the nonresponder group. PFS refers to the time interval from the start of treatment to tumour progression or death from any cause. In contrast, OS refers to the time interval from the beginning of treatment to death from any reason or the last follow-up.

Flow cytometry

The flow experiment steps were as follows: (1) three flow cytometry tubes were required for each peripheral blood sample and labelled Tube 1, Tube 2, and Tube 3. (2) Different premixed monoclonal antibodies were added to the three tubes, and 50 µL of anticoagulated blood was added to each tube. (3) The samples were mixed well and incubated at room temperature in the dark for 15 min. (4) Then, 500 µL of haemolysin was added and mixed well with vortex shaking, and the samples were placed in the dark for 15 min. (5) One millilitre of sheath fluid was added to terminate haemolysis. (6) The blood samples were centrifuged at 1500 rpm for 5 min, and the supernatant was discarded. (7) Then, 500 µL of sheath fluid was added to resuspend the sample, and the percentages of peripheral blood lymphocyte subsets were measured using a Beckman Coulter Cytometer FC500 flow cytometer. Detailed operating procedures and associated materials are provided in Additional file 1.

Statistical analysis

Continuous variables are presented as medians,

and categorical data are presented as counts and percentages. Comparisons between groups were made using the Wilcoxon two-independent-samples rank-sum test, and comparisons within groups were made using the Wilcoxon paired-samples rank-sum test. Categorical variables were compared using the chi-square test. In predictive analysis, for each lymphocyte subset, the median was set as a cut-off, and each subset was divided into two subgroups (high and low). Univariate Cox proportional hazards regression models were used to assess the effect of clinical history information and lymphocyte subsets (expressed as HR and their corresponding 95% confidence intervals (CI)) on PFS and OS. PFS and OS were estimated with the Kaplan-Meier method and analysed using the log-rank test. All statistical tests were two-sided, and P values <0.05 were considered statistically significant. All analyses were conducted using SPSS 25.0 software (International Business Machines Corporation, Armonk, NY, USA) and GraphPad Prism 9.0 (GraphPad Software Corporation, San Diego, CA, USA).

Results

Participant characteristics

From 01 January 2019 to 31 December 2021, 61 patients with advanced HCC met the inclusion and exclusion criteria (Figure 1). The baseline characteristics of all patients are shown in Table 1. Forty-two (68.9%) patients were 60 years or younger, and 51 (83.6%) were male. Of these patients, the majority had hepatitis B infection (46, 75.4%). Fifty-three (86.9%) patients had Child-Pugh class ratings of A, while 8 (13.1%) patients had class ratings of B. In addition, 26 (42.6%) patients had an AFP level of \geq 400 IU/ml at baseline. The BCLC stage was C in 50 (82.0%) patients and B in 11 (18.0%). In addition, 43 patients (70.5%) had an ECOG score of 1, and 18 others (29.5%) had an ECOG score of 0. Forty-four patients (72.1%) received first-line therapy, 50 patients (82.0%) were treated with concomitant targeted treatment, and 24 patients (39.3%) were treated with concomitant local treatment. In addition, 27 patients (44.3%) had distant metastasis.

Efficacy

As of 14 November 2022, the median follow-up time was 20.1 months (95% CI: 16.0 to 24.2 months). The median PFS and OS were 8.3 months (95% CI: 5.5 to 11.1 months) and 19.4 months (95% CI: 6.0 to 32.8), respectively. The objective response rate (ORR) was 32.79% [20/61, 0 CR, 20 PR]. The disease control rate (DCR) was 78.69% [48/61, 0 CR, 20 PR, 28 SD].



Figure 1. Workflow of this study. Abbreviation: HCC, hepatocellular carcinoma.

Changes in lymphocyte subsets in peripheral blood after ICI treatment

The proportions of changes in peripheral blood lymphocyte subsets after ICI treatment are shown in Figure 2 and Table 2. The results showed an increase in the percentage of CD3+CD8+ T cells (P = 0.034) and a decrease in the percentage of B cells (P = 0.036) after immunotherapy compared with baseline (Figure 2 and Table 2). This suggests an effect of ICI treatment on peripheral lymphocyte subsets.

Table 1.	Clinical a	nd patho	logical o	characteristics.
----------	------------	----------	-----------	------------------

	N	0/_
Carr	Ĩ	70
Sex		
Male	51	83.6
Female	10	16.4
Age		
≤60	42	68.9
>60	19	31.1
Type of hepatitis		
No hepatitis	15	24.6
Hepatitis B	46	75.4
Child-Pugh class		
А	53	86.9
В	8	13.1

	N	%	
AFP			
<400(IU/ml)	35	57.4	
≥400(IU/ml)	26	42.6	
BCLC stage			
В	11	18.0	
С	50	82.0	
ECOG PS			
0	18	29.5	
1	43	70.5	
Line of therapy			
First line	44	72.1	
Later lines	17	27.9	
Combined targeted therapy			
No	11	18.0	
Yes	50	82.0	
Combined local therapy			
No	37	60.7	
Yes	24	39.3	
Drinker			
No	35	57.4	
Yes	26	42.6	
Distant metastasis			
No	34	55.7	
Yes	27	44.3	

Abbreviations: AFP, alpha-fetoprotein; BCLC, Barcelona Clinic Liver Cancer; ECOG PS, Eastern Cooperative Oncology Group Performance Status.



