

**Research Paper** 



## Runs of homozygosity associate with decreased risks of lung cancer in never-smoking East Asian females

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#### Abstract

Although genome-wide association studies (GWASs) have identified some risk single-nucleotide polymorphisms in East Asian never-smoking females, the unexplained missing heritability is still required to be investigated. Runs of homozygosity (ROHs) are thought to be a type of genetic variation acting on human complex traits and diseases. We detected ROHs in 8,881 East Asian never-smoking women. The summed ROHs were used to fit a logistic regression model which noteworthily revealed a significant association between ROHs and the decreased risk of lung cancer (P < 0.05). We identified 4 common ROHs regions located at 2p22.1, which were significantly associated with decreased risk of lung cancer ( $P = 2.00 \times 10^{-4} - 1.35 \times 10^{-4}$ ). Functional annotation was conducted to investigate the regulatory function of ROHs. The common ROHs were overlapped with potential regulatory elements, such as active epigenome elements and chromatin states in lung-derived cell lines. *SOS1* and *ARHGEF33* were significantly up-regulated as the putative target genes of the identified ROHs in lung cancer samples according to the analysis of differently expressed genes. Our results suggest that ROHs could act as recessive contributing factors and regulatory elements to influence the risk of lung cancer in never-smoking East Asian females.

Key words: lung cancer, runs of homozygosity, GWASs, genetic risk factors, regulatory elements

## Introduction

Lung cancer is a major public health problem worldwide and constitutes an enormous burden on global society [1]. Epidemiological studies of lung cancer have shown that the highest incidence rates among females occur in North America, Europe, Australia and East Asia [2]. The highest incidence rate among females occurs in Northern America, which is 33.8 cases per 100,000. For the East Asian females, the incidence rate is 19.2 cases per 100,000 in average. Moreover, lung cancer is the most commonly diagnosed cancer among females in China and North Korea [3]. Although most of lung cancers are attributed to tobacco smoking, genetic factors also play a pivotal role in lung cancer development [3]. Heritability of lung cancer has been estimated to be 31% [4]. Moreover, it is now fairly accepted that lung cancer occurring in lifetime never-smokers is distinct from smoking-associated lung cancer [5]. Compared to never-smokers, the risk of developing lung cancer is 20-40 times higher in lifetime smokers [6]. Furthermore, it has been reported that there are different clinical features and outcomes of lung cancer between never-smokers and smokers. Several studies have reported a higher proportion of adenocarcinoma histology in never-smokers with lung cancer compared to smokers [5, 7]. Never-smokers with lung cancer have better response rates to epidermal growth factor receptor (EGFR) tyrosine kinase inhibitors than smokers [2, 8]. Identifying genetic factors in never-smoking females could exclude environmental confounding risk factors and offer new insights into the progression of lung cancer.

association studies Previous genome-wide (GWASs) have identified over 70 susceptibility single nucleotide polymorphisms (SNPs) associated with lung cancer (http://www.genome.gov/gwastudies), including some SNPs identified in never-smoking females in Asia. For instance, susceptibility loci at 3q28, 5p15.33, 6p21.1, 6p21.32, 6q22.2, 9p21, 10q25.2, 12q13.13 and 17q24.3 are reported to be associated with the risk of lung cancer in never-smoking Asian females [9-13]. However, the susceptibility loci identified by GWASs only accounted for about 10% of the total heritability in Asian populations, remaining a large part of the heritability to be interpreted [14]. Therefore, studies with innovative methodologies are needed to detect other genetic factors.

