Figure S1. Expression levels of CXCR4 detected in lung adenocarcinoma tissues. (A)
Western blot analysis was used to detect expression levels of CXCR4. Detection of actin served as a loading control. The sample number labels shown in black and red represent samples from patients with and without lymph node metastasis, respectively.
(B) Quantitation of CXCR4 levels (median, [P₂₅-P₇₅]) according to metastasis.

Figure S2. Validation of a PC-9 stable cell line expressing low levels of CXCR4. (A) PCR amplification of CXCR4-shRNA cassettes in: (1) negative control (ddH₂O); (2) DNA marker; and (3-8) CXCR4-shRNA clones 3-8 samples. (B) Sequencing of a representative PCR product from A. (C) Detection of *CXCR4* mRNA in the PC-9/CXCR4 stable cell line with and without shRNA silencing as indicated (*P < 0.01). (D) Cell morphology (magnification, 100×) was observed by microscope and confocal microscope. GFP imaging was performed to confirm expression of the infecting lentiviruses. (E) Western blotting was performed to detect expression of CXCR4 in the cell lines indicated. Detection of GAPDH was included as a loading control. (F) Expression of *CXCR4* mRNA was detected in RT-qPCR assays of the indicated cell groups. PC9-KD: PC-9/CXCR4-shRNA1 group; PC9-C: PC-9/shCtrl group; PC9: untransfected PC-9 group. * P < 0.001.

Figure S3. Effect of *CXCR4* silencing on various phenotypes of the PC14 cell line. Low levels of CXCR4 expression were compared with higher levels of CXCR4 expression in: (A) soft agar colony formation assays, (B) wound healing assays, and (C) transwell assays. These assays were performed to evaluate colony formation, cell migration, and cell invasion, for the PC14 cell groups indicated, respectively. In panel C, crystal violet stained cells (at left) and the corresponding quantitation of these stained cells (at right) indicate the number of invasion cells for each group. PC14-KD: PC14/CXCR4-shRNA1 group; PC14-C: PC14/shCtrl group; PC14: untransfected PC14 group.

Table S1. ShRNA sequences for targeting CXCR4

Table S2. Sequences of the primers used in RT-qPCR assays.

Table S3. CXCR4-related circRNAs identified in database search

Figure S1



Figure S2







Table S1. ShRNA sequences for targeting CXCR4

Primer name	Orientation	Sequence $(5' \rightarrow 3')$
shRNA1	Forward	gatccGGATCAGCATCGACTCCTTTTCAAGAGAAAGGAGTCGATGCTGATCCTTTTTTg
	Reverse	aattcAAAAAAGGATCAGCATCGACTCCTTTCTCTTGAAAAGGAGTCGATGCTGATCCg
shRNA2	Forward	gatccGGATCAGTATATACACTTCTTCAAGAGAGAGAGTGTATATACTGATCCTTTTTTg
	Reverse	aattcAAAAAAGGATCAGTATATACACTTCTCTCTTGAAGAAGTGTATATACTGATCCg
shRNA3	Forward	gatccGCAAGGCAGTCCATGTCATTTCAAGAGAATGACATGGACTGCCTTGCTTTTTTg
	Reverse	
shRNAc	Forward	ccggtGCTTCGACATTTAACCAATTTCAAGAGAATTGGTTAAATGTCGAAGCTTTTTTg

	Gene		Sequence $(5' \rightarrow 3')$	
_		Forward	GGAGAGTTGTAGGATTCTAC	
	CACK4	Reverse	CCTCGGTGTAGTTATCTGAAG	
		Forward	GGCGATGCTGGCGCTGAGTAC	
	GAPDH	Reverse	GAGGCTGTTGTCATACTTCTC	
		Forward	TTTTGCCTCAGAGCATACCT	
	hsa_circRNA_0056616	Reverse	GTCTTTGTTCTTTACTTCTCCCA	

Table S2. Sequences of the primers used in qRT-PCR assays.

Table S3. CXCR4-related circRNAs identified in databased
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CircRNA ID	Genomic length (bp)	Spliced length (bp)	Scores	Repeats	Annotation
		(*P)	200100	1000000	
has_circ_0056615	3389	3389	NA	NA	ALT_ACCEPTOR.CDS.coding, OVCOED, OVEXON, upstream_start, UTR3, UTR5
					ANNOTATED.CDS.coding, OVCOED, OVEXON, UTR3,
has_circ_0056616	3807	1674	NA	NA	UTR5
has_circ_0117403	253	253	3	NA	ALT_ACCEPTOR.ALT_DONOR., CDS.coding, INTERNAL, OVCOED, OVEXON, UTR3
NA: not available					