Figure 2. Percentage of different subpopulations of peripheral blood lymphocytes before and after Immune checkpoint inhibitor (ICI) treatment. (A) CD3⁺ T cells, (B) CD3⁺CD4⁺ T cells, (C) CD3⁺CD8⁺ T cells, (D) CD4/CD8 ratio, (E) natural killer (NK) cells (CD3⁻CD56⁺), (F) B cells (CD3⁻CD19⁺), (G) natural killer T (NKT) cells (CD3⁺CD56⁺), (H) Ts cells (CD4⁺CD45RA⁺), (I) memory T cells (CD4⁺CD45RO⁺), (J) activated T cells (CD45RA⁺CD45RO⁺), and (K) activated CD8⁺ cells (CD8⁺CD38⁺). *, P<0.05.

 Table 2. Percentage of different subpopulations of peripheral blood lymphocytes before and after ICI treatment.

Lymphocyte subset	Percentage of HCC patients								
	Before (median (IQR))	After (median (IQR))	Р						
CD3+ T cells	69.3 (60.2,78.7)	72.2 (60.5,75.1)	0.874						
CD3+CD4+ T cells	42.4 (35.4, 49.3)	38.4 (35.2,48.3)	0.653						
CD3+CD8+ T cells	22.5 (16.3 26.6)	23.9 (18.6,27.6)	0.034						
CD4/CD8 ratio	1.8 (1.5, 2.6)	1.7 (1.4,2.4)	0.098						
NK cells	16.5 (10.7, 28.4)	21.0 (12.3,28.8)	0.367						
B cells	8.4 (5.1,12.0)	7.0 (4.3,13.5)	0.036						
NKT cells	2.8 (1.7, 4.1)	2.7 (2.0,3.6)	0.707						
Ts cells	10.2 (5.1, 14.6)	9.5 (5.8,14.1)	0.236						
Memory T cells	25.3 (23.3, 33.0)	24.7 (22.5,31.6)	0.411						
Activated T cells	2.8 (2.3, 3.4)	2.7 (2.5,3.5)	0.661						
Activated CD8+ T cells	6.3 (4.0,9.3)	6.9 (5.3,8.8)	0.719						

Abbreviation: IQR, Inter-Quartile Range.

Table 3. Correlation between the changes in the proportions oflymphocyte subsets in peripheral blood and efficacy after ICItreatment.

Ν	Response		Non-res	ponse	P value
	Populat	ion %	Populat	ion %	-
61	48	78.69	13	21.31%	
					0.517
28	21	75.00	7	25.00	
33	27	81.82	6	18.18	
					0.460
32	24	75.00	8	25.00	
29	24	82.76	5	17.24	
					0.065
24	16	66.67	8	33.33	
37	32	86.49	5	13.51	
					0.732
35	27	77.14	8	22.86	
26	21	80.77	5	19.23	
					0.202
28	20	71.43	8	28.57	
33	28	84.85	5	15.15	
					0.661
49	38	77.55	11	22.45	
12	10	83.33	2	16.67	
					0.561
	N 61 28 33 29 24 37 35 26 28 33 49 12	N <u>Respons</u> <u>Populati</u> 61 48 28 21 33 27 32 24 29 24 29 24 24 16 37 32 35 27 26 21 28 20 33 28 49 38 12 10	N Response Population % 61 48 78.69 28 21 75.00 33 27 81.82 34 75.00 29 24 75.00 29 24 82.76 24 37 32 86.49 35 27 77.14 26 21 80.77 28 20 71.43 33 28 84.85 49 38 77.55 12 10 83.33	N Response Non-res Population % Populat 61 48 78.69 13 28 21 75.00 7 33 27 81.82 6 32 24 75.00 8 29 24 82.76 5 24 16 66.67 8 37 32 86.49 5 35 27 77.14 8 26 21 80.77 5 28 20 71.43 8 33 28 84.85 5 49 38 77.55 11 12 10 83.33 2	N Response Non-response Population % Population % 61 48 78.69 13 21.31% 28 21 75.00 7 25.00 33 27 81.82 6 18.18 32 24 75.00 8 25.00 29 24 82.76 5 17.24 24 16 66.67 8 33.33 37 32 86.49 5 13.51 35 27 77.14 8 22.86 26 21 80.77 5 19.23 28 20 71.43 8 28.57 33 28 84.85 5 15.15 49 38 77.55 11 22.45 12 10 83.33 2 16.67

