

Review

Gene Expression Detection Assay for Cancer Clinical Use

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Abstract

Cancer is a genetic disease where genetic variations cause abnormally functioning genes that appear to alter expression. Proteins, the final products of gene expression, determine the phenotypes and biological processes. Therefore, detecting gene expression levels can be used for cancer diagnosis, prognosis, and treatment prediction in a clinical setting. In this review, we investigated six gene expression assay systems (qRT-PCR, DNA microarray, nCounter, RNA-Seq, FISH, and tissue microarray) that are currently being used in clinical cancer studies. Some of these methods are also commonly used in a modified way; for example, detection of DNA content or protein expression. Herein, we discuss their principles, sample preparation, design, quantification and sensitivity, data analysis, time for sample preparation and processing, and cost. We also compared these methods according to their sample selection, particularly for the feasibility of using formalin-fixed paraffin-embedded (FFPE) samples, which are routinely archived for clinical cancer studies. We intend to provide a guideline for choosing an assay method with respect to its oncological applications in a clinical setting.

Key words: gene expression, assay, cancer, clinical use, qRT-PCR, DNA microarray, nCounter, RNA-Seq, FISH, tissue microarray

Introduction

It is generally accepted that driver gene mutations initialize cancer development. Proto-oncogenes, such as *Ras*, are transcribed into products such as receptors, growth factors, transcription factors, and signaling enzymes for cellular proliferation.¹ Gain-of-function mutations in proto-oncogenes, resulting in dominant oncogenes that differ from their proto-oncogenes or are over-expressed, occur by point mutations, localized reduplication, or chromosomal translocation. An oncogene therefore disrupts a cell's normal activity and can lead to uncontrolled cell division, and ultimately cancerous cells. Genes for the inhibition of cell growth are termed tumor-suppressor genes and include *APC* and *TP53*; the loss or under-expression of these genes also results in an uncontrolled cell division.^{1,2}

Therefore, by analyzing gene expression levels and the biological pathways associated with the genes involved in a cancer, one can study the difference between normal cell and cancerous cell pathways to determine the genetic origin of the faulty pathway, thereby identifying potential targets for treating cancer. The potential subtypes of that cancer can be identified through class discovery, the identification of novel cancer subtypes, and class prediction, the assignment of tumor samples to pre-defined classes in order to aid in predicting the outcome.³ Gene expression analysis also allows for biomarker and gene signature discovery. The use of gene expression profiling and development of gene biomarkers/signatures for cancer allows for the diagnosis, progression and aggressiveness analyses, prognosis, prediction of therapeutic treatment, and/or

identification of patients who would benefit from therapeutic treatment to better understand the disease and its biology.⁴

Various assay technologies have been developed for gene expression analysis (**Fig 1**). For example, qRT-PCR amplifies a gene of interest and uses fluorescent probes or dyes to depict the gene expression; NanoString nCounter uses probe pairs to anneal to a region of RNA and detect genes; DNA microarrays allow cDNA targets to hybridize to probes on a solid slide and can use fluorescence to detect genes; Illumina MiSeq RNA-Seq bridge amplifies nucleic acid samples to create clusters, which are then interpreted by the MiSeq system; and lastly, tissue microarrays (TMAs) study circular punches from tissue sample blocks with labelled probes or antibodies to determine the gene expression. Each of these assays has been used in diagnosis, prognosis, and treatment prediction for a variety of cancers. Some assays have been commercialized for clinical cancer use. For example, Afirma® is a microarray test for thyroid cancer diagnosis and the Oncotype DX qRT-PCR test is for guiding breast cancer treatment. Many assays are under investigation in clinical trials or studies.

However, numerous challenges involved with tumor sample collection, experimental design and determining the proper assay to use, analytical and

diagnostic factors (such as the interpretation of samples and biomarker performance analysis), assay detection limits and specificity, drug development for rare cancers, and clinical distribution of a significant biomarker must be overcome to develop a novel clinical assay for cancer patients. Biomarker development is also naturally affected by a patient's natural biology and history, intratumor heterogeneity, cancer progression, and germ-line mutations. This results in a prolonged period from the biomarker discovery to patenting and clinical translation stages.^{4, 5, 6, 7} Therefore, we intend to provide a guideline for choosing an assay method with respect to its oncological applications in a clinical setting.

qRT-PCR Principle and Clinical Assay

Principle

PCR (polymerase Chain Reaction) was first invented by Kary Mullis and his colleagues from the Cetus Corporation when they exploited the ability of *Taq* polymerase from *Thermus Aquaticus*, which amplifies DNA, to withstand elevated temperatures. In reverse transcription PCR (RT-PCR), RNA is first reverse transcribed into cDNA and then amplified. The first generation of RT-PCR uses a band's intensity in gel for a semi-quantification.⁸ Real-time quantitative reverse transcription PCR (real-time

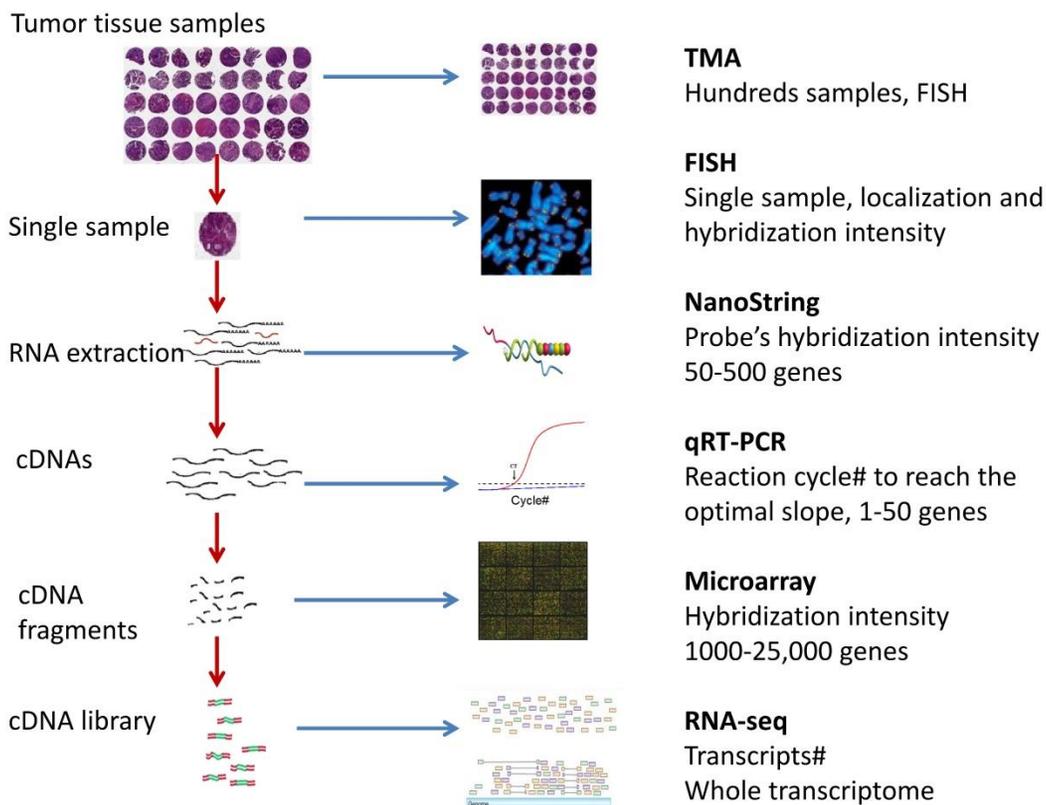


Fig 1. Overview of the gene expression detection assays for cancer tissue sample.