Runs of homozygosity (ROHs) are referred to as a consecutively homozygous segment with large numbers of SNPs along chromosomes. ROHs can represent as a new type of genetic variation since ROHs varies among individuals and populations [15-17]. With the advancement of genome-wide SNP array, genome-wide homozygosity can be assessed conveniently using high-density SNPs data [18]. ROHs can reflect the level of inbreeding and reveal non-additive genetic effects, hence complex traits and diseases could be influenced by ROHs and corresponding recessive genetic effects. Several studies have reported the associations between ROHs and complex traits or diseases, including height, bone mineral density, Alzheimer's disease and thyroid cancer [17, 19-21]. In the investigation of the association between ROHs and the risk of lung cancer, Cheng Wang et al. found that the ROHs level was negatively related to the risk of lung cancer, and a ROHs region at 14q23.1 was associated with the risk of lung cancer in Han Chinese population [22]. This study used population comprising both males and females. In East Asia, the incidence rates of lung cancer in males are much higher than in females, suggesting the gender difference in lung cancer risks [23]. Besides, smoking and never-smoking individuals were used in this study at the same time, it was difficult to examine whether the identified genetic variants were associated with lung carcinogenesis or nicotine addiction [24]. Therefore, the investigation of the effects of ROHs in never-smoking females with a larger sample size may enhance the understanding of the relationship between ROHs and lung cancer.

In this study, we screened ROHs on

whole-genome regions in 8,881 never-smoking East Asian females and filtered the common ROHs regions by statistical methods. With the purpose of examining underlying function of ROHs, we annotated these common ROHs regions with located genes and investigated their effects on neighbor genes. We also investigated the underlying regulatory function of the common ROHs regions with annotation using markers, regulatory elements and epigenetics We long-range interaction data. identified differentially expressed genes (DEGs) between lung adenocarcinoma samples and control samples to evaluate the expression level of genes possibly regulated by ROHs. Our results reveal that ROHs play an important role as recessive genetic factors and act as regulatory elements in the underlying genetic mechanism of lung cancer.

## Materials and Methods

## Subjects

A total of 8,881 never-smoking East Asian females were enrolled in this study, including 4,922 (55.42%) lung cancer cases and 3,959 (44.58%) controls from China, South Korea, Japan, Singapore, Taiwan, and Hong Kong. The mean age of the subjects was 58.2 years old. The diagnoses of all cases were confirmed histologically. All subjects had complete phenotypes, including age, sex, and histological type of lung cancer. The basic characteristics of the subjects are shown in Table 1. The phenotype and genotype data were obtained from Database of Genotypes and (dbGaP). data Phenotypes The we used (phs000716.v1.p1) have passed the embargo date [25]. All samples used in this study have been reported in Lan et al [25].

Table 1. Basic characteristics of the study samples

Characteristic	Never-smoking East Asian females GSE40791				
Sample Size (n)	8,881	194			
Age (year)	58.2(9.29)	68.01(10.46)			
Female (%)	100%	42.8%			
Cases (%)	55.42%	48.45%			
Never-smokers (%)	100.00%	7.22%			
Histology					
Adenocarcinoma (%)	73.04%	100%			
Squamous (%)	13.41%	0%			
Other (%)	13.55%	0%			

Age values are presented as mean (standard deviation).

## Genotyping and quality control

Samples were genotyped using two similar high-density SNP arrays (Illumina 610Q SNP microarray and Illumina 660W SNP microarray). The intersection of SNPs on both arrays exceeded 570,000 SNPs. The genotyping data passed the quality control (QC) with measurements have been provided by the previous GWASs [25]. The individuals with over 2% missing rate were excluded. SNPs with call rate less than 95% or minor allele frequency (MAF) less than 5% were excluded. Only autosomal SNPs were analyzed in this study. After QC, 4,922 cases, 3,959 controls and 420,680 autosomal SNPs were remained for subsequent analyses. We estimated the proportion of phenotypic variance explained by SNPs among all of the 8,881 individuals using a software called genome-wide complex trait analysis (GCTA) [26], with the estimated prevalence of lung cancer in East Asian females as 19.2 cases per 100,000 (Globocan 2012 http://globocan.iarc.fr/Default.aspx).