Variable	Ν	Respons	se	Non-	response	P value
		Population %		Popu	lation %	
Decrease	38	29	76.32	9	23.68	
Increased	23	19	82.61	4	17.39	
Ts cells						0.608
Decrease	32	26	81.25	6	18.75	
Increased	29	22	75.86	7	24.14	
Memory T cells						0.016
Decrease	39	27	69.23	12	30.77	
Increased	22	21	77.42	1	22.58	
Activated T cells						0.460
Decrease	32	24	75.00	8	25.00	
Increased	29	24	82.76	5	17.24	
Activated CD8+ T cells						0.984
Decrease	28	22	78.57	6	21.43	
Increased	33	26	78.79	7	21.21	

Correlation between the proportion change in lymphocyte subsets in peripheral blood and efficacy after ICI treatment

After immunotherapy, 48 patients were classified as responders (0 CR, 20 PR, 28 SD), and 13 were classified as nonresponders. In addition, the changes in the percentage of lymphocytes were divided into decreased groups and increased groups. The results showed that the effective rate was 69.23% in the decreased memory T cells group and 77.42% in the increased memory T cells group (Table 3). There was a significant difference in the effective rate between the two groups (P = 0.016).

Factors influencing lymphocyte subsets

We collected clinicopathological information before ICI treatment and analysed its effect on the percentage of changes in different lymphocyte subsets (Table 4). The results showed that the percentage of activated T cells and activated CD8⁺ cells decreased in HCC patients over 60 years of age after ICI treatment compared with patients under 60 years (Table 4). In addition, hepatitis B infection history may influence the changes in CD3⁺ T cells and activated CD8⁺ T cells after ICI treatment (Table 4). The percentage of CD3⁺ T cells decreased and that of activated CD8⁺ T cells increased in patients with a history of hepatitis B infection after ICI treatment compared with patients without a history of hepatitis B infection (Table 4). Moreover, compared with HCC patients who did not receive first-line therapy, those who did receive first-line therapy after ICI therapy had decreased proportions of B cells and memory T cells (Table 4). Changes in the percentages of CD3⁺CD4⁺ T cells, the CD4/CD8 ratio, and B cells were associated with distant metastasis. Compared with HCC patients without distant metastasis, HCC patients with distant metastasis after ICI treatment had decreased percentages of CD3+CD4+ T cells and CD4/CD8 ratio, while the percentage of B cells increased (Table 4). These results suggest that age, history of hepatitis B infection, first-line therapy, and distant metastasis may influence the change in the percentage of peripheral blood lymphocyte subsets after ICI treatment.

Correlation between clinicopathological features, baseline lymphocyte subset percentages or changes in peripheral blood lymphocyte subset percentages and survival

Among the 61 HCC patients, the median PFS was 8.3 months (95% CI: 5.5 to 11.1 months), and the median OS was 19.4 months (95% CI: 6.0 to 32.8). Univariate analysis showed that all clinicopathological characteristics and the percentage of peripheral blood lymphocyte subsets before ICI treatment were not prognostic factors for PFS and OS in HCC patients treated with ICI. However, the changes in the percentage of NK cells after ICI treatment were independent prognostic factors for PFS (P = 0.011) and OS (P = 0.023) in HCC patients treated with ICI, and the changes in the percentage of other lymphocyte subsets were not prognostic factors for PFS and OS (Figure 3 A and B). Survival analysis was performed to further determine the predictive value of ICI treatment based on peripheral blood lymphocyte subsets in HCC patients. The results showed that HCC patients with a high percentage of NK cells had longer OS and PFS than HCC patients with a low percentage of NK cells after ICI treatment, which was consistent with the univariate Cox regression analysis results (Figure 4).

Table 4. Correlation between the changes in peripheral blood lymphocyte subset percentages and clinicopathological features after ICI treatment.