qRT-PCR) and digital PCR were then developed. Instead of detecting how much PCR product is generated (conventional PCR), qRT-PCR counts how many reaction cycles are used to reach an amplification slope status in real-time. The Ct (cycle threshold) levels, defined as the number of cycles required for the fluorescent signal to cross the threshold (i.e. exceeds background level) are inversely proportional to the amount of target nucleic acid in the sample (i.e. the lower the Ct level, the greater the amount of target nucleic acid in the sample). The first step requires reverse transcription of the mRNA template. Then, a primer pair anneals to the target cDNA template and a DNA polymerase (e.g. Taq) extends from the ligated primers. A thermal cycler varies the temperature settings to perform the three stages of the PCR: denaturation, annealing, and extension. Finally, a standard dissociation curve can be studied to determine the initial amount of RNA (cDNA) transcripts.^{9,10}

Primer design

Primers are required in qRT-PCR analysis to anneal to a specific portion of the mRNA so that the DNA polymerase can extend from the primer and amplify the gene of interest. The factors to consider in primer design are the melting temperatures (T_m) and GC content of the forward and reverse primers. A range of T_m s have been reported, with 58°~60°C considered as stringent. Tight annealing of the primer to the mRNA template will reduce the chances of primer-dimers or hairpins.^{11, 12} Lastly, primers spanning exon-exon junctions allow for the strict amplification of mRNA samples and not any contaminating DNA as DNA contains alternating regions of exons and introns while mRNA lacks introns.¹¹ There are a few freeware to help the primer/probe design, for example Primer3, the primer designing tool from NCBI.

Enzyme/Kit

Many commercial companies provide qRT-PCR Taq polymerase enzyme and mixture reaction solutions. Besides the digital PCR that detects the actual copy of mRNAs and other alternative PCR methods, there are two major qRT-PCR systems: TaqMan and SYBR green. In TaqMan qRT-PCR, a specific probe labeled with a fluorescent reporter is bound on the cDNA of the target RNA. Taq polymerase enzyme will degrade the probes during the chain extension such that the fluorescent reporter is released. The amount of PCR product can be quantitatively measured by the fluorescence reporter detection.¹⁰ As different reporter dyes can be utilized for a TaqMan assay, the sensitivity of a TaqMan assay

is one of its major advantages. It also has a high specificity because of the specific probe but it cannot be used for multiplex gene targets. In SYBR Green qRT-PCR, the fluorescent dye SYBR Green binds with the amplified double stranded DNA and emits an intense green light. Therefore, the PCR products can be measured by detecting the bound SYBR Green fluorescence. Compared to TaqMan qRT-PCR, this method is inexpensive, easy to use, and can be used for multiplex gene targets. However, due to its ability to bind to any double-stranded DNA, nonspecific binding can lead to over quantification of PCR product.^{9,13}

Data Analysis

The quantitation of results generally requires the study of the output dissociation curve, which allows a graph of known RNA concentration to be used as a reference for a curve constructed from the unknown RNA quantity. This approach is known as the standard curve method. The comparative Ct method involves comparing the Ct values of interest with a control, such as a non-treated or normal sample, which are both then normalized to a housekeeping gene.^{9,11} A biostatistical analysis can also be done where the final gene expression values are log-transformed for a normalized or symmetrical data distribution.

Cancer clinical

ThyraMIR™ is a commercial RT-PCR assay developed by Interpace Diagnostics Inc. It is used for the diagnosis of thyroid nodules, one of the most common cancer types, based on the evaluation of expression of 10 miRNAs.¹⁴

The well-known Oncotype DX, a gene expression profiling technique developed by Genomic Health, was the first genomic biomarker assay advising on breast cancer treatment options. Through the identification of 250 candidate genes with known functions and relevancy to hormone receptor-positive breast cancer, a 21-gene signature of 5 reference genes and 16 cancer-related genes involved in tumor proliferation, invasion, and estrogen signaling was established. It is able to predict the recurrence risk in early-stage (stages I and II), estrogen receptor (ER) positive, lymph node negative breast cancer patients to assess and predict the efficacy of adjuvant chemotherapy (ACT) in treating a patient.^{15,16}

Techniques similar to Oncotype DX have also been applied in colon cancers. For example, two separate studies established 18-gene signatures, both of 13 cancer-related genes and 5 reference genes to study stage II colon cancers.^{17,18} qRT-PCR has also been applied to monitor the development of severe

acute respiratory syndrome (SARS)-associated coronavirus and detect the prognostic markers in leukemia.¹⁹ In a study establishing a gene signature for human prostate cancer, RNA was extracted from snap-frozen human prostate surgical tissue samples and reverse transcribed using random hexamers and Superscript II Reverse Transcriptase. qRT-PCR then aided in the identification of 8 genes of interest: ornithine decarboxylase, ornithine decarboxylase antizyme, adenosylmethionine decarboxylase, spermidine/spermine N(1)-acetyltransferase, histone H3, growth arrest specific gene, glyceraldehyde 3-phosphate dehydrogenase, and Clusterin.²⁰ qRT-PCR has also been successfully applied in non-small cell lung cancer (NSCLC) studies to establish a 14-gene signature with 11 cancer-related target genes and 3 reference genes and identify 5 genes essential in the treatment outcome prediction.^{21,22}

DNA Microarray Principle and Clinical Assay

Principle

The development of DNA microarrays can be traced to the late 1990s and early 2000s where methods for fluorescent detection were adapted for the progression of array technology.²³ DNA microarrays can be used for quantification of either genomic DNA or RNA. There are two types of DNA microarray: 2-channel and 1-channel arrays. In 2-channel array, the array slides are made by spotting with cDNA fragments (custom cDNA array) or oligonucleotide probes (Agilent). The treated sample's cDNA and the normal control sample's cDNA are labeled by two types of fluorescence (2-channel). For example, cDNA from tumor samples may be labeled with Cy5 while cDNA from normal tissue samples may be labeled with Cy3. The mixture of treated and control samples are then hybridized on array. The ratio of the 2-channel intensities of each spot represents the gene expression of treated sample relative to control. Now Agilent produces commercial 2-channel arrays.^{24, 25} In 1-channel array, the oligonucleotide probes are synthesized on the surface of slide. The fluorescent labeled sample cDNAs are then hybridized on the array and the absolute intensity of hybridization signal is measured. Affymetrix GeneChip is a typical 1-channel array.^{24, 26} Illumina BeadArray is a variation of 1-channel array which synthesizes barcoded probes on a surface of microbeads and makes cells filled with beads.²⁷ A DNA microarray assay needs a commercial array or custom designed array and cDNA labelling preparation from each RNA sample. Following hybridization of the cDNA samples on array, a

scanning device images and measures the fluorescent levels of the microarray.

Arrays/kit

There are three main manufactures for commercial microarrays: Affymetrix GeneChips, Illumina BeadArrays, and Agilent 2-channel arrays.^{25,26,27} These microarrays meet all kinds of needs for human gene expression detections, such as the whole genome-wide transcriptome detection.²⁵ If a special panel of gene expression detection is needed, a custom array can be designed by these commercial companies.