### Identification of ROHs

In this study, ROHs were defined as segments with at least 50 consecutive homozygous SNPs and ROHs calling were performed using PLINK v1.07 (http://pngu.mgh.harvard.edu/purcell/plink/) [27]. We set a range of the minimum length for calling ROHs segments, including 0.50 Mb, 1.00 Mb, 1.25 Mb, 1.50 Mb, 1.75 Mb, 2.00 Mb and 3.00 Mb. At the same time, ROHs detected in this study must have more than one SNP per 50 kb, and the gap between two adjacent ROHs segments was set more than 1.00 Mb. PLINK used a sliding window of 5.00 Mb with at least 50 SNPs to define ROHs. One heterozygous site and five missing calls in a window were allowed.

## Association analyses between ROHs and lung cancer

A logistic regression model was used to examine the association between the summed ROHs and lung cancer. Age and three significant principal components (EV1, EV2, and EV4) obtained from the software GCTA [26] were used as covariates. The summed ROHs were defined as the total length of ROHs in an individual. After fitting the logistic regression model,  $F_{ROHs}$  and  $\beta_{FROHs}$  were used to examine the degree of homozygosity and the effects of ROHs on phenotype. FROH was defined as the ratio of the summed ROHs to the total genome length (approximately 2.5 × 10<sup>6</sup>) and  $\beta_{\text{FROHs}}$  was the estimated effect of F<sub>ROHs</sub> on the trait calculated by  $F_{ROHs}$  divided by the standard deviation of the summed ROHs [28].

## The common ROHs regions selection

The "--homozyg-group" option of PLINK was used to produce files consisted of the overlapping ROHs regions which were divided into pools including the ratio of cases and controls containing the same overlapping ROHs regions. Although ROHs regions were defined with parameters described above, some overlapping regions were small because of little common segments in different individuals, e.g. only including one SNP. Such regions were excluded from the subsequent annotation. Overlapping ROHs regions contained in less than 5% individuals were also excluded from subsequent analyses. The chi-squared goodness-of-fit test was then performed to detect the common regions, which were defined as the overlapping ROHs regions with significant different ratios in cases and controls. The Benjamini and Hochberg (BH) procedure was used for multiple-testing corrections.

# Inbreeding coefficients calculation and statistical analysis

We used PLINK to get the genomic inbreeding coefficients estimated (F) [27]. The inbreeding coefficients were calculated from all the homozygous SNPs counts on each autosome. The differences between cases and controls were tested using the Student's t-test on whole-genome and a single chromosome separately. We fitted a logistic regression model with inbreeding coefficients and the covariates on the case-control status of lung cancer. The covariates were the same as in the previous regression model, including age and three principal components (EV1, EV2, and EV4).

## Testing the effects of natural selection

We used haplotter (http://haplotter.uchicago. edu/) to evaluate the effects of natural selection on the common ROHs [29]. Haplotter is a tool that can detect positive selection in a genomic region using the HapMap Phase II data. We estimated the integrated haplotype score (iHS), Fay and Wu's H and fixation index (F<sub>st</sub>) for ROHs separately. The value of iHS was used to detect signals of the recent selection through scanning SNPs data at the whole genome. Voight *et al.* reported that SNPs with |iHS| > 2 indicated a powerful signal of selection [29]. Fay and Wu's H is powerful to detect positive selection based on the frequencies of the polymorphisms in the region [30]. F<sub>st</sub> can be used to measure the level of differentiation at the locus between populations due to selection [31].