	CD3⁺ T c	ells	CD3+CD4 cells	1 ⁺ T	CD3+CD8 cells	8+ T	CD4/CD8 ratio	3	NK cells		B cells		NKT cell	s	Ts cells		Memory cells	Т	Activated cells	ΙT	Activateo CD8⁺ cel	1 ls
	z (after-be fore)	Р	z (after-be fore)	Р	z (after-be fore)	Р	z (after-be fore)	Р	z (after-be fore)	Р	z (after-be fore)	Р	z (after-be fore)	Р	z (after-be fore)	Р	z (after-be fore)	Р	z (after-be fore)	Р	z (after-be fore)	Р
Age		0.1		0.8		0.9		0.6		0.5		0.3		0.3		0.4		0.7		0.0		0.0
		23		09		63		46		75		96		03		00		20		49		46
≤60 years	1.3		-2.1		1.4		-0.2		3.1		-3.6		-0.4		-2.1		-2.6		0.3		0.7	
>60	3.3		-0.7		1.8		-0.3		2.5		-2.4		-0.3		0.2		-2.8		-0.2		-1.2	
years																						
Type of hepati		0.0 24		0.6 09		0.9 33		0.8 93		0.3 53		0.1 01		0.8 01		0.7 00		0.1 47		0.5 97		0.0 34
115	0.6		2.4		4.0		0.2		11		2.2		0.5		10		20		0.2		11	
hepatit is	9.0		-2.4		4.0		-0.5		1.1		-2.2		-0.5		-1.0		5.0		-0.2		-1.1	
hepatit is B	-0.6		1.5		0.9		-0.3		2.9		-3.6		-0.3		0.4		-2.9		0.1		0.7	
Line		0.3		0.8		0.5		0.8		0.5		0.0		0.5		0.7		0.0		0.3		0.4
of therap		81		79		20		85		68		02		57		29		29		80		69
у																						
first line	2.0		-1.2		0.9		-0.3		3.5		-3.9		-0.4		-1.9		-3.5		-0.1		0.4	
later lines	2.3		-2.7		2.6		-0.2		-1.3		-1.7		-0.3		0.2		3.8		0.4		-1.0	
Distan		0.1		0.0		0.6		0.0		0.5		0.0		0.1		0.3		0.9		0.7		0.4
t		79		21		47		09		47		18		42		83		94		38		77
metast asis																						
No	4.6		2.5		1.5		0.1		3.0		-4.0		-0.3		0.5		-2.7		-0.2		0.3	
Yes	-2.4		-3.8		1.6		-0.6		-1.3		-2.2		-0.4		-2.0		-2.8		-0.1		0.2	

	HR (95% CI)	P value
	0.748 [0.334,1.674]	0.480
	0.743 [0.391,1.414]	0.365
	1.035 [0.532,2.015]	0.919
•	→ 1.398 [0.624,3.133]	0.416
	1.441 [0.811,2.562]	0.213
- -	1.285 [0.599,2.756]	0.520
	1.234 [0.655,2.326]	0.515
	1.216 [0.649,2.275]	0.542
	0.918 [0.442,1.905]	0.818
—	0.950 [0.529,1.705]	0.863
	0.782 [0.433,1.412]	0.415
	1.106 [0.621,1.970]	0.733
	0.669 [0.371,1.206]	0.181
	0.750 [0.420,1.339]	0.331
	0.992 [0.557,1.767]	0.979
	0.923 [0.520,1.638]	0.784
	1.387 [0.769,2.498]	0.277
	1.106 [0.622,1.966]	0.732
	1.144 [0.643,2.034]	0.648
	0.973 [0.547,1.730]	0.925
•	1.303 [0.734,2.314]	0.366
	0.630 [0.351,1.132]	0.122
	0.760 [0.424,1.364]	0.358
	1.212 [0.682,2.154]	0.512
	1.136 [0.623,2.072]	0.678
	0.916 [0.514,1.630]	0.765
4	0.840 [0.472,1.497]	0.555
	0.449 [0.241,0.835]	0.011
	1.112 [0.625,1.979]	0.718
1	1.142 [0.639,2.041]	0.653
	1.617 [0.898,2.914]	0.109
	0.705 [0.397,1.253]	0.233
•1	1.297 [0.729,2.310]	0.376
	0.884 [0.498,1.571]	0.674
2	3 4	

Hazard ratio (HR)

ò

Variables

Sex (male vs. female)

BCLC stage (B vs. C)

ECOG PS (0 vs. 1)

Drinker (No vs. Yes)

Age (d60 vs. >60 years)

Child-Pugh class (A vs. B)

AFP (<400 vs. e400 IU/ml)

Type of hepatitis (no hepatitis vs. hepatitis B)

Line of therapy (first line vs. later lines)

Combined local therapy (No vs. Yes)

Baseline CD3+ T cells (Low vs. High)

Baseline CD4/CD8 ratio (Low vs. High)

Baseline Memory T cells (Low vs. High)

Baseline Activated T cells (Low vs. High)

Change of CD4/CD8 ratio (Low vs. High)

Change of Memory T cells (Low vs. High)

Change of Activated T cells (Low vs. High)

Change of Activated CD8+ cells (Low vs. High)

Change of NK cells (Low vs. High)

Change of NKT cells (Low vs. High)

Change of Ts cells (Low vs. High)

Change of B cells (Low vs. High)

Baseline Activated CD8+ cells (Low vs. High)

Change of CD3+ T cells (Low vs. High) Change of CD3+CD4+ T cells (Low vs. High)

Change of CD3+CD8+ T cells (Low vs. High)

Baseline CD3+CD4+ T cells (Low vs. High)