Data Analysis

Normalization is required to remove the variation in an experiment that affects gene expression levels and is necessary in the earlier stages of microarray examination as the following analyses will be affected by the normalization. Following imaging of the microarray, the raw data is first analyzed by subtracting the background fluorescence from the fluorescence of each spot. Selecting an appropriate housekeeping gene or genes is a delicate task as some genes previously thought to be ideal references have been shown to vary in expression level, depending on circumstances such as a donor's biological state.²⁸ A common normalization method, such as the non-linear locally weighted scatterplot smoothing (LOWESS) method, is applied to the background-adjusted intensities.²⁹

Cancer clinical

Afirma®, a microarray assay developed by Veracyte Corporation (South San Francisco, California), is a gene expression classifier that measures the expression of 167 gene transcripts. It classifies aspirated material from thyroid nodules as either benign or suspicious.³⁰

MammaPrint is a 70-gene signature derived from 5000 expressed genes using a DNA microarray assay to predict the distant disease-free survival and overall survival in lymph-node negative patients. It has also been shown to have significant prognostic results in lymph-node positive tumors. Being cost-effective and shown to improve quality-adjusted survival, it identifies patients in the "low-risk" group with a >90% chance of being disease-free for a minimum of 5 years, allowing them to refrain from unnecessary adjuvant chemotherapy.^{31,32}

DNA microarrays have been used to determine the differences between the subtypes of lung squamous cell carcinomas (SCC) and stratify the patients for accurate prognoses and targeted research. One study used 382 SCC patients from five published cohorts of DNA microarrays to detect the subtypes

and the collection and assay of 56 SCC patients for validation. Four independent subtypes were identified and statistics and bioinformatics were used to determine subtype survival outcome, clinical covariates, and biological processes.³³

DNA microarrays have also been employed to evaluate the expression of *ZAP-70* in purified leukemic cells from chronic lymphocytic leukemia (CLL) patients. It was determined that the *ZAP-70* expression level, which is high in T-cells, can be used to identify patients requiring certain treatments by assigning patients with CLL to the correct *IgVH* mutational subtype. High correlation concordance was shown for a spread of *ZAP-70* mRNA levels between the DNA microarray and qRT-PCR assay.³⁴ The DASL™ (cDNA-mediated annealing, selection, extension, and ligation) assay to target gene-specific sequences was established using 16 FFPE samples from prostate, lung, colon, and breast tissues types.³⁵

NanoString nCounter Principle and Clinical Assay

Principle

The nCounter technology is a robust, sensitive, reproducible, and easy-to-use method. Developed by NanoString Technologies, nCounter is a variation on the DNA Microarray. It employs two ~50 base probes per mRNA that hybridize in solution. The reporter probe carries the signal, while the capture probe allows the complex to be immobilized so that the Digital Analyzer can detect the colour codes. The Digital Analyzer senses the hybridized colour-coded probes and captures the signal intensities as gene expression mRNA levels. As the probes anneal directly to an RNA sample, no reverse transcription to produce cDNA is required. This assay has a wide range of potential applications such as in the diagnostics and validation of gene expression experiments for diseases (e.g. cancer) and research of gene regulatory pathways, ultimately leading to the translation of the data for clinical use.^{36,37}

Probe design

Probe pairs are constructed for each unique gene. The probe pair is comprised of the capture and reporter probes, each 35-50 bases long and complementary to the same individual RNA molecule. The capture probe is attached via its 3' end to an affinity tag, such as biotin, while the reporter probe is detected through coupling via its 5' end to a colour-coded tag (a backbone of single-stranded DNA ligated to RNA molecules labeled with fluorophores). There are four different coloured fluorophores and six available positions on a colour code, conferring a wide range of tags that can be mixed in a single-well

reaction and successfully be resolved and identified during data analysis.^{36,37,38}

Enzyme/kit

As the probes attach directly to an RNA sample for its detection, no enzyme is required. The capture and reporter probes are mixed with RNA in solution prior to forming tripartite structures via hybridizing to their designated segments. Two kits of reagents are required: the CodeSet and the Master Kit. The CodeSet has customized or pre-designed probes, depending on one's experimental protocol, and a set of system controls. The Master Kit contains ready-to-load consumables and reagents for an assay. The Digital Analyzer identifies, scans, and counts the molecular barcodes per sample.^{36,37,38}

Data analysis

The Digital Analyzer uses a CCD camera through a microscope objective lens to magnify and image the immobilized Reporters, capturing hundreds of images per sample and outputting hundreds of thousands of counts; it processes the images and exports the data as comma separated (.csv) output files which can then be used as input for the nSolver™ Analysis software. The nSolver™ software performs quality control (QC) and data normalization and analysis.^{36,38}

Normalization is needed to account for variations in hybridization, purification, binding efficiency, and other experimental factors. NanoString recommends applying the internal positive controls in each CodeSet to remove variability. In addition to the positive control and reference (housekeeping) gene normalization, global and Plex² normalization may also be performed.³⁸

Cancer clinical

PROSIGNA is a well-known 50-gene NanoString nCounter assay for measuring the risk of relapse (ROR), a prognostic factor for relapse-free survival in breast cancer patients with node-negative tumors who have not received adjuvant systemic therapy.^{31,39}

One study used a custom designed nCounter CodeSet for 414 gene elements for studying colon cancer and discovered a subset of genes serving as potential biomarkers for cancer prognosis in FFPE tissue samples. There was a moderate correlation between the nCounter and microarray platforms. The assay included multiple published gene signatures for colon cancer prognosis and several candidate genes elements from ongoing studies in intestinal stem cell biology and epithelial-to-mesenchymal transition (EMT).⁴⁰

In another study, the NanoString nCounter technology was chosen to assay RNA extracted from

FFPE samples from stage II gastric cancer patients based on its ability to evaluate the expression levels of up to 800 genes. After screening 186 patients for the identification of prognostic genes and evaluating the strength of the prognostic algorithms, 8 genes (*LAMP5*, *CDC25B*, *CDK1*, *CLIP4*, *LTB4R2*, *MATN3*, *NOX4*, and *TFDP1*) were identified that, together, efficiently predicted a patient's outcome for the recurrence of cancer post-surgery, regardless of prior adjuvant chemotherapy. There is hope this prognostic signature will be validated in a phase III trial.⁴¹

Through the extraction and analysis of 100 ng of RNA from fresh-frozen tissue and FFPE samples, a 25-gene signature for medulloblastoma patients was established using a specific NanoString nCounter CodeSet through demonstrating a high concordance between the nCounter assay and an Affymetrix expression array data using 101 medulloblastoma samples and a second group of 130 non-overlapping medulloblastomas of known subgroup. Three laboratories in Canada, Switzerland, and the United States demonstrated reproducibility of the assay.⁴²

MiSeq RNA-Seq Principle and Clinical Assay

Principle

RNA sequencing (RNA-Seq), which directly sequences and counts the mRNA molecules in the whole transcriptome, can be selected to measure gene expression to detect genomic changes in disease states. Four steps are involved in RNA-Seq: the mRNA transcript is fragmented and primers are allowed to randomly anneal to mRNA segments; first and second strand cDNA synthesis from reverse transcription of the mRNA fragments result in double-stranded cDNA; the ends of the fragments are tagged with a phosphate group and poly(A) tail; and finally, adaptors are ligated onto the cDNA fragments so that PCR amplification and sequencing can be performed.⁴³ Sequencing involves the continual addition of fluorescently labelled nucleotide bases of four colours to the adaptor sequence, followed by photo imaging by a machine each time a base is added. These images are then translated into a sequence of bases.⁴⁴, ⁴⁵ The MiSeq® bench-top appliance sequences the cDNA templates and generates sequence reads in densities of 10 million individual molecular clusters per square centimeter through cluster generation, sequencing by synthesis (SBS), and data analysis.⁴⁶

Kit/RNA library preparation

MiSeq RNA preparation kits, such as Illumina's Tru-Seq RNA-Seq kit, can be purchased from manufacturers. The kit includes reverse transcriptase for the reverse transcription of RNA into cDNA,

primers, and adaptors. Sample preparation results in the ligation of adaptors to both ends of a nucleic acid fragment.^{46,47}