## Functional annotation of the common ROHs regions

To evaluate the underlying regulatory function of the common regions, we annotated the common ROHs regions in different levels. First, we identified genes located in the common ROHs regions using ANNOVAR [32]. The common ROHs regions were also annotated for histone markers in A549 cell line by using the ChIP-seq data from ENCODE in the UCSC genome database [33]. To assess the chromatin states of ROHs regions, we downloaded 15-state chromatin state segmentation data generated by ChromHMM based on the Roadmap histone modification data [34]. We identified the common ROHs regions within putative enhancer regions, including "Enhancers" and "Genic enhancers". We annotated the chromatin states of the common ROHs regions in A549 cells, IMR90 cells and normal lung tissue cells, separately. To detect the regulatory function of ROHs regions through long-range interaction, we used the Hi-C data downloaded from 4D genome (http://4dgenome.research.chop.edu/) [35]. We only annotated the Hi-C regions in IMR90 because high-quality Hi-C data from other lung cell lines are still unavailable. We identified long-range interaction regulated genes which overlapped with transcription start sites (including "Active TSS" and "Flanking TSS" according to 15-state chromatin state segmentation annotation) lied within long-range interaction pairs of enhancer regions.

#### Gene expression profiling of lung tumors

DEGs from lung adenocarcinoma tissues were identified from a publicly available dataset (GEO accession number: GSE40791). The dataset consists of 94 lung adenocarcinoma samples and 100 normal controls. Basic information of GSE40791 samples was summarized in Table 1. The age row of GSE40791 samples refers to the age at surgery. The gene expression data were generated on Affymetrix human genome u133 plus 2.0 arrays. Before statistical analyses, the gene expression data were normalized by using the Robust Multi-array Average (RMA) method [36]. The conditions of differential expression were calculated by using the unpaired t-test. The significant cutoff was set as *P*-value < 0.05.

## **Results**

#### Estimates of heritability explained by SNPs

We estimated the heritability with GWAS data in 8,881 unrelated individuals. The SNPs explained on average 18.68% (s.d. 2.57%) of the proportion of the total phenotypic variance, which was approximate to the result of previous published studies. Sampson et al. estimated that the heritability of the lung cancer in Asian population was 12.1% (95 % CI = 6.40% to 17.7%) based on the GWAS dataset [37].

## Identification of ROHs on autosomal chromosomes in subjects

We identified total ROHs regions in all 8,881 individuals with different minimum length thresholds. Summary statistics of the total ROHs size and the total ROHs number are shown in Table 2. The total number and the total size of ROHs in individuals decreased with the minimum ROHs size threshold increased. The mean size per ROHs became longer with the minimum length increased except the threshold of ROHs size of 3 Mb. When the threshold of the minimum length for detecting ROHs was set above 1.50 Mb, the lengths of ROHs which carried by some individuals were shorter than the set threshold (Table 2). Using 1.5 Mb as the minimum threshold of ROHs length, 99 % of cases and 99 % of controls were detected with more than 1 ROHs. When the threshold was set as 3 Mb, we detected 4 % of cases and 6 % of controls carrying one or more ROHs.