Baseline CD3+CD8+ T cells (Low vs. High)

Distant metastasis (No vs. Yes)

Baseline NK cells (Low vs. High)

Baseline NKT cells (Low vs. High)

Baseline B cells (Low vs. High)

Baseline Ts cells (Low vs. High)

Combined targeted therapy (No vs. Yes)



A

Variables	
Sex (male vs. female)	
Age (d60 vs. >60 years)	
Type of hepatitis (no hepatitis vs. hepatitis B)	
Child-Pugh class (A vs. B)	
AFP (<400 vs. e400 IU/ml)	
BCLC stage (B vs. C)	
ECOG PS (0 vs. 1)	
Line of therapy (first line vs. later lines)	
Combined targeted therapy (No vs. Yes)	
Combined local therapy (No vs. Yes)	
Drinker (No vs. Yes)	
Distant metastasis (No vs. Yes)	
Baseline CD3+ T cells (Low vs. High)	
Baseline CD3+CD4+ T cells (Low vs. High)	
Baseline CD3+CD8+ T cells (Low vs. High)	
Baseline CD4/CD8 ratio (Low vs. High)	
Baseline NK cells (Low vs. High)	
Baseline B cells (Low vs. High)	
Baseline NKT cells (Low vs. High)	
Baseline Ts cells (Low vs. High)	
Baseline Memory T cells (Low vs. High)	
Baseline Activated T cells (Low vs. High)	
Baseline Activated CD8+ cells (Low vs. High)	
Change of CD3+ T cells (Low vs. High)	
Change of CD3+CD4+ T cells (Low vs. High)	
Change of CD3+CD8+ T cells (Low vs. High)	
Change of CD4/CD8 ratio (Low vs. High)	
Change of NK cells (Low vs. High)	
Change of B cells (Low vs. High)	
Change of NKT cells (Low vs. High)	
Change of Ts cells (Low vs. High)	
Change of Memory T cells (Low vs. High)	
Change of Activated T cells (Low vs. High)	
Change of Activated CD8+ cells (Low vs. High)	

В

HR (95% CI) P value 0.926 [0.353,2.428] 0.875 0.667 [0.297,1.499] 0.327 1.006 [0.450,2.252] 0.988 1.082 [0.377,3.106] 0.883 1.773 [0.872,3.608] 0.114 1.680 [0.586,4.818] 0.335 0.767 0.892 [0.419,1.897] 1.717 [0.828,3.561] 0.147 0.059 0.473 [0.217.1.030] 1.089 [0.528,2.247] 0.817 1.114 [0.549,2.262] 0.764 1.520 [0.746,3.099] 0.249 0.778 [0.379,1.598] 0.494 0.592 [0.284,1.237] 0.163 1.136 [0.559,2.307] 0.724 0.775 [0.381,1.576] 0.482 1.246 [0.603,2.573] 0.553 0.910 [0.449,1.842] 0.793 1.186 [0.585,2.408] 0.636 1.130 [0.558,2.291] 0.734 1.146 [0.566,2.321] 0.705 0.651 [0.315,1.345] 0.246 0.721 [0.350,1.487] 0.376 0.959 [0.473,1.942] 0.907 1.318 [0.637.2.728] 0.457 0.549 [0.267.1.129] 0.103 1.095 [0.534,2.245] 0.803 0.423 [0.201,0.890] 0.023 1.443 [0.700,2.975] 0.320 0.889 [0.438,1.806] 0.745 1.299 [0.640,2.639] 0.469 0.610 [0.298, 1.249] 0.177 1.050 [0.518.2.129] 0.892 0.847 [0.417,1.723] 0.647 2 4 ò Hazard ratio (HR)

Figure 3. Univariate Cox regression analysis of progression-free survival (PFS) (Figure 3A) and overall survival (OS) (Figure 3B) for clinical characteristics and peripheral blood lymphocyte subsets. A median cell percentage was employed as the threshold to distinguish between low and high levels.



Discussion

In this study, 61 HCC patients treated with ICI were recruited. The results showed an increased CD3⁺CD8⁺ T cells percentage and a decreased B cells percentage after immunotherapy. Moreover, there were statistically significant differences in the changes in percentages of memory T cells in different immune efficacy groups. In addition, changes in the percentages of activated T cells and activated CD8+T cells after ICI treatment were age-related; changes in the proportions of CD3⁺ T cells and activated CD8⁺ cells were related to hepatitis B infection history; changes in the proportions of B cells and memory T cells were associated with first-line treatment; and changes in the percentages of CD3+CD4+ T cells, the CD4/CD8 ratio and B cells were associated with distant metastasis. Additionally, univariate analysis and survival analysis showed that high percentage changes of NK cells predicted good PFS and OS in ICI-treated HCC patients.