Data analysis

MiSeq conducts the basic data analysis with the MiSeq Control Software (MCS) and the Sequencing Analysis Viewer (SAV) installed on another computer. The MiSeq Reporter generates FASTQ-format files containing the sequenced reads for each sample and their quality scores. The MiSeq Analysis folder contains a BaseCalls folder where the FASTQ files are sent to. The Targeted RNA workflow outputs a BAM file that contains the reference genome, with various annotations and set chromosome sizes, and a target hit file with aligned replicate counts per transcript.^{47,48}

The fastq files exported from MiSeq can be analysed alternatively. Read alignment poses as a complication for RNA-Seq as the reads are compared to a reference genome where exon-exon splice junctions exist in RNA transcripts but not DNA transcripts. RNA-Seq alignment tools that correct for this include GSNAP, MapSplice, RUM, STAR, and TopHat.^{48, 49} MapSplice, for example, searches for exon-splice junctions in a sample mRNA transcriptome and then determines a likely alignment for each mRNA tag to a reference genome. In the first phase, the "tag alignment" phase, potential alignments of the mRNA tags to the reference genome are identified. The second phase, "splice interference phase", splice junctions appearing in one or more tags are further studied for their quality and diversity of alignments to output a splice significance score. Following read alignment, transcripts are formed from the mapped reads using computer software.⁵⁰ Programs such as Cufflinks and FluxCapacitor count the total number of reads mapping to transcripts to quantify gene expression.⁵¹

Data between each sample can be normalized by reads per kilobase per million library reads (RPKM). Normalization on expression levels per replicate is performed based on the number of aligned reads. Variance estimation requires dividing the raw variance of all sample IDs by the square of the mean. Deriving the p-value for differential expression per transcript requires a negative binomial distribution model normalized for transcript abundance. A Benjamini-Hochberg procedure is used to calculate the q-values to correct for multiple hypothesis testing and the false discovery rate (FDR). The statistical results are summarized and output to the Alignment folder.⁴⁸

Cancer clinical

Several studies have used next-generation sequencing (NGS) technologies for cancer gene analysis and/or signature development. For example, one study used NGS to analyze 21-genes, including the *BRCA1* and *BRCA2* tumor suppressor genes, associated with breast and ovarian cancers.⁵² NGS was also used to determine the instability and examine the pathogenic properties of and clinical impact resulting from the large number of somatic microdeletions at exon 19 affecting the Epidermal Growth Factor Receptor (EGFR) in NSCLCs.⁵³

By selecting 33 patients with leukemia and designing amplicons targeting *RUNX1*, *CEBPA*, *CBL*, *NRAS*, *KRAS*, *DNMT3A*, *EZH2*, and *TP53*, one study analyzed the robustness of the PCR amplification strategies and GS FLX Titanium and Illumina MiSeq amplicon-deep sequencing platforms. The aim was to evaluate the technologies for their routine clinical use in characterizing and monitoring diseases using individual patient assays. MiSeq could detect *TP53* variants and a 24-base pair insertion-deletion, conferring a predictive characteristic of the assay to detect residual disease and identify mutations for the assignment of treatments.⁵⁴

The MiSeq RNA-Seq technique was also used to assess the purpose of tumor-educated blood platelets (TEPs) in systemic and local responses in tumor growth. Using a total of 283 platelet samples (228 patients with localized and metastasized tumors and 55 non-disease individuals), mutated *KRAS* and *EGFR* in plasma DNA and platelet RNA was detected. Blood-based onco-signatures can be derived from TEP mRNA profiles to examine tissue biomarkers for the stratification of patients into low- and high-risk groups and the selection of their therapies. Although the clinical relevance of blood platelets in the molecular diagnosis of patients presenting with several types of cancer was established, systemic factors such as chronic disease or transient disease states and/or non-cancerous diseases could have affected the TEP mRNA profile, prompting their further validation in blood-based therapy selection, longitudinal studies, and disease recurrence monitoring.⁵⁵

Fluorescence in situ hybridization (FISH)

Principle

FISH was developed in the early 1980s.⁵⁶ It applies fluorescent-labeled short DNA probes that hybridize to target DNA or RNA sequences *in situ* and fluorescence microscopy to localize and detect the targets in tissue slides. FISH can determine the presence or absence of mRNA expression from a gene of interest, as well as localize these gene expressions

in specific cells.^{57,58}

Pre-treatment is required to preserve RNA and tissue morphology. Cells, FFPE, or frozen tissue sections are fixed, then permeabilized to allow target accessibility. A target-specific probe hybridizes to the target RNA(s). Separate but compatible signal amplification systems enable the multiplex assay. Signal amplification is achieved via series of sequential hybridization steps. Multicolor FISH can be used to identify as many labeled features as there are different fluorophores used in the hybridization.^{57,58}

Probe design

Many different types of probes are used for in FISH, including cDNA, crRNA, and synthetic oligonucleotide probes. Probe size is important because longer probes hybridize more specifically than shorter probes, so that strands of DNA or RNA (often 20–50 nucleotides), which are complementary to a given target sequence, are often used to locate a target.^{57,58,59}

Data analysis

Automated fluorescence signal analysis system can analyze fluorescence signal patterns / FISH spots in cells or cell nuclei automatically, precisely, and reproducibly. Signal channels can be combined to an assay, and the images obtained in each channel can be processed individually. Morphology of target cells, nuclei, or other objects can be precisely defined with a number of cell selection parameters. Multiple features of objects (e.g. area, shape, intensity, and signal distribution) can be measured.⁵⁷

Cancer clinical

The application of FISH is growing rapidly in genomics, cytogenetics, prenatal research, tumor biology, radiation labels, gene mapping, gene amplification, and basic biomedical research, as well as clinical applications, for diagnostic purposes. As a gene expression assay, FISH is used for localization and detection of specific RNA targets (mRNA, lncRNA and miRNA) in tumor cells and tissues.

In one study, FISH was used to assess *HER2* transcript distribution in a fresh frozen *HER2*-positive human breast cancer tissue section and *KRAS* in wild-type and mutant cells. Padlock probes, oligonucleotides that are circularized following hybridization to a target sequence that is complementary to the probe ends, were designed to detect the cDNA molecules. Rolling circle amplification can then be used to amplify the circularized padlock probes. *HER2* expression varied, as expected, and a somatic point mutation was identified in *KRAS*.⁶⁰

23 small molecule inhibitors and antibodies that target *MET*, a proto-oncogene, and its ligand, hepatocyte growth factor (HGF), have been developed and tested in over 240 clinical trials in 2014.⁶¹ One study analyzed *MET* expression and activity in a matched set of FFPE vs. fresh frozen tumor samples consisting of 20 cases of gastric cancer using both FISH and IHC. It was found that RNA FISH can be used to confirm findings obtained by IHC and potentially may replace IHC for certain targets if no suitable antibodies are available. RNA FISH is a valid platform for testing predictive biomarkers for patient selection.⁶²

MicroRNAs (miRNAs) are excellent tumor biomarkers because of their cell-type specificity and abundance. A reliable multicolor miRNA FISH technique was developed that enables miRNA visualization in FFPE tissues. They tested two skin tumors, basal cell carcinoma (BCC) and Merkel cell carcinoma (MCC), to visualize differentially expressed miRNAs in FFPE tumor samples.⁶³

qRT-PCR analysis and FISH was applied in a study to assess miRNA expression in the HER2-positive breast cancer cells. It was found that miR-489 was expressed at a significantly lower level in tumor tissues compared to the adjacent normal tissues and directly targets HER2.⁶⁴ FISH can also be used for lncRNA expression detection in cancer. For example, one study used RNA FISH to investigate the association between lncRNA AK023391 expression and the clinicopathological characteristics and prognosis of gastric cancer patients.⁶⁵ A separate study demonstrated two antisense lncRNAs at the 5' (5'aHIF-1 α) and 3' (3'aHIF-1 α) ends of the human *HIF-1 α* gene in kidney tumor tissues using RNA FISH.⁶⁶