**Table 2.** Summary of ROHs Characteristics identified withdifferent length

Total ROHs size			Total ROHs number			Mean size
	Range			Range		
Mean(kb)	Min(kb)	Max(kb)	Mean(n)	Min(n)	Max(n)	(kb)
148,270	58,436	445,666	176.90	75	272	837.8
51,936	13,854	378,231	34.79	11	115	1,471
32,633	2,769	358,867	17.39	2	81	1,808
21,650	-	350,432	9.32	-	58	2,153
14,884	-	343,937	5.12	-	44	2,513
10,938	-	334,826	2.99	-	33	2,760
5,947	-	335,623	0.84	-	25	2,185
	Total ROF Mean(kb) 148,270 51,936 32,633 21,650 14,884 10,938 5,947	Total ROUs size           Range           Mean(kb)         Min(kb)           148,270         58,436           51,936         13,854           32,633         2,769           21,650         -           14,884         -           10,938         -           5,947         -	Fordal ROF size           Range           Min(kb)         Max(kb)           148,270         58,436         445,666           51,936         13,854         378,231           32,633         2,769         358,867           21,650         -         350,432           14,884         -         343,937           10,938         -         34,826           5,947         -         355,623	Total ROHs size         Total RO           Range         Mara(k)         Mara(k)         Mean(n)           148,270         58,436         445,666         176.90           51,936         13,854         378,231         34.79           32,633         2,769         358,867         17.39           21,650         -         350,432         9.32           14,884         -         343,937         5.12           10,938         -         334,826         2.99           5,947         -         335,623         0.84	Total ROFF size         Total ROFF size         Total ROFF size           Range         Range         Range           Mean(b)         Min(kb)         Max(kb)         Mean(n)         Min(n)           148,270         58,436         445,666         176.90         75           51,936         13,854         378,231         34.79         11           32,633         2,769         358,867         17.39         2           21,650         -         350,432         9.32         -           14,884         -         343,937         5.12         -           10,938         -         334,826         2.99         -           5,947         -         335,623         0.84         -	Total ROHs size         Total ROHs number           Range         Range         Range           Mean(h)         Max(h)         Mean(h)         Man(n)         Max(n)           148,270         58,436         445,666         176.90         75         272           51,936         13,854         378,231         34.79         11         115           32,633         2,769         358,867         17.39         2         81           21,650         -         350,432         9.32         -         58           14,884         -         343,937         5.12         -         44           10,938         -         334,826         2.99         -         33           5,947         -         335,623         0.84         -         25

Size: the minimum threshold for calling ROHs segments. The mean and range data were calculated by adding together all the values for the sizes of ROHs in each individual.

## Associations between ROHs and lung cancer risk

To examine whether the total length of ROHs per individual between cases and controls could affect case-control status, we performed a logistic regression analysis. The values of  $\beta_{\text{FROHs}}$  were all less than zero (ranging from -11.09 to -12.20). Therefore, we found that the increased level of ROHs was strongly associated with the decreased lung cancer risk under all of the different lengths conditions (P < 0.05) (Table 3).

Table 3. Effects of genome-wide burden of ROHs on subjects

Size	Р	$\beta_{FROH}$	$eta_{{\scriptscriptstyle FROH-se}}$	
> 500 kb	0.007	-11.50	4.24	
> 1.00 Mb	0.010	-11.09	4.33	
> 1.25 Mb	0.007	-11.81	4.38	
> 1.50 Mb	0.009	-11.58	4.42	
> 1.75 Mb	0.006	-12.20	4.48	
> 2.00 Mb	0.007	-12.13	4.52	
> 3.00 Mb	0.009	-12.08	4.65	

*P*: P-value for association with the logistic regression model;  $\beta$ : the estimated effects of ROHs on whole genome in units of standard deviations; se: standard error.

#### Identification of common ROHs regions

We identified ROH\_14857 ( $P = 1.43 \times 10^4$ , adjusted P = 0.044), ROH\_14715 ( $P = 1.41 \times 10^4$ , adjusted P = 0.044), ROH\_14344 ( $P = 1.35 \times 10^4$ , adjusted P = 0.044) and ROH\_14342 ( $P = 2.00 \times 10^4$ ,

adjusted P = 0.045) as the common ROHs regions. These regions were used in the subsequent analyses. Consistent with results of the logistic model (Table 4), the number of controls which had the common ROHs regions were more than the number of cases. The frequencies of the common ROHs regions were over 0.01 within the subjects (range from 0.048 to 0.050). Notably, we found that all of the common ROHs regions were located in chromosome 2. Each region contains one or two protein-coding genes (*DHX57*, *MORN2*, *ARHGEF33* and *SOS1*). Genes with exonic regions overlapped with these common ROHs regions are also listed in Table 4.