In the development of tumours, tumour cells bind to PD-1 on the surface of T cells by overexpressing PD-L1, inactivating T cells and making them unable to kill tumour cells promptly, resulting in immune escape of tumour cells(6). This study showed that the CD8⁺ T cells percentage increased after ICI treatment. This observation of systemic proliferation of CD8+ T cells after ICI treatment is consistent with the results reported by Fei et al. and Alexander et al.(19, 20). It may be possible that blocking the PD-1/L1 pathway with ICI treatment relieves immunosuppression, restores the viability of CD8⁺ T cells, and induces their activation and proliferation(21). Moreover, blockade of the PD-1/PD-L1 pathway increases B cells activation, proliferation, and immunoglobulin secretion(22). Nevertheless, we found that the percentage of B cells decreased after immunotherapy. It may be that B cells undergo isotype switching upon activation by antigens(19, 23).

According to the expression differences of cell surface molecules, CD4+ T cells were divided into Ts cells (CD4⁺ CD45RA⁺) and memory T cells (CD4⁺ CD45RO⁺)(24, 25). Compared with Ts cells, memory T cells reencounter the same antigen and respond and proliferate faster(26). Furthermore, we analysed the correlation between changes in lymphocyte subset percentages after immunotherapy and clinical efficacy. The results showed statistically significant differences in the changes in the percentages of memory T cells in different immune efficacy groups. This finding suggests that differences in memory T cells responses may contribute to differential immune efficacy. Interestingly, in Peng Yang et al.'s study of the predictive role of the peripheral CD4⁺ memory ratio in non-small cell carcinoma, patients with increased memory CD4⁺ T cells may have a poor PFS (P = 0.138)(27). Memory CD4⁺ T cells may proliferate less than primary reactive cells when they reencounter antigens in peripheral blood and do not elicit a robust immune response(28, 29). This hypothesis requires further exploratory validation.

With age, the body's immunity and immune cells continue to decline, and the ability of the immune system to activate against tumours also decreases(30). We found that the percentages of activated T cells and activated CD8+ cells were inversely correlated with age, which is consistent with previous studies(29, 31). The immunopathological mechanism of hepatitis B infection is mainly T cells immunity, including CD3+ T cells, CD4⁺ T cells, CD8⁺ T cells, and activated cells, in which the immune response to viral antigens is associated with viral elimination and pathogenesis(32, 33). This study showed that patients with a history of hepatitis B infection had decreased percentages of CD3⁺ T cells and increased percentages of activated CD8⁺ T cells. These results suggest that hepatitis B infection causes disturbances in T cells subsets.

Increasing evidence suggests that peripheral blood lymphocyte subsets have predictive and prognostic value in cancer patients(34-36). However, predictive biomarkers for immunotherapy in HCC patients need further exploration. In this study, we showed that a high NK cells percentage predicted a better prognosis in HCC patients. However, none of the other peripheral blood lymphocyte subsets were prognostic factors for PFS and OS in HCC patients with ICI treatment. Therefore, prospective studies are needed to recruit more patients to validate the further association between peripheral blood lymphocyte subsets and PFS and OS.

This study had several limitations. First, its retrospective nature and limited number of patients may have introduced case selection bias and narrow results, and prospective studies are needed to recruit more patients to avoid this bias. Second, whether combined with local therapy, different ICI drugs may be potential confounders. Finally, the follow-up period of this study was a relatively short median of 20.1 months, and future studies will need to extend the observation time and treatment cycles, longitudinally monitor changes in lymphocyte subsets, and further explore their correlation with PFS and OS.

Conclusions

ICI treatment induced changes in the percentages of peripheral blood lymphocyte subpopulations. The changes in the proportion of memory T cells were significantly different among different immune efficacy groups. Significant percentage changes in NK cells were associated with longer PFS and OS. Therefore, peripheral blood lymphocyte subsets may serve as valuable markers for the efficacy and prognosis of immunotherapy in advanced HCC patients.

Supplementary Material

Supplementary information. https://www.jcancer.org/v14p2946s1.pdf

Acknowledgements

Funding

Zhejiang Traditional Chinese Medicine Scientific Research Fund, Category B (Grant 20212B037) and Zhejiang Basic Public Welfare Research Program (Grant LGF22H160084).

Ethics approval and consent to participate

This study was undertaken in accordance with the World Medical Association- Declaration of Helsinki-ethical principles for medical research, and the study was granted ethical approval by the Ethical Committee of the Hospital (IRB-2023-87).

Availability of data and materials

The datasets analyzed during the current study are available from the corresponding author on reasonable request.

Author contributions

Conceived and designed the analysis: Qu Xie, Can Hu; Collected the data: Qu Xie; Contributed data or analysis tools: Cong Luo; Performed the analysis: Qu Xie; Wrote the paper: Qu Xie; Can Hu; Cong Luo.