Tissue Microarray Principle and Clinical Assay

Principle

Developed in 1998 by Kononen and associates, tissue microarrays (TMAs) require combining small tissue samples into "sausage blocks" to study the connection between the tissue samples on the slide with the clinical data.⁶⁷ By using *in situ* analyses, such as fluorescent *in situ* hybridization (FISH), RNA *in situ* hybridization (ISH), or immunohistochemistry techniques, TMAs can detect protein staining and cellular locations, offering its application in molecular diagnostic studies. However, the number of proteins to be detected is limited by the antibodies.^{68,69} TMAs are expanded from gene expression serial analysis and cDNA microarrays so that they only require a single experiment to study the gene expression from many genes in a single tumor sample, allowing multiple patient samples representing various stages

of a disease to be simultaneously investigated.⁷⁰ With the use of a TMA slide, probes for hybridization, and a signal detection instrument or scanner for data, followed by data analysis, TMAs offer the advantage of permitting a single set of tissues with associated clinical data to be used in various and unlimited studies. The steps generally required to prepare a TMA are as follows: preparation of a high-adhesive glass slide; acquisition of donor tissues; tissue core mapping and arrangement; sampling, sectioning, and transferring of the tissue donor cores; and tissue staining and molecular analysis. First, samples are taken from FFPE tissues. After mapping a tissue section stained with hematoxylin and eosin (H and E), a tissue arrayer removes cores from the donor block and places them in the recipient block.

Primer/probe design

FISH requires a DNA probe typically from cloned sources (e.g. plasmids) and target DNA in the form of chromosomes or interphase chromatin that will bind to the probe. The DNA probe can be labelled with haptens, fluorochrome, or dye-based nucleotides. The DNA probe and target DNA are permitted to hybridize to each other only following their denaturation into single strands. Fluorescent microscopy can allow for the visualization of the DNA probe-target DNA complexes.⁷¹ Similar to FISH, RNA ISH requires a gene-specific nucleotide probe to bind to a certain sequence on a DNA or RNA strand. The probes may be labelled with radio-, antigen-, or fluorescent-labelled bases and analyzed by autoradiography, immunohistochemistry, or fluorescent microscopy, respectively.⁵⁸

Data analysis

Minute tissue samples (typically 0.6 mm in diameter) from up to 1000 different tissues can be analyzed on one microscope glass slide. Since all tissue samples are simultaneously analyzed with the same set of reagents, standardization, speed of processing, and cost are all efficient so that normalization to account for technical experimental variances between samples need not be extensively performed.^{67,68,70}

The tissue sections can first be analyzed via H and E staining or immunofluorescent ISH. An automated scanning system can be purchased to scan and analyze the prepared TMA slides. As the tests carried out on a TMA may vary depending on one's experimental design and purpose, a different combination of gene detection and analysis may be performed. Autoradiography or immunohistochemistry may be applied for detection followed by fluorescent microscopy or imaging by an array

scanner may be used for the assessment of fluorescent levels in a TMA.⁶⁹ For example, one can use digital imaging scanning and processing technologies and Microsoft Access and Excel as databases, then compare their results to clinical data.⁶⁸

As various techniques exist to visualize TMAs, numerous data analysis and normalization methods have been developed. The Tissue Array Management and Evaluation Environment (TAMEE) web-based database application that stores experimental parameters, TMA images, and evaluation results is available for TMA data analysis and management for the whole TMA workflow.⁷² The Tissue Array Co-Occurrence Matrix Analysis (TACOMA) uses local inter-pixel relationships to quantify cellular phenotypes to recognize specific staining patterns and output a score of categorization.⁷³

Cancer clinical

Through studying candidate genes and the RNA, DNA, and/or protein molecular alterations in each sample, TMAs can be used in the research of diseases, such as cancer, and for drug target discovery, to determine the therapeutic importance of these genes in a patient, as well as their diagnostic and prognostic significance. In cancers, this allows for the examination of tumor progression and the identification and validation of prognostic factors or genes that can potentially be translated into the clinical setting for therapeutic treatments. For example, TMAs were used in an original experiment to study the amplifications of six genes, as well as the expression of *p53* and estrogen receptor (ER), in breast cancers, allowing for the further stratification of the cancers into subgroups.⁶⁷ To examine how the molecular changes in bladder cancer patients affect their clinical outcomes, a tissue microarray was constructed using 2317 samples from 1842 patients to study the outcome of the amplification of *CCNE* and its protein expression.⁷⁴

Four independent TMAs were used to analyze the prognostic markers (ER, PR, and *p53*) in impacting molecular changes and clinical endpoints in 553 breast carcinomas. 0.6 mm samples were taken from one central and three peripheral regions from FFPE samples. Multiple punches per tumor were taken that demonstrated the tumors could be distinguished into three subgroups (positive, negative, and heterogeneous). 12 TMAs constructed using three antibodies on four arrays demonstrated significant associations with tumor-specific survival than large section analyses. A single sample per tumor was sufficient to identify associations between molecular alterations and clinical outcome. Further validation of the established molecular markers is required using

larger tissue samples from clinical trials.⁷⁵

As many previous studies assessing ER expression using immunohistochemistry have not been reproduced, one study used TMAs to examine the variability in ER expression reporting for breast cancers. 5 independent laboratories constructed TMAs by coring an invasive breast cancer donor block from 29 patients twice. A moderate to high interlaboratory agreement was determined, demonstrating that TMAs are efficient for the identification of ER variability reporting and have the potential to be applied to similar projects.⁷⁶

TMAs of 544 clinical samples from varying stages of prostate cancer were used to study the expression of candidate genes discovered by cDNA microarrays of the CWR22 xenograft model. Using 60 prostatic intraepithelial neoplasia, 264 primary tumors, 41 distal metastases, and 134 hormone refractory specimens, with 45 samples of benign prostatic hyperplasia acting as the control, they demonstrated the use of *S100P*, *CRYM*, and *LMO4* in affecting androgen-independent growth and therapy failure in prostate cancers. Following mRNA *in situ* hybridization and immunohistochemical analyses of the TMAs, strong correlation was found between the cDNA microarrays and mRNA. Over-expression in hormone-refractory CWR22R xenografts of *S100P* was seen while *CRYM* and *LMO4* were down-regulated.⁷⁷

Another study used TMAs from 1023 samples of prostate tissues to examine the significance of the Polycomb group protein EZH2 in the progression of prostate cancer. Included in the cohort were 400 samples from 23 individuals who passed away from hormone refractory metastatic prostate cancer. Highly expressed EZH2 is mainly seen in the nucleus and its staining increases across the benign, prostatic atrophy, prostatic intraepithelial neoplasia, and clinically localized prostate cancer subgroups. Metastatic prostate cancer demonstrated the highest intensity staining. The group also analyzed the validity of EZH2 as a tissue biomarker for prostate cancers, considering the clinical and pathological factors, using TMAs from patients with provided clinical follow-ups. It was demonstrated that EZH2 is a potential biomarker for prostate cancer as its concentrations can affect the disease aggressiveness and abnormal EZH2 expression may progress the cancer.⁷⁸

Comparison

We compared the qRT-PCR, DNA Microarray, NanoString nCounter, Illumina MiSeq RNA-Seq, FISH, and Tissue Microarray assays in primer/probe design, sample preparation, instrument used, data analysis, reproducibility, specificity, sensitivity,

throughput, complexity, processing, data analysis, use in research, and commercialization (Table 1), and the cost and time used (Table 2).