### Measurement and association between Inbreeding coefficient and lung cancer

We calculated the inbreeding coefficient (F) using SNPs information of all samples. We found a significant difference of F between cases and controls on whole-genome level (P-value =  $8.65 \times 10^{-4}$ ). The means and standard deviations (SDs) for F in cases and controls were 0.0004 (0.1277) and 0.0110 (0.1087), separately. Furthermore, we calculated F for each chromosome separately. As shown in Table 5, chromosomes 2, 6, 8 and 16 showed the significant differences in the three inbreeding coefficients (P-value = 0.0026, 0.044, 0.0004 and 0.0369, separately). All of the common ROHs regions were located in chromosome 2. After fitting a logistic regression model with F, F also showed significant P-value of 0.0087 with a negative estimate of effect as -4.774.

Likewise,  $F_{ROH}$  was also higher in controls than cases. The means and standard deviations (SDs) for  $F_{ROH}$  were 0.0204 (0.0094) in cases and 0.0211 (0.0109) in controls. We detected a significant difference for  $F_{ROH}$  between cases and controls (P-value = 0.0013). Moreover, the inbreeding coefficient and  $F_{ROH}$  were significantly associated with each other according to Pearson's correlation coefficient (r = 0.7517, P-value =  $2.2 \times 10^{-16}$ ).

## Natural selection on the common ROHs

We used iHS, Fay and Wu's H test and Fst to

measure the selective pressure of ROHs. In the East Asian population, ROH\_14715 and ROH\_14342 showed extremely positive iHS scores (iHS > 2) of 2.623 and 2.010, respectively. The Fay and Wu's H test scores for all four ROHs were extremely negative and less than -10 (from -56.961 to -22.097). According to the Fst test, the Fst values of the ROHs were derived as 0. 360, 0. 433, 0. 290, and 0. 271, respectively. Based on the threshold of Fst > 0.2 used by Thomsen *et al* [17], we found that all of the ROHs were different between Yoruba and East Asian populations with the Fst values.

# Regulatory functional annotation of the common ROHs regions

We annotated the common ROHs regions with active histone markers in A549 cell line, including H3K27ac, H3K4me1 and H3K4me2. As shown in Figure 1, ROH 14342, ROH 14344 and ROH 14715 were enriched in a set of activated enhancer histone modifications in A549 cell line, including H3K27ac, H3K4me1 and H3K4me2. Therefore, these 3 ROHs regions were regarded as enhancers in A549 cell line according to the chromatin states annotation (Figure 1). In IMR90 cell line, ROH\_14342 and ROH\_14857 were also annotated as enhancers. The results of the common ROHs regions annotated with chromatin states and long-range interactions are shown in Figure 2. We also explored the effects of ROHs on genes as distal regulatory elements. In IMR90 cell line, we found that the active promoter regions of SOS1 and ARHGEF33 were the target regions of enhancer within ROH 14342.

## **DEGs of the common ROHs regions**

We evaluated the expression levels for the four genes located in the common ROHs in lung cancer samples. The gene expression data of 100 normal samples and 94 lung adenocarcinoma samples were obtained by using microarray. We found that *SOS1* (P =  $1.21 \times 10^{-8}$ ) and *ARHGEF33* (P =  $3.25 \times 10^{-4}$ ) were significantly up-regulated in lung cancer samples.

 Table 4. Basic characteristics of significant ROHs associated with lung cancer

ROH	Chr	Position		Position		Freq	Cases:	Chi <sup>2</sup>	Р	$P_{ad}$	Located gene	iHS <sub>max</sub>	Fay and Wu's Fst max	
		Start	End		controls						H max			
ROH_14857	2	39025631	39047154	0.048	198:230	14.47	1.43×10-4	0.044	DHX57	1.284	-22.097	0.360		
ROH_14715	2	39084756	39135927	0.049	200:232	14.49	$1.41 \times 10^{-4}$	0.044	DHX57, MORN2	2.623	-36.364	0.433		
ROH_14344	2	39198965	39226271	0.049	206:238	14.57	1.35×10-4	0.044	ARHGEF33, SOS1	1.798	-39.925	0.290		
ROH_14342	2	39247288	39293530	0.050	207:237	13.85	2.00×10-4	0.044	SOS1	2.010	-56.961	0.271		