All authors contributed to manuscript revision, read and approved the submitted version.

Competing Interests

The authors have declared that no competing interest exists.

References

- 1. Arnold M, Ferlay J, van Berge Henegouwen MI, Soerjomataram I. Global burden of oesophageal and gastric cancer by histology and subsite in 2018. Gut. 2020;69(9):1564-71.
- Vogel A, Meyer T, Sapisochin G, Salem R, Saborowski A. Hepatocellular 2. carcinoma. Lancet (London, England). 2022;400(10360):1345-62
- Park JW, Chen M, Colombo M, Roberts LR, Schwartz M, Chen PJ, et al. Global patterns of hepatocellular carcinoma management from diagnosis to death: the BRIDGE Study. Liver international : official journal of the International Association for the Study of the Liver. 2015;35(9):2155-66.
- Allemani C, Matsuda T, Di Carlo V, Harewood R, Matz M, Nikšić M, et al. 4. Global surveillance of trends in cancer survival 2000-14 (CONCORD-3): analysis of individual records for 37 513 025 patients diagnosed with one of 18 cancers from 322 population-based registries in 71 countries. Lancet (London, England). 2018;391(10125):1023-75.
- Llovet JM, Castet F, Heikenwalder M, Maini MK, Mazzaferro V, Pinato DJ, et 5 al. Immunotherapies for hepatocellular carcinoma. Nature reviews Clinical oncology. 2022;19(3):151-72. McNutt M. Cancer in
- immunotherapy. Science (New York, NY). 2013:342(6165):1417.
- 7. Cheng AL, Qin S, Ikeda M, Galle PR, Ducreux M, Kim TY, et al. Updated efficacy and safety data from IMbrave150: Atezolizumab plus bevacizumab sorafenib for unresectable hepatocellular carcinoma. J Hepatol. 2022;76(4):862-73.
- Zhu AX, Finn RS, Edeline J, Cattan S, Ogasawara S, Palmer D, et al. Pembrolizumab in patients with advanced hepatocellular carcinoma previously treated with sorafenib (KEYNOTE-224): a non-randomised, open-label phase 2 trial. The Lancet Oncology. 2018;19(7):940-52.
- 9 Yau T, Park JW, Finn RS, Cheng AL, Mathurin P, Edeline J, et al. Nivolumab versus sorafenib in advanced hepatocellular carcinoma (CheckMate 459); a randomised, multicentre, open-label, phase 3 trial. The Lancet Oncology. 2022;23(1):77-90.
- 10. Ducreux M, Abou-Alfa G, Ren Z, Edeline J, Li Z, Assenat E, et al. O-1 Results from a global phase 2 study of tislelizumab, an investigational PD-1 antibody, in patients with unresectable hepatocellular carcinoma. 2021;32:S217-
- 11. Finn RS, Qin S, Ikeda M, Galle PR, Ducreux M, Kim TY, et al. Atezolizumab plus Bevacizumab in Unresectable Hepatocellular Carcinoma. The New England journal of medicine. 2020;382(20):1894-905.
- Finn RS, Ikeda M, Zhu AX, Sung MW, Baron AD, Kudo M, et al. Phase Ib 12. Study of Lenvatinib Plus Pembrolizumab in Patients With Unresectable Hepatocellular Carcinoma. Journal of clinical oncology : official journal of the American Society of Clinical Oncology. 2020;38(26):2960-70.
- Kudo M, Ikeda M, Motomura K, Okusaka T, Kato N, Dutcus CE, et al. A phase Ib study of lenvatinib (LEN) plus nivolumab (NIV) in patients (pts) with unresectable hepatocellular (uHCC): Study 117. carcinoma 2020;38(4 suppl):513-
- 14. Xu J, Shen J, Gu S, Zhang Y, Wu L, Wu J, et al. Camrelizumab in Combination with Apatinib in Patients with Advanced Hepatocellular Carcinoma (RESCUE): A Nonrandomized, Open-label, Phase II Trial. Clinical cancer research : an official journal of the American Association for Cancer Research. 2021;27(4):1003-11.
- 15. He Y, Lu M, Che J, Chu Q, Zhang P, Chen Y. Biomarkers and Future Perspectives for Hepatocellular Carcinoma Immunotherapy. Front Oncol. 2021;11:716844.
- 16. Pallozzi M, Di Tommaso N, Maccauro V, Santopaolo F, Gasbarrini A, Ponziani FR, et al. Non-Invasive Biomarkers for Immunotherapy in Patients with Hepatocellular Carcinoma: Current Knowledge and Future Perspectives. Cancers. 2022;14(19).
- 17. Hung YP, Shao YY, Lee JM, Hsu C, Hsu CH, Yang MH, et al. Potential of circulating immune cells as biomarkers of nivolumab treatment efficacy for advanced hepatocellular carcinoma. Journal of the Chinese Medical Association : JCMA. 2021;84(2):144-50.
- 18. Hong JY, Cho HJ, Sa JK, Liu X, Ha SY, Lee T, et al. Hepatocellular carcinoma patients with high circulating cytotoxic T cells and intra-tumoral immune signature benefit from pembrolizumab: results from a single-arm phase 2 trial. Genome medicine. 2022;14(1):1.
- 19. Yang F, Xu GL, Huang JT, Yin Y, Xiang W, Zhong BY, et al. Transarterial Chemoembolization Combined With Immune Checkpoint Inhibitors and Tyrosine Kinase Inhibitors for Unresectable Hepatocellular Carcinoma: Efficacy and Systemic Immune Response. Frontiers in immunology. 2022;13:847601.