Sample Preparation

Cell lysates, fresh frozen (FF) tissue, and FFPE samples are widely used in cancer genomics studies by the qRT-PCR, DNA microarray, NanoString nCounter, and Illumina MiSeq RNA-Seq assays to provide data for patient diagnosis and prognosis. FISH and TMAs also employ FFPE specimens. FF samples allow for better preserved RNA but are limited in availability and use as tissue banks and research groups mainly collect and study them; they also have the disadvantage of sample size, transport, and clinical annotation.^{79,80} The RNA isolation and purification steps are generally the same for the qRT-PCR, DNA microarray, NanoString nCounter, and Illumina MiSeq RNA-Seq methods as commercial kits may be purchased for RNA extraction from cell lysate, FFPE, and FF samples. The TMA has a longer process for sample preparation using FFPE samples.

The purpose of the formalin fixation and paraffin embedding (FFPE) protocol is to preserve proteins for

tests such as FISH or immunohistochemistry, rather than nucleic acid strands. As TMAs do not require RNA extraction and apply fluorescent detection tests such as FISH and ISH, they can make use of the long-term storage and abundance of samples acquired from FFPE samples. Although FF samples provide better quality samples for microarray analyses, microarray oligonucleotide probes have consistency in detections from both FFPE and FF specimens. FFPE samples are also commonly used in microarray gene expression analyses. Due to differing tissue characteristics between FFPE and FF samples, the yield of cDNA after reverse transcription and amplification from FFPE and FF samples may vary. Through matched FF and FFPE samples studies to evaluate the integrity and consistency of gene expression in FFPE samples, it has been shown that FFPE samples may be used in gene expression analysis.^{80,81} Similarly, FF samples are preferred for RNA-Seq experiments as they provide better quality data.

Table 1. Comparison of qRT-PCR, DNA Microarray, NanoString nCounter, Illumina MiSeq RNA-Seq, and Tissue Microarray assay properties.

Assay	qRT-PCR	DNA Microarray	NanoString nCounter	Illumina MiSeq RNA-Seq	Tissue microarray & FISH
Primer/probe design	Gene-specific primer with attached quencher and reporter fluorophores; SYBR green	DNA oligo probes complementary to cDNA samples	Capture probe with 3' affinity tag and Reporter probe with colour-coded tag	Primers on flow cell and adaptors to ligate to ends of sample	Gene-specific RNA probes; gene-specific fluorochrome-labelled probes; monoclonal antibodies
Sample preparation	RNA extraction; reverse transcribe sample	RNA extraction; reverse transcribe sample, fragmentation	RNA extraction	RNA extraction; reverse transcribe sample; fragmentation, library construction	Map donor block; place into recipient block; make TMA
Instrument	Thermal cycler	Microarray scanner	Prep Station and Digital Analyzer	MiSeq benchtop sequencer	Tissue arrayer; microscope or array scanner
Reproducible	Yes	Yes	Yes	Yes	Yes
Specificity	Forward and reverse primer design, oligonucleotide probe	Density of probes annealed to the slide, probe design	Design of Capture and Reporter probes	Rely on data analysis	Rely on probes to be used
Sensitivity	10-200 copies/cell	1-10 copies/cell	<1 copy/cell	<1 copy/cell	1-10 copies/cell
Clinic study	Yes	Yes	Yes	Yes	Yes
Commercialized for clinical use	Oncotype DX	MammaPrint	Prosigna	No	No
Number of genes or transcripts detected	1-100	50 000	800	Whole transcriptome	3
Up to sample# per assay	1-96	1-12/array	12	96	1000
Processing steps	Prep reaction mixture, PCR cycles, Result analysis	Label cDNAs, hybridization to array, Data analysis	Label probes, hybridization to array, Data analysis	cDNA lib prep, sequencing, data analysis	Make TMA, Slide TMA Staining, Analysis
Raw Data analysis	by machine in 30 minutes	by machine in 1 hour 40 minutes	by machine in 2.7 hours	by machine in 3 hours	by machine or microscopy in 6 minutes
Normalization	3-5 housekeeping genes	Housekeeping genes; RMA; LOWESS method	Housekeeping genes; positive controls	RPKM	Tissue array co-occurrence matrix analysis
Data analysis	Absolute and relative quantification; standard dissociation curve; statistical tests	Visualization; statistical tests	Colour-coded images are taken and output as code counts	Data output as sequenced reads with quality scores or read alignments	PCR; H and E staining; FISH, ISH; fluorescent microscopy

Table 2. Cost and time of qRT-PCR, DNA Microarray, NanoString nCounter, Illumina MiSeq RNA-Seq, and Tissue Microarray assay. The prices could vary from different facilities.

Assay	qRT-PCR	DNA Microarray	NanoString nCounter	Illumina MiSeq RNA-Seq	Tissue microarray & FISH
<i>Cost on Sample preparation</i>	\$0.56 (SYBR) \$0.82 (probe)/Singleplex reaction	~\$50/array	\$20	~\$200-300	\$75-2000/array, depending on cancer type
<i>Cost on Kit or plate</i>	\$1416 (SYBR) \$1834 (probe)/100 preps	~\$350/plate	\$280-350/1 cartridge for 12 samples (Master Kit); \$3600-4800/12 assays (Custom CodeSet)	\$1200-2320/~12 reactions	\$515 for FISH
<i>Cost on Processing</i>	\$0.50/sample	\$0.025/data point; ~\$100/array	\$4.17/data point; \$35-41.67/sample	\$90/sample	\$12.50/sample
<i>Cost on Data analysis</i>	\$55	~\$100	\$65-250	\$160-175	\$95/slide using 3 antibodies
<i>Cost on Instrument</i>	\$25 000- \$95 000 Or ~\$25/run	\$50 000- \$110 000	\$235 000-\$285 000	\$128 000	\$55 000- \$98 000
<i>Time on Sample preparation</i>	1 hour	20 minutes	5 minutes	8 hours	30 minutes
<i>Time on Sample processing</i>	2-4 hours	60 minutes to 17-18 hours or overnight	5 minutes + 2.5 hours	24 hours	24 hours
<i>Time on Data Analysis</i>	30 minutes	1 hour 40 minutes	5 minutes + 2.7 hours	3 hours	6 minutes

Studies have shown that the FFPE/FF pairs demonstrate a high correlation when analyzing gene expression if the storage time of specimens is less than two years, meaning that reliable RNA can be extracted from FFPE samples in younger tissue blocks.⁸¹ Since it only requires the Capture and Reporter probes to anneal to the template, with the Reporter probe depicting the gene of interest, the sample is prevented from accumulating errors through the reverse transcription step, allowing FFPE samples to be utilized by NanoString nCounter. It has been shown that an nCounter experiment has a high concordance (90%) rate with immunohistochemistry, demonstrating that it is reproducible. Although nCounter shows promise in using FFPE samples, it is limited to detecting 770 genes at once. This limitation is present no matter which sample type is chosen.⁸¹

Due to source variation and intratumor heterogeneity, individual FFPE tissues can have varying RNA qualities. Therefore, to produce the best results from FFPE tissues, a standardized method of RNA extraction must be established. In a study comparing the Trizol, Qiagen RNeasy FFPE, and Arcturus PicoPure RNA Isolation kits for quantity and quality of RNA for microarray gene expression studies, an efficient method for RNA extraction, from the deparaffinization to microarray hybridization steps, was developed. The protocol includes an RNeasy FFPE kit with a modified deparaffinization protocol for RNA isolation from FFPE tissues and a combination of X3P Array Ovation and FFPE WTA System kit for amplification and hybridization. This workflow improves on FFPE tissues as a reliable alternative for FF samples.⁸⁰