Freq: frequency; Chr: chromosome; Chromosomal positions are shown according to NCBI Build 37 (hg19); P: P-value for testing the differences of homozygosity status between cases and controls with two-sided chi-square test; *P<sub>ad</sub>*: P-value adjusting with the Benjamini and Hochberg (BH) procedure; Located gene: ROHs located gene; iHS max; the maximal absolute values for iHS; Fay and Wu's H max: the maximal absolute values for Fay and Wu's H; and Fst max: the maximal absolute values for Fst; iHS max, Fay and Wu's H max and Fst max were derived for Asian population from Haplotter (http://haplotter.uchicago.edu/) using Phase II HapMap Project data.

## Discussion

In this study, we screened whole-genome ROHs in 8,881 never-smoking East Asian females. We found that low level of ROHs was associated with the risk of lung cancer. We successfully identified four ROHs significantly associated with lung cancer. Our study reveals that ROHs might be involved in the development of lung cancer through affecting gene expression in never-smoking East Asian females.

In this study, we used two indicators containing the total length and the numbers of ROHs to reveal the level of homozygosity in an individual from different perspectives. According to the statistical summaries, most of the ROHs regions in the subjects were relatively short. These results indicate that homozygosity in the subjects is more likely to be the consequence of selective pressure compared with inbreeding [38]. In other words, ROHs in the samples inherited from common ancestors are improbable. Consistent with the previous analyses, our results support the conclusion that the ROHs might be the consequences of selection. Some studies have reported that consanguineous parents are more likely to generate long ROHs [39, 40]. In contrast, short ROHs are supposed to be a risk factor of complex traits and diseases [41]. Therefore, in this study, ROHs could present as a type of genetics variant in the population, and contribute genetics effects to the risk of lung cancer. The distribution of total length and the numbers of ROHs in this study are consistent with the results of previous studies that detected ROHs in East Asian populations under the same minimum length [28, 42, 43]. For instance, at the minimum length of ROHs >1.50 Mb, we found that the mean size of total ROHs was 21.65 Mb, while Joshi *et al.* reported that the size of ROHs ranged from 20.00 Mb to 39.00 Mb in diverse East Asian populations [28].

 Table 5. P-values for the statistical analyses of inbreeding coefficients between cases and controls

Chromosome	Total length	FI
1	249250621	0.088905
2	243199373	0.002689
3	198022430	0.717751
4	191154276	0.575738
5	180915260	0.171943
6	171115067	0.043707
7	159138663	0.489741
8	146364022	0.000393
9	141213431	0.207553
10	135534747	0.163951
11	135006516	0.223446
12	133851895	0.599272
13	115169878	0.985238
14	107349540	0.543299
15	102531392	0.151867
16	90354753	0.036879
17	81195210	0.75997
18	78077248	0.282267
19	59128983	0.303959
20	63025520	0.561602
21	48129895	0.995754
22	51304566	0.537312

\*bold values represent significant differences between cases and controls at P < 0.05



Figure 1. Functional annotation for the common ROHs regions. The common ROHs regions overlapped with histone marks which classified as active enhancer in A549 cell line, including H3k4me1, H3k4me2 and H3k27ac, and annotated as enhancers according to 15-state chromatin state segmentation.

We found that the risk of lung cancer decreased with the increasing homozygosity in East Asian females. Our results are consistent with the results of a previous study [22]. Because we did not detect ROHs in some individuals with the threshold of minimum length at 1.25 Mb, we considered that the results of chi-square test were meaningful when the minimum length is below 1.25 Mb. However, defining ROHs with shorter length may overestimate the true level of homozygosity in individual genome [15]. Several studies have indicated that outbred populations usually carry ROHs with the length of more than 1.00 Mb [39]. Considering most of previous studies defined ROHs with a minimum length of 1.00 Mb to balance those factors, we also chose 1.00 Mb as length of ROHs regions to explore their functions.