- 20. Huang AC, Postow MA, Orlowski RJ, Mick R, Bengsch B, Manne S, et al. T-cell invigoration to tumour burden ratio associated with anti-PD-1 response. Nature. 2017;545(7652):60-5.
- Liu B, Hu X, Feng K, Gao R, Xue Z, Zhang S, et al. Temporal single-cell tracing 21. reveals clonal revival and expansion of precursor exhausted T cells during anti-PD-1 therapy in lung cancer. Nature cancer. 2022;3(1):108-21.
- Thibult ML, Mamessier E, Gertner-Dardenne J, Pastor S, Just-Landi S, Xerri L, 22 et al. PD-1 is a novel regulator of human B-cell activation. International immunology. 2013;25(2):129-37.
- 23. Yuan S, Liu Y, Till B, Song Y, Wang Z. Pretreatment Peripheral B Cells Are Associated With Tumor Response to Anti-PD-1-Based Immunotherapy. Frontiers in immunology. 2020;11:563653.
- 24. Hannet I, Erkeller-Yuksel F, Lydyard P, Deneys V, DeBruyère M. Developmental and maturational changes in human blood lymphocyte subpopulations. Immunology today. 1992;13(6):215, 8.
- Poppema S, Lai R, Visser L, Yan XJ. CD45 (leucocyte common antigen) 25. expression in T and B lymphocyte subsets. Leukemia & lymphoma. 1996;20(3-4):217-22.
- Chang JT, Wherry EJ, Goldrath AW. Molecular regulation of effector and 26. memory T cell differentiation. Nature immunology. 2014;15(12):1104-15.
- Yang P, Ma J, Yang X, Li W. Peripheral CD4+ naïve/memory ratio is an 27 independent predictor of survival in non-small cell lung cancer. Oncotarget. 2017;8(48):83650-9
- 28. David A, Crawford F, Garside P, Kappler JW, Marrack P, MacLeod M. Tolerance induction in memory CD4 T cells requires two rounds of antigen-specific activation. Proceedings of the National Academy of Sciences of the United States of America. 2014;111(21):7735-40.
- 29. Klebanoff CA, Scott CD, Leonardi AJ, Yamamoto TN, Cruz AC, Ouyang C, et al. Memory T cell-driven differentiation of naive cells impairs adoptive immunotherapy. The Journal of clinical investigation. 2016;126(1):318-34
- Zhang X, Meng X, Chen Y, Leng SX, Zhang H. The Biology of Aging and 30. Cancer: Frailty, Inflammation, and Immunity. Cancer journal (Sudbury, Mass). 2017;23(4):201-5.
- Xu X, Wang D, Chen W, Li N, Suwinski R, Rossi A, et al. A nomogram model 31. based on peripheral blood lymphocyte subsets to assess the prognosis of non-small cell lung cancer patients treated with immune checkpoint inhibitors. Translational lung cancer research. 2021;10(12):4511-25.
- Trépo C, Chan HL, Lok A. Hepatitis B virus infection. Lancet (London, 32. England). 2014;384(9959):2053-63.
- Shimizu Y. T cell immunopathogenesis and immunotherapeutic strategies for 33. chronic hepatitis B virus infection. World journal of gastroenterology. 2012:18(20):2443-51.
- Yang J, Xu J, E Y, Sun T. Predictive and prognostic value of circulating blood 34. lymphocyte subsets in metastatic breast cancer. Cancer medicine. 2019;8(2):492-500.
- Yan Y, Wang X, Liu C, Jia J. Association of lymphocyte subsets with efficacy 35. and prognosis of immune checkpoint inhibitor therapy in advanced non-small cell lung carcinoma: a retrospective study. BMC pulmonary medicine. 2022;22(1):166.
- Liu A, Xia Y, Li W, Zhang G, Liu Y, Ye S, et al. The Predictive Value of 36. Changes in the Absolute Counts of Peripheral Lymphocyte Subsets for Progression and Prognosis in Breast Cancer Patients. Contrast media & molecular imaging. 2022;2022:3444360.