Reproducibility

Although qRT-PCR tests have shown consistent results across replicate assays, they are not 100% reproducible as errors may be amplified along with the exponential amplification of the original nucleotide strand and the PCR can result in some loss of information during amplification.⁸² Expression microarrays have also been shown to be highly reproducible, depending on the ability of a probe to anneal to the same number of transcripts across replicate experiments. The protocols used for RNA extraction and sample purification, data acquisition and normalization, type and construction of probes, and the methods of labelling and hybridization used greatly affect the reproducibility of DNA microarrays.^{83, 84} Numerous studies have confirmed the reproducibility of the NanoString nCounter platform due to its requirement of little input RNA and lack of a reverse transcription step. For example, its reproducibility has been demonstrated using diffuse B-cell lymphoma (DLBCL), gastric, pancreas, xenograft, lung, breast, blood, and melanoma samples, while a separate study demonstrated inter-laboratory (Canada, Switzerland, and the United States) reproducibility of nCounter with medulloblastoma samples.^{42, 85, 86} Similarly, studies have used techniques such as PCR, qRT-PCR, and microarrays to show the MiSeq platform and RNA-Seq to be reproducible.^{87, 88, 89, 90} For example, the SEQC/MAQC-III Consortium have demonstrated RNA-Seq to have a >80% validation with quantitative PCR (qPCR) when sequencing unannotated exon-exon junctions, yet microarrays do not provide

reliable reproducibility data.⁸⁹ One of the greatest drawbacks of TMAs is the tumor inter-heterogeneity – the fact that a single spot punched from a tumor block is not representative of the rest of the tumor. Regardless, TMAs have a high degree of reproducibility due to the maximum 1000 spots on a single slide simultaneously being sampled and subjected to an experimental test using the same reagents. Multiple punches can be sampled from one tumor block, allowing for the same tumor block to be used in multiple experiments; for example, TMAs from one recipient block can be distributed between labs. Immunohistochemical and FISH tests can then be standardized for reproducible results.⁹¹

Sensitivity

Sensitivity refers to the minimum amount of substance, such as RNA, detected per sample in an experimental assay. qRT-PCR and PCR have a reported sensitivity of 10 copies mRNA per cell, with as many as 200 copies per cell having been detected for PCR.^{92,93} As little as 3 copies per cell have been detected in PCR, implying that this number could be similar for qRT-PCR.⁹⁴ Similarly, DNA microarrays can detect from 1-10 copies of mRNA per cell.⁸³ Both nCounter and MiSeq have the ability to sense <1 copy of mRNA per cell; nCounter utilizes region-specific colour-coded probes to bind to a single transcript, whereas MiSeq can generate a cluster of 1000 copies from a single transcript.^{46,95} Lastly, sensitivity for TMAs can be analyzed in a number of ways as it may depend on the size of the tissue spot in the array or the sensitivity of the tests (e.g. FISH, ISH) being used. For example, TMAs can array up to 1000 different sample spots on a single array, allowing 0.6 mm as the smallest sample size that can be taken with efficient detection results; *in situ* hybridization is shown to have a sensitivity of 10-20 mRNA copies per cell.^{67,68,96}

Specificity

The specificity for each of the qRT-PCR, DNA microarray, NanoString nCounter, MiSeq RNA-Seq, and TMA depends on the primers/probes and/or reagents being used. As previously discussed, a strict design is required to ensure the qRT-PCR primer tightly anneals to the mRNA transcript and amplifies only the section of interest. Therefore, the specificity of qRT-PCR depends on the forward and reverse primer design, as well as the oligonucleotide probe used in detection.^{11,12,97} The specificity of DNA microarrays depends on a number of factors, such as the density of probes annealed to the slide, the single probe (hybridization of a probe to a single target), single spot (a spot of multiple probes may contain probes that are perfectly or partially hybridized to a

target), and spot-set (multiple spots represent different sections of the same sequence).⁹⁸ Similar to PCR, the specificity may also depend on the probe design; for example, cDNA microarrays can employ probes up to a few thousand base pairs in length while oligonucleotide arrays perform best with shorter probes that can be 25-30mer or 60-70mer in length.⁸³ The specificity of NanoString nCounter is due to its target-specific Capture and Reporter probes, as these are designed to anneal to a nucleic acid region of interest. These combine with the internal controls used to form the CodeSet, which confers the overall specificity.⁹⁹ TMA quality is dependent on the type of analysis being done. For example, FISH, RNA FISH, and immunohistochemistry each confer different specificities to the TMA assay.^{100,101} RNA probes about 250-1500 nucleotides in length, with a common recommendation of 800 nucleotides, tend to demonstrate high specificity.¹⁰² Chemically-synthesized oligonucleotides labelled with fluorophores and calibrated to a certain region tend to generate RNA FISH probes with high specificity.¹⁰⁰

High-throughput

High throughput refers to the advantage of a technology to simultaneously analyze many (i.e. hundreds of thousands) samples under a certain set of conditions. It therefore may depend on the number of genes that can be detected by a technique and/or the number of samples that can be processed. Well-plates that can hold a maximum of 384 samples can be purchased for PCR. Although control samples (tests priming the housekeeping genes) and all replicates (a recommended number of three) should be run on a single plate, a qRT-PCR assay has the potential to assess 384 samples at once. This substantial number of wells also allow for the analyses of a few hundred genes in Singleplex reactions.^{12,103} The number of samples analyzed on a DNA microarray can depend on the purpose of the microarray and the number of genes being assessed. Whole-genome arrays can be performed for an individual or a small number of individuals can be simultaneously analyzed for a certain set of genes on one array. For example, Prosigna is able to assess up to 8 breast cancer patient samples for 1900 features on a single array.^{104,105} It has been reported that up to 50 000 different probes are available for detections on a microarray, with each detection representing a different gene.¹⁰⁶ The nCounter has the capacity to input 12 samples and detect 800 genes.¹⁰⁷ Approximately 1000 genes can be targeted and 96 samples can be processed using the MiSeq benchtop sequencer; clusters of about 1000 molecules or 1536 amplicons can be generated from a single transcript and anywhere from 700 to thousands

of targets have been reported.^{108, 109} Similar to DNA microarrays, the number of samples assessed and genes detected on the TMA depends on its purpose. Generally, 1000 spots representing different samples can be arrayed on a TMA.^{67,68} Although many genes have the ability to be detected as different coloured probes can be designed to anneal to their target region, studies tend to depict 1-3 genes using fluorescent detection on a TMA.^{74,75,76,77} This may be because resolution can be difficult using many different probes in an *in situ* detection assay.