We identified four ROHs significantly associated with the risk of lung cancer. The proportion of ROHs in cases and controls indicated that more controls had these four ROHs. For example, we detected ROH\_14857 in 198 cases and 230 controls. The chi-square test showed that the distribution of the



**Figure 2. Regulatory annotation of the common ROHs regions in IMR90 cell line.** ROH\_14342 and ROH\_14857 were annotated with enhancers and interactions within the chromosome regions in IMR90 cell line. *SOS1* and *ARHGEF33* with active promoters were the target genes of the enhancer region overlapped with ROH\_14342. Status of the enhancer regions were annotated according to 15-state chromatin state segmentation data generated by ChromHMM based on the Roadmap histone modification data. The interactions between regions on the chromosome detected by chromatin capture Hi-C are shown in the center.

ROHs was significantly different between cases and controls. Therefore, these consequences and the results of logistic regression pointed out the same conclusion that the increased level of ROHs was strongly associated with the decreased risk of lung cancer. Furthermore, all the identified common ROHs regions located in chromosome 2p22.1. Lui et al. detected the high level of amplification in small cell lung samples at chromosome 2p22 [44]. According to Yan et al., gains of chromosome 2p were associated with advanced clinical stage and metastases of lung squamous cell carcinomas [45]. Pifarre et al. found that chromosome 2p25-p22 was frequent targets for replication errors (RER) in lung cancer, and RER-positive tumors were correlated with worse survival [46]. Therefore, the increased ROHs might suggest the deletion of heterozygosity mutation, and consequently lead to the activation of tumor suppressor genes and/or the inactivation of oncogene located in this region. We found that these four ROHs located at 2p22.1 overlapped with several genes, including SOS1, ARHGEF33, DHX57, and MORN2.

> The ROHs at 14q23.1 reported by Wang *et al.*[47] was also detected in our study. However, there was no significant association between it and lung cancer after multipletesting corrections.

> Intriguingly, we found that the common ROHs regions also had potential regulatory function to distal genes. As the results of chromatin state segmentation annotation in IMR90 cell line and A549 cell line, ROH\_14342 was annotated as enhancers in these two cell lines. These results suggest that ROHs can be distal regulatory elements.

> SOS1 and ARHGEF33 were potentially regulated by ROH 14342 according to the results of long-range interaction annotation and DEGs. SOS1 is a famous guanine nucleotide-exchange factor, acting in directing exchange of RAS-GDP to RAS-GTP and leading to the ERK activation[48]. Sequentially, the activation of RAS-RAF-MEK-ERK-MAP kinase pathway accelerates tumor cell proliferation [49, 50]. Several studies found that the reduction in SOS1 expression and corresponding RAS-MAPK activity depression could be great

importance in anticancer activity [51-53]. ARHGEF33 has been identified as the gene for cell differentiation, of cell primary function survival and caspase-dependent cell death pathway. Considering ROHs are associated with lower risk of lung cancer, we suggest that the specific ROH regions could display as inhibitors to down-regulate the cell proliferation related genes. These results provide the new evidence that more homozygous levels inhibit the expression of cancer-associated genes and consequently lower the risk of lung cancer.

There are some potential limitations of our study. Some other genetic factors which could influence the detection of ROHs such as one-copy deletion were not able to be definitely excluded [54]. Therefore, we used five missing calls in one window when calling ROHs. That reduced the absence of one allele in genotype calling affecting in ROHs identification. More explorations of the common ROHs regions require more details of sequencing and chromosome conformation capture techniques.

In conclusion, our study provides evidence that ROHs could act as recessive contributing factors and regulatory elements for the risk of lung cancer, which offers a new insight for discovering missing heritability of lung cancer. For confirming and illuminating the potential mechanism of lung cancer, further molecular studies would be demanded.

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#### **Competing Interests**

The authors have declared that no competing interest exists.

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