Cost per assay

A qRT-PCR reaction per sample can be considered cheap in cost. Various kits can be obtained according to one's experimental design to reverse transcribe the mRNA to cDNA and carry out the remainder qRT-PCR steps for about \$300-\$550. According to BioSearch Technologies, after all reagent costs have been accounted for, a single qRT-PCR reaction totals under \$1.00. When probes are being used as the detection method, the total cost is about \$0.82 per reaction; when SYBR is being used, the total cost is about \$0.56 per reaction. These increase to about \$0.89 and \$1.13 if probes and SYBR are being used, respectively, when additional targets are present.¹¹⁰ A sample processing of \$0.50 has been reported. A qRT-PCR reaction per sample has been reported to cost as little as \$0.50 per reaction.¹¹¹ A whole microarray slide can cost between \$150-\$400, with a full genome array being \$500, and a processing cost of about \$325 per array. Some studies have reported a cost of \$0.025 per data point (or gene being analyzed).^{111,112} The NanoString nCounter provides a Master Kit and CodeSet to be purchased separately, with all necessary consumables and reagents. The Master Kit costs about \$280-350 per 1 cartridge for 12 samples.¹¹³ Custom or pre-designed CodeSets are available for specific gene detection. Pre-designed CodeSets can be purchased for \$3600-\$4800 per 12 assays for analyzing samples of human stem cells, human cancer reference, or leukemia, to name a few. A custom CodeSet for 48 assays assessing 25 genes can cost about \$6000, resulting in a cost of \$125 per assay and \$5 per data point. Doubling the target to 50 genes nearly doubles the cost to \$10 000 for 48 assays, \$208.33 per assay, and \$4.17 per data point.¹¹⁴ Several institutes provide services to carry out an nCounter experiment for up to \$530 (University of Kentucky and the Whitehead Institute) per cartridge of 12.^{112,113} A sample processing for targeted gene sequencing costs \$90 on the MiSeq Benchtop sequencer, with a 50-500 base pair sequencing kit valued at \$1400-\$2000.^{115, 116} Some institutes provide data analysis for \$160-\$175.^{116, 117} Prepared tissue

microarrays may cost as much as \$6000 (stage I breast cancer tissue array).¹¹⁸ Studies have reported the cost to build and analyze a TMA to be \$255 (18 cases) and \$12 240 (48 cases), resulting to \$12.50 per case.¹¹⁹ In addition, the instruments are valued at \$25 000-\$95 000 (thermal cycler for PCR), \$50 000-\$110 000 (microarray scanner), \$235 000-\$285 000 (NanoString Prep Station and Digital Analyzer), \$128 000 (MiSeq Benchtop sequencer), and \$55 000-\$98 000 (tissue arrayers), respectively.^{9,120,121,122,123,124,125}

Complexity and Time

qRT-PCR, DNA microarrays, NanoString nCounter, and Illumina MiSeq all require the isolation and purification of RNA. Following RNA extraction, qRT-PCR requires the mixing of various reagents before allowing the sample to reverse transcribe. A well-plate must be carefully prepared prior to subjecting the sample for the PCR and dissociation curve analysis steps in the thermal cycler machine and by the computer, respectively; this can take about 5 hours when including an RNA extraction step. Similarly, DNA microarrays require a reverse transcription step after the RNA extraction. Time is then required to allow the sample to hybridize to the array; the arrays can be pre-ordered and do not have to be constructed in the laboratory. The microarray scanner analyzes and slides and computer software can be purchased for data analysis, resulting in a total time of 20 hours.¹²⁶ NanoString nCounter only requires the user to prepare the initial sample and then transfer the sample to and from the Prep Station and Digital Analyzer machines. The machines carry out any processing and data analysis that needs to be done, resulting in a total of about 5.5 hours.¹⁰⁷ Similar to the nCounter, Illumina MiSeq only requires the user to prepare the library and load it onto the reagent cartridge; the rest of the sample processing and data analysis is carried out by the MiSeq benchtop sequencer, with the MiSeq Reporter analysis software launching following completion of the trial. A total of about 40 hours is required.¹¹⁵ TMA construction is a tedious procedure and therefore requires the most work, demanding an average of up to two days, depending on the exact procedure followed. If donor tissue blocks are not available, one must collect samples prior to preparing the block. Then, slides of the tissue sample must be stained so that one can use the tissue arrayer to accurately remove cores from the donor block and place them on the slide. Following construction of the slide, reagents for FISH, ISH, or immunohistochemistry tests must be prepared and applied to the slide. The slides can then be analyzed by eye or with a (digital) slide scanner.¹²⁷

Clinical use/ Commercialized

As discussed earlier, each of the five assays have been use in clinical-related studies to analyze the gene expression from cancer patient samples. The qRT-PCR, DNA microarray, and NanoString nCounter assays have commercialized products (Oncotype DX, MammaPrint, and PROSIGNA, respectively) for testing clinical samples. Illumina MiSeq does not have such an assay, yet one can send samples to an institute for processing.^{116,117} Tissue microarrays also do not have a commercialized assay as they are used to detect up to a few genes in a maximum of 1000 samples, requiring a cohort of patient samples to be gathered to construct a TMA. They are therefore ideal for validating studies using qRT-PCR, DNA microarrays, et cetera, and not for the assessment of individual or a small number of samples.

Table 3. Main advantages and disadvantages of gene expression detection assays.

	Advantages	Disadvantages
qRT-PCR	a popular technology, low cost	only for small number of genes
Microarray	works for more than 1000 genes, commercial chip/arrays ready to use	requires expertise for data normalization and analysis
NanoString	standard workflow and analysis pipeline are setup by the company	machine is costly, constrained by one company
RNA-seq	whole transcriptome, MiSeq pipeline for data analysis	machine is costly, requires expertise for lib prep and sequencing
FISH	localizes and detects gene expression	complicated procedure
TMA	detects hundreds of samples simultaneously	mainly for research, not for clinical settings

Table 4. General guideline for gene expression assay for cancer clinical use

Assay	qRT-PCR	DNA microarray	nCounter	RNAseq	TMA&FISH
gene#					
1-50	++				+
50-500			++		
500-2000		++			
>2000				++	
purposes					
proteins					++
mRNAs	+	+	+	+	+
novel discovery				++	
facility					
clinic lab	++				
hospital lab	++				
Institution core		+	+	++	
national lab			+	++	++
use frequency					
daily	++				
weekly		+	+		
monthly			++	++	+

Conclusion

We summarized the main advantages and disadvantages of each method as **Table 3** and setup a matrix (**Table 4**) to guide in choosing the most practical assay based on the gene number to be tested, purposes of the assay, location of experiment to be performed, and frequency of the assay.

With a sample processing fee of as little as \$0.50 for a single gene, an experimental time of about 5 hours that includes RNA isolation and purification, as well as PCR and data analysis, qRT-PCR offers the cheapest and quickest assay for processing an individual clinical sample for 1-50 genes. Small laboratories such as a hospital lab or a clinic lab suited to a clinical processing environment can purchase a thermal cycler for as little as \$25 000 to process and analyze their sample.

Other techniques employ machines that can cost up to \$285 000, which are better suited for larger research institutes and feasible if clinical samples are sent to these institutes or a national lab to be processed. With the cost constraint, the nCounter is suited for detection of 50-500 genes and microarray is good for more than 500 genes or genome-wide gene expression profiling. RNA-Seq can detect a large number of genes with the additional benefit of gene discovery, which is suited for cancer research institutions. TMAs are appropriate for protein level detection in thousands of samples at one time, which is ideal for tumor archive centers at provincial or national facilities.

In addition to the above proposed guidelines, developing an assay into clinical use must pass stringent Clinical Laboratory Improvement Amendments (CLIA) standards and a College of American Pathology certification. Diagnostic tests can result in large costs due to demonstrating improved patient mortality outcomes through replicated and randomized clinical trials.⁵ The cost of an assay, then, is an important parameter in experimental design and should be minimized for the desire to translate results into clinical use. Medical costs are increasing in the current healthcare system as cures are being sought for all illnesses and the general population ages and increases in life expectancy, resulting in more money being put towards the elderly population's care and medical costs. Cost-effectiveness studies may be performed on an assay to determine its economic utility for a consumer's and society's well-being.^{6,128} New biomarkers may undergo cost-utility studies measured in sum of money per quality adjusted life year (QALY) gained, where the ratio of the cost of a test to its benefits in the number of years gained is taken. \$50 000 USD per QALY gained is considered the upper-limit for cost-effectiveness.¹²⁶ In addition to

the necessity of developing an assay with low cost, the FDA guidelines for approval must be met. Once FDA-approved, it is then up to clinicians to accept and employ the assay.⁶

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Competing Interests

The authors have declared that no competing interest exists